

---

VOL 83 (2)

# ACTA BOTANICA CROATICA



ACTA BOT. CROAT., 87–180  
ZAGREB, 2024

# ACTA BOTANICA CROATICA

Acta Bot. Croat. 83 (2), 2024

**Publisher:** Department of Biology, Faculty of Science, University of Zagreb, Croatia

## Editors-in-Chief:

Nenad Jasprica, Institute for Marine and Coastal Research, Dubrovnik, Croatia

Mirta Tkalec, Department of Biology, Faculty of Science, University of Zagreb, Croatia

## Subject Editors:

Biljana Balen, Zagreb, Croatia (plant micropropagation and plant stress physiology)

Sandro Bogdanović, Zagreb, Croatia (systematic botany, biogeography)

Snježana Bolarić, Zagreb, Croatia (plant genetics)

Sunčica Bosak, Zagreb, Croatia (algal taxonomy)

Mirna Ćurković Perica, Zagreb, Croatia (plant microbe interactions)

Sven Jelaska, Zagreb, Croatia (ecology, diversity, modelling)

Marin Ježić, Zagreb, Croatia (plant-pathogen interactions)

Mitja Kaligarič, Maribor, Slovenia (vegetation)

Dario Kremer, Zagreb, Croatia (systematic botany, flora)

Dunja Leljak-Levanić, Zagreb, Croatia (plant developmental and molecular biology)

Zrinka Ljubešić, Zagreb, Croatia (phytoplankton ecology)

Mirjana Pavlica, Zagreb, Croatia (genetic toxicology, ecotoxicology)

Dragica Purger, Pécs, Hungary (systematics, biogeography, vegetation)

Sandra Radić Brkanac, Zagreb, Croatia (plant stress physiology)

Ivana Rešetnik, Zagreb, Croatia (genetic diversity and evolution)

Marko Saboljević, Belgrade, Serbia (ecology and taxonomy of bryophytes)

Boštjan Surina, Koper, Slovenia (plant evolution, systematics, biogeography, vegetation)

Željko Škvorc, Zagreb, Croatia (forest vegetation and biostatistics)

Massimo Terzi, Bari, Italy (vegetation ecology and phytosociology)

## Editorial advisory board:

Éva Ács, Göd

Nikola Ljubešić, Zagreb

Andraž Čarni, Ljubljana

Montserrat Martinez-Ortega, Salamanca

Daniel Hagège, Orleans

Helmut Mayrhofer, Graz

József Horváth, Keszthely

Božena Mitić, Zagreb

Ljudevit Ilijanić, Zagreb

Toni Nikolić, Zagreb

Áron Keresztes, Budapest

Giorgio Socal, Venice

Branka Salopek Sondi, Zagreb

Damir Viličić, Zagreb

Jaromir Lukavsky, Třeboň

Lajos Vörös, Tihany

**Technical Editor-in-Chief:** Petra Peharec Štefanić, Zagreb, e-mail: [acta@biol.pmf.hr](mailto:acta@biol.pmf.hr)

**Managing Editor:** Mateja Jagić, Zagreb, e-mail: [acta@biol.pmf.hr](mailto:acta@biol.pmf.hr)

## Technical Editors:

Ana Car, Zagreb (Reference checking)

Renata Biba, Zagreb (Record keeping of submissions, archive)

Željka Vidaković-Cifrek, Zagreb (Plagiarism control)

Sandra Vitko, Zagreb (Creating extensible markup language-xml files)

**Exchange:** Grozdana Sirotić, e-mail: [grozdana.sirotic@biol.pmf.hr](mailto:grozdana.sirotic@biol.pmf.hr)

**Language editing:** Graham McMaster, Zagreb

**Acta Botanica Croatica is published two times yearly.**

**Annual subscription:** individuals, institutions and companies 50 EUR. Payment should be made after receipt of the invoice. The order should be sent to the Editorial Office ([acta@biol.pmf.hr](mailto:acta@biol.pmf.hr))

**Printing is financially supported** by the Ministry of Science and Education of the Republic of Croatia.

Financial support has been given by The Foundation of the Croatian Academy of Sciences and Arts (HAZU).

**Cover illustration:** *Degenia velebitica* (Degen) Hayek, published in Magyar Botanikai Lapok 8, 1909

**Cover design:** Juraj Balen

Acta Botanica Croatica offers **free access to full-text articles** on the Portal of the Scientific Journals of Croatia – Hrčak (<http://hrcak.srce.hr>) and [www.abc.botanic.hr](http://www.abc.botanic.hr)

**Printed by:** DENONA d.o.o. Zagreb

**On the cover:**

*Melaleuca williamsii* subsp. *synoriensis* was found along the banks of the Tiber River in Rome, Italy, representing its first record outside its native range. This finding is reported by Iamónico and Nicoletta (pp. 115-118).



## ACTA BOTANICA CROATICA

An international journal of botany  
issued by:  
the Department of Biology,  
Faculty of Science, University of Zagreb, Croatia

Vol. 83 (2)

Zagreb, October 2024

## ACTA BOTANICA CROATICA

The journal originally entitled *Acta Botanica Instituti Botanici Regalis Universitatis Zagrebensis* was founded in 1925. In 1957 its name was changed to Acta Botanica Croatica. In 1998, it became an entirely English-language journal.

The journal covers field (terrestrial and aquatic) and experimental research on plants and algae; including plant viruses and bacteria; from the subcellular level to ecosystems. Manuscripts focusing upon the lowland and karstic areas of southern Europe, karstic waters, other types of fresh water, and the Adriatic (Mediterranean) Sea are particularly welcome. More detailed information is available on the link

**<http://www.abc.botanic.hr/index.php/abc/about>**

The following points make Acta Botanica Croatica an attractive publishing medium: 1) article submission and publishing are free of charge, 2) manuscripts subject to international review, 3) covered by major abstracting and indexing services.

Impact Factor calculated by Journal Citation Reports:

1.1 (2023)

5-year Impact Factor: 1.2

Acta Botanica Croatica is a member of CrossCheck by iThenticate. iThenticate is a plagiarism screening service that verifies the originality of content submitted before publication. The iThenticate software checks submissions against millions of published research papers, documents on the web, and other relevant sources. Authors and researchers can also use the iThenticate system to screen their work before submission by visiting [research.ithenticate.com](http://research.ithenticate.com).

# Nomenclature of the Balkan alliance *Romuleion graecae* (*Poetea bulbosae*)

Massimo Terzi<sup>1\*</sup>, Nenad Jasprica<sup>2</sup>, Andraž Čarni<sup>3,4,5</sup>, Vlado Matevski<sup>4</sup>, Erwin Bergmeier<sup>6</sup>, Jean-Paul Theurillat<sup>7,8</sup>

<sup>1</sup> Institute of Bioscience and Bioresources (IBBR), National Council of Research (CNR), via Amendola 165/A, IT-70126, Bari, Italy

<sup>2</sup> University of Dubrovnik, Institute for Marine and Coastal Research, Kneza Damjana Jude 12, HR-20000 Dubrovnik, Croatia

<sup>3</sup> Institute of Biology, Research Centre of the Slovenian Academy of Sciences and Arts, Novi trg 2, SI-1000, Ljubljana, Slovenia

<sup>4</sup> Macedonian Academy of Sciences and Arts, Krste Misirkov 2, MK-1000 Skopje, Republic of North Macedonia

<sup>5</sup> University of Nova Gorica, School for Viticulture and Enology, Vipavska 13, SI-5000 Nova Gorica, Slovenia

<sup>6</sup> University of Göttingen, Albrecht-von-Haller Institute of Plant Sciences, Department of Vegetation Analysis and Plant Diversity, Untere Karspüle 2, D-37073 Göttingen, Germany

<sup>7</sup> Centre Alpien de Phytogéographie, Fondation J.-M. Aubert, Case postale 71, CH-1938 Champex-Lac, Switzerland

<sup>8</sup> University of Geneva, Section of Biology, Department of Botany and Plant Biology, Case postale 71, CH-1292 Chambésy, Switzerland

**Abstract** – The *Romuleion*, the only alliance of the order *Poetalia bulbosae* (class *Poetea bulbosae*) found on the Balkan Peninsula, represents Mediterranean perennial and ephemeral pastures. It has been found in several Balkan countries, from Greece to the Republic of North Macedonia, Montenegro, Croatia and Bulgaria. However, a revision of its nomenclature according to the fourth edition of the International Code of Phytosociological Nomenclature shows that the name of the alliance was not validly published. In this paper we therefore describe the new alliance *Romuleion graecae*, together with the new association *Plantagini lagopodis-Poetum bulbosae*.

**Keywords:** Balkan vegetation, ICPN, Mediterranean grassland, nomenclature, phytosociology, *Poetea bulbosae*, syntaxonomy

## Introduction

The class *Poetea bulbosae* is accepted in the standard European vegetation classification (EVC), as representing the “Mediterranean and Maghrebinian seasonal perennial and ephemeroïd pastures in the thermo- to oro-Mediterranean belts” (Mucina et al. 2016, EVS 2017, FloraVeg.EU 2023, Terzi et al. 2024).

The class includes only one order, *Poetalia bulbosae*, and six alliances, namely *Trifolio-Periballion*, *Plantaginion serrariae*, *Poo bulbosae-Astragalion*, *Ornithogalo corsici-Trifolion subterranei*, *Plantaginion cupanii*, and *Romuleion* (FloraVeg.EU 2023, Fernández-González et al. 2023, Terzi et al. 2024).

The distribution range of most of these alliances is restricted to the western Mediterranean, i.e. the Iberian Peninsula and the Tyrrhenian Islands, and to Italy (i.e. *Plantaginion cupanii*), while only the alliance *Romuleion* has been found on the Balkan Peninsula and its islands (Preislerová et al. 2022). More specifically, the *Romuleion* has been reported in Greece, the Republic of North Macedonia, Croatia, and Montenegro, while its occurrence is considered uncertain in Albania, Bulgaria, and Bosnia and Herzegovina (Oberdorfer 1954, de Bolòs et al. 1996, Čarni et al. 2014, Škvorc et al. 2017, Preislerová et al. 2022, Stanišić-Vujačić et al. 2023, Terzi and Jasprica 2024).

\* Corresponding author e-mail: massimo.terzi@cnr.it

The *Romuleion* was originally placed in the class “*Thero-Brachypodietea*” (Oberdorfer 1954), and then moved to the *Brachypodio-Chrysopogonetea* (Horvat et al. 1974), the *Saginetea maritimae* (Rodwell et al. 2002), and the *Tuberarietea guttatae* (Čarni et al. 2014). Finally, in the EVC, it has been framed within the *Poetea bulbosae* (Mucina et al. 2016).

Although the *Romuleion* is the only alliance of the class *Poetea bulbosae* occurring in the Balkans, with a potentially rather wide distribution, its original description raises some problems from a nomenclatural point of view (see also Čarni et al. 2014). Therefore, the aim of this paper is to revise the nomenclature of the alliance and thus contribute to the stabilisation of the European vegetation system by the use of correct names.

## Material and methods

The revision of the nomenclature of the alliance *Romuleion* is based on the fourth edition of the International Code of Phytosociological Nomenclature (ICPN, Theurillat et al. 2021), whose articles (Art.) are quoted in brackets in the text. The names of the syntaxa, which are given exactly as in the mentioned articles, are given in quotation marks (“”).

In order to assess possible synonymy between some of the names of the associations involved in the validation of the alliance name, Jaccard’s similarity index (J, Kent 2012) was calculated between the phytosociological synoptic tables of some of the associations.

The taxonomic nomenclature follows Euro+Med (2006–2023), while the syntaxonomic nomenclature follows FloraVeg.EU (2023) and its updates as proposed by Fernández-González et al. (2023) and Terzi et al. (2024).

To facilitate comparison with the EVC, the names of the syntaxa in the syntaxonomic scheme at the end of the discussion have been supplemented with the codes already assigned in the EVC (three digits for classes, five for orders and six for alliances).

## Results and discussion

The “*Romulion*” alliance [recte: *Romuleion*] was originally described by Oberdorfer (1954) with a diagnosis comprising two associations. Another, the “*Biareto-Poetum timoleontis*”, is merely mentioned as provisional (Art. 3b) and is moreover invalid as it lacks a sufficient original diagnosis (Art. 2b). The two substantiated associations, “*Tortileto-Poetum timoleontis*” and “*Lagopeto-Poetum timoleontis*”, are represented by a synoptic table derived from 14 and 17 relevés, respectively, from the surroundings of the city of Thessaloniki, Thessaly, Isthmus of Corinth, Attica, Southern Macedonia and Thrace (all in Greece). However, this synoptic table contains neither *Poa timoleontis* Boiss. (Fl. Orient. 5: 607. 1884), nor *Poa bulbosa* subsp. *timoleontis* (Boiss.) Hayek (Repert. Spec. Nov. Regni Veg. Beih. 30(3): 260. 1933), but only *Poa bulbosa*. Consequently, these asso-

ciations were not validly published because one of the name-giving taxa is missing in their diagnosis (Art. 3f), although mentioned in the text (Oberdorfer 1954: 89, “...von *Poa bulbosa* beherrschten Gesellschaften”). Since all associations assigned to the diagnosis of the alliance in Oberdorfer (1954) are thus invalid, the name *Romuleion* Oberdorfer 1954 is also invalid for lack of a sufficient diagnosis (Art. 2b).

In Horvat et al. (1974: 120–121) the two associations described by Oberdorfer (1954) within the *Romuleion* alliance, namely “*Tortileto-Poetum timoleontis*” and “*Lagopeto-Poetum timoleontis*”, were united under a single association name, “*Poetum timoleontis*”, with two subassociations (“Subass. mit *Stipa tortilis*” and “Subass. mit *Plantago lagopus*”). However, the diagnosis of the *Poetum timoleontis* is still based on the same synoptic relevés published by Oberdorfer (1954), where the name-giving taxon *Poa timoleontis* is missing. Consequently, the names *Poetum timoleontis* Oberdorfer ex Horvat, Glavač et Ellenberg 1974 (Art. 3f) and *Romuleion* Oberdorfer ex Horvat, Glavač et Ellenberg 1974 (Art. 2b) are still invalid names.

Later, Bolòs et al. (1996) renamed the alliance “*Romulion graecae* Oberd. 1954 em. nom. (*Romulion* Oberd.)” and assigned a new association, “*Airo elegantissimae-Trifolietum dalmaticae*”, from the Ionian island of Cephalonia (Greece). This association was validly published with a sufficient original diagnosis consisting of seven relevés (Table 12, on page 106). This was not so for the alliance because the name-giving taxon *Romulea linaresii* subsp. *graeca* Bég. (Bot. Jahrb. Syst. 38: 325, 1907) was missing in the only valid element of the diagnosis, namely in the relevés of the new association (the two associations of Oberdorfer (1954), which were also referred to, were invalidly published). The name *Romuleion graecae* Oberdorfer ex de Bolòs, Masalles, Ninot, et Vigo 1996 is therefore invalid under Art. 3f.

Čarni et al. (2014) provided a different interpretation on the validity of the names published by Oberdorfer (1954), considering the alliance *Romuleion* and the two associations “*Tortileto-Poetum timoleontis*” and “*Lagopeto-Poetum timoleontis*” as validly published. They argue that the taxon name “*Poa bulbosa*” given in the table on page 90 (“Liste I”) in Oberdorfer (1954) is a printing error for “*Poa bulbosa* coll.” as written in the two other tables (“Liste II” and “Liste III”) of the work on pages 92 and 94, respectively. In their view, “*Poa bulbosa* coll.” would include all subspecies of *Poa bulbosa* (or species of the *Poa bulbosa* aggregate), including *Poa timoleontis* (*Poa bulbosa* subsp. *timoleontis*), and would therefore comply with Art. 3f. The justification of this interpretation would be the statement made by Oberdorfer on page 88 that *Brachypodium ramosum* [recte: *B. retusum*] or *Brachypodium phoenicoides*, which are widespread in the western Mediterranean, are completely receding eastwards where they are replaced by various subspecies (“div. ssp.”) of *Poa bulbosa*. To some extent, such an interpretation might have been supported by the content of Art. 3f in the third edition of the ICPN (Weber et al. 2000), which only states that the name-giving taxon should be “indicated in

the original diagnosis". This interpretation is no longer valid with the present rules, as they state (Art. 3f Note 1) that the name-giving taxon must be present in the relevés belonging to the "original diagnoses of the associations that have been quoted in the original diagnosis of the alliance". Therefore, the taxon *Poa timoleontis*, which is not explicitly included in the collective species *Poa bulbosa*, is not mentioned in the synoptic table of the two associations. Consequently, the associations *Stipo tortilis-Poetum timoleontis* and *Lagopo-Poetum timoleontis* are not validly published in Oberdorfer (1954). In considering these two association names as validly published, Čarni et al. (2014) also made some corrections. In both names, the first name-giving taxon corresponds to a specific epithet used without the generic name, namely "Tortileto" for "*Stipa tortilis*" and "Lagopeto" for *Plantago lagopus*. In accordance with Art. 14b, Čarni et al. (2014) corrected the former name to *Stipo tortilis-Poetum timoleontis*. For the latter name, where the specific epithet "lagopus" is also a validly published generic name, they retained the association name and corrected the orthographic error ("Lagopo" instead of "Lagopeto"; Arts. 10a and 41b). Indeed, *Lagopus arvensis* Fourr. (Ann. Soc. Linn. Lyon sér. 2, 17: 140, 1869) is a valid name (International Plant Names Index, <https://www.ipni.org/n/32110-1>, accessed 3 Oct 2023) for *Plantago lagopus* L., and according to the third edition of the ICPN (Weber et al. 2000) this name should be retained as the name-giving taxon. However, as *L. arvensis* Fourr. is an illegitimate name, this would no longer be the case with the current rules (Art. 44). Čarni et al. (2014, p. 124) also corrected the second name giving-taxon of the name *Lagopo-Poetum timoleontis*, namely *Poa timoleontis*, to *Poa bulbosa*, in accordance with Art. 43, because they considered the presence of the taxon *Poa timoleontis* in the research area to be doubtful. On the other hand, they did not make this correction for the name *Stipo tortilis-Poetum timoleontis*, considering that the taxon *Poa timoleontis* could occur where Oberdorfer had originally sampled the association. Čarni et al. (2014) provided a neotype for the *Lagopo-Poetum timoleontis* Oberdorfer 1954 corr. Čarni et al. 2014, but the latter name was nevertheless not incidentally validated, as it was not reported as new (Art. 3i).

As the name *Lagopo-Poetum* remains invalidly published, we validate it, and correct the name here based on the results of Čarni et al. (2014), namely *Plantagini lagopodis-Poetum bulbosae* Čarni, Matevski, Šilc et Čušterevska ex Terzi, Jasprica, Čarni, Matevski, Bergmeier et Theurillat ass. nov. hoc loco. The original diagnosis of the new association includes relevés 1-12 of Table 1, on page 112, in Čarni et al. (2014), and its holotypus is relevé 5 in this table, which is the same relevé selected as the neotype for the invalid *Lagopo-Poetum bulbosae*.

A second association, *Romuleo graecae-Poetum bulbosae*, was also validly published by Čarni et al. (2014) and classified in the *Romuleion*. However, the name of this alliance was not validated either, as it was not reported as new (Art. 3i). Recently, Stanišić-Vujačić et al. (2023) maintained the same nomenclatural interpretation for the *Romuleion*

as Čarni et al. (2014), and they described two new associations to be included in this alliance, the *Romuleo bulbocodii-Poetum bulbosae* and the *Ornithogalo exscapi-Poetum bulbosae*.

Attempts to find a neotype for the "*Tortileto-Poetum timoleontis*" have failed, as no adequate relevé could be found. Two relevés containing *Poa bulbosa* together with *Stipa capensis* (= *S. tortilis*) were published by Čarni et al. (2014) among the relevés of the *Romuleo graecae-Poetum bulbosae* (relevés 14 and 20, Table 1). However, the floristic composition of these two relevés appears to be more similar to the other relevés of the *Romuleo graecae-Poetum bulbosae* than to those in the synoptic table of the "*Tortileto-Poetum timoleontis*" in Oberdorfer (1954). A comparison of the synoptic table of the original diagnosis of the *Plantagini lagopodis-Poetum bulbosae* in Čarni et al. (2014) (relevés 1-12, Table 1) showed that it is also closer to the *Romuleo graecae-Poetum bulbosae* ( $J = 0.51$ ) than to the relevés in Oberdorfer's synoptic table of the "*Lagopo-Poetum timoleontis*" ( $J = 0.27$ ) or the "*Tortileto-Poetum timoleontis*" ( $J = 0.27$ ). Conversely, a comparison between the two synoptic relevés of Oberdorfer (1954) showed that their floristic composition is very similar ( $J = 0.78$ ). Therefore, the syntaxonomic interpretation by Horvat et al. (1974), which considers Oberdorfer's two syntaxa to be two subassociations of the same association seems justified, and the *Lagopo-Poetum timoleontis* Oberdorfer 1954 could correspond to a different association than the *Plantagini lagopodis-Poetum bulbosae*. However, in order to verify this hypothesis, relevés from the regions from which Oberdorfer described his two associations are essential.

In terms of alliance, the *Romuleion* currently contains five associations (see the syntaxonomic scheme below). A sixth association (*Festuco valesiacae-Poetum bulbosae*) has recently been described (Terzi and Jasprica 2024), but questions remain about its inclusion in this alliance. Another undescribed association may occur in Crete, according to unpublished relevés of E. Bergmeier. Those new syntaxa are not considered in the present discussion. The name of the alliance is derived from the genus *Romulea* Maratti (Pl. Romul. Saturn. 13, 1772), which is present with two taxa in the original diagnoses of the five described associations, (1) *Romulea bulbocodium* (L.) Sebast. & Mauri (Fl. Roman. Prodr. 17, 1818) and (2) *Romulea linariesii* subsp. *graeca*. *Romulea bulbocodium* is a widespread Mediterranean species (Euro+Med 2006-2023) whereas *Romulea linariesii* subsp. *graeca* is a Balkan-Anatolian taxon occurring in Turkey, Greece, including the Aegean islands, and some Balkan countries (e.g. Hadžiablahović and Bulić 2004, Dimopoulos et al. 2013, Raycheva et al. 2021).

Since the alliance is centred in the southern Balkans, we follow the proposal of Bolòs et al. (1996) to consider *Romulea linariesii* subsp. *graeca* as the name-giving taxon, and we validate the name *Romuleion graecae* Oberdorfer ex Terzi, Jasprica, Čarni, Matevski, Bergmeier et Theurillat all. nov. hoc loco. The nomenclatural type (holotypus) of the new alliance is the *Romuleo graecae-Poetum bulbosae* Čarni,

Matevski, Šilc et Čušterevska 2014 (Čarni et al. 2014, p. 125) from the southern part of the Balkans (Greece). According to Oberdorfer (1954) and Čarni et al. (2014), the characteristic species of the alliance are: *Allium guttatum*, *Alyssum minutum*, *Alyssum repens*, *Campanula ramosissima*, *Gagea reticulata*, *Hedypnois rhagadioloides*, *Hypochaeris cretensis*, *Lagoecia cuminoides*, *Linaria simplex*, *Lotus angustissimus*, *Ornithogalum collinum*, *Ornithogalum armeniacum*, *Picris pauciflora*, *Romulea bulbocodium*, *Romulea linaresii* subsp. *graeca*, *Romulea columnae*, *Sedum aetnense*, *Silene graeca*, *Ziziphora capitata*.

Therefore, we propose the following syntaxonomic scheme (the author citation of the class follows Terzi et al. 2024):

BUL – *Poetea bulbosae* Rivas Goday et Rivas-Martínez ex Navarro Andrés et Valle Gutiérrez 1984

BUL-01 *Poetalia bulbosae* Rivas Goday et Rivas-Martínez in Rivas Goday et Ladero 1970

BUL-01F *Romuleion graecae* Oberdorfer ex Terzi, Jasprica, Čarni, Matevski, Bergmeier et Theurillat all. nov. hoc loco [holotypus: *Romuleo graecae-Poetum bulbosae* Čarni, Matevski, Šilc et Čušterevska 2014; synonyms: *Romuleion* Oberdorfer 1954 (Art. 2b), *Romuleion* Oberdorfer ex Horvat, Glavač et Ellenberg 1974 (Art. 2b), *Romuleion graecae* Oberdorfer ex de Bolòs, Masalles, Ninot et Vigo 1996 (Art. 3f)]

*Plantagini lagopodis-Poetum bulbosae* Čarni, Matevski, Šilc et Čušterevska ex Terzi, Jasprica, Čarni, Matevski, Bergmeier et Theurillat ass. nov. hoc loco [syn. *Lagopo-Poetum bulbosae* Oberdorfer 1954 corr. Čarni, Matevski, Šilc et Čušterevska 2014 (corr. superfl.)]

*Romuleo graecae-Poetum bulbosae* Čarni, Matevski, Šilc et Čušterevska 2014

*Airo elegantissimae-Trifolietum dalmatici* Bolòs, Masalles, Ninot et Vigo 1996

*Romuleo bulbocodii-Poetum bulbosae* Stanišić-Vujačić, Stešević, Hadžiablahović et Šilc 2023

*Ornithogalo exscapi-Poetum bulbosae* Stanišić-Vujačić, Stešević, Hadžiablahović et Šilc 2023

*Poetum timoleontis* Oberdorfer ex Horvat, Glavač et Ellenberg 1974 (Art. 3f) [incl. *Lagopo-Poetum timoleontis* Oberdorfer 1954 (Art. 3f), *Tortileto-Poetum timoleontis* Oberdorfer 1954 (recte: *Stipo capensis-Poetum timoleontis*) (Art. 3f)]

## Acknowledgments

The authors thank Steve Latham (United Kingdom) for improving the English, and Kiril Vassilev for his help in trying to find a neotype for the *Tortileto-Poetum timoleontis* Oberdorfer 1954. Additional thanks are extended to two anonymous reviewers for their comments and suggestions.

A.Č. is funded by the ARIS (P1-0236). The other authors declare that no funds, grants, or other support were received during the preparation of this manuscript.

## Author contribution statement

M.T. and J.-P.T. conceived and wrote the manuscript; all authors critically revised and approved the manuscript.

## References

- Bolòs, O. de, Masalles, R. M., Ninot, J. M., Vigo, J., 1996: A survey on the vegetation of Cephalonia (Ionian islands). *Phytocoenologia* 26(1), 81–123.
- Čarni, A., Matevski, V., Šilc, U., Čušterevska, R., 2014: Early spring ephemeral therophytic non-nitrophilous grasslands as a habitat of various species of *Romulea* in the southern Balkans. *Acta Botanica Croatica* 73(1), 155–177. <https://doi.org/10.2478/botcro-2013-0017>
- Dimopoulos, P., Raus, T., Bergmeier, E., Constantinidis, T., Iatrou, G., Kokkini, S., Strid, A., Tzanoudakis, D., 2013: Vascular plants of Greece: An annotated checklist. *Englera* 31, 1–372.
- Euro+Med, 2006-2023: Euro+Med PlantBase – the information resource for Euro-Mediterranean plant diversity. Retrieved January 25, 2024 from <http://www.europplusmed.org/>
- EVS [IAVS Working Group European Vegetation Survey], 2017: Procedures for updating the standard European vegetation classification. Retrieved April 2021 from <http://euroveg.org/download/EuroVegChecklistupdate-procedures-2017-09-14-APPROVED.pdf>
- Fernández-González, F., Terzi, M., Di Pietro, R., Theurillat, J. P., 2023: Proposals (33–34) to conserve the name *Poo-Astragalion* and to conserve the name *Poo-Astragaletum sesamei* with a conserved type, and requests (5–7) for a binding decision on the name-giving taxa in the same names and the inversion of the name *Poo-Astragaletum sesamei*. *Vegetation Classification and Survey* 4, 203–207. <https://doi.org/10.3897/VCS.108769>
- FloraVeg.EU, 2023: Database of European Vegetation, Habitats and Flora. Retrieved October 2, 2023 from [www.floraveg.eu](http://www.floraveg.eu)
- Hadžiablahović, S., Bulić, Z., 2004: On distribution of some Balkan endemic and rare species in the flora of Montenegro. *Glasnik Republičkog Zavoda za Zaštitu Prirode Podgorica* 27–28, 43–50.
- Horvat, I., Glavač, V., Ellenberg, H., 1974: *Vegetation Südos-teuropas*. *Geobotanica selecta* 4. Gustav Fischer Verlag, Stuttgart.
- Kent, M., 2012: *Vegetation description and data analysis. A practical approach*. Ed. 2. John Wiley & Sons, Chichester, UK.
- Mucina, L., Bültman, H., Dierssen, K., Theurillat, J.-P., Dengler, J., Čarni, A., Šumberová, K., Raus, T., Di Pietro, R., Gavilán García, R., Chytrý, M., Iakushenko, D., Schaminée, J. H. J., Bergmeier, E., Santos Guerra, A., Daniëls, F. J. A., Ermakov, N., Valachovič, M., Pigantti, S., Rodwell, J. S., Pallas, J., Cape-lo, J., Weber, H. E., Lysenko, T., Solomeshch, A., Dimopoulos, P., Aguiar, C., Freitag, H., Hennekens, S. M., Tichý, L., 2016: *Vegetation of Europe: Hierarchical floristic classification system of plant, lichen, and algal communities*. *Applied Vegetation Science* 19(S1), 3–264. <https://doi.org/10.1111/avsc.12257>
- Oberdorfer, E., 1954: Nordägäische Kraut- und Zwergstrauchfluren im Vergleich mit den entsprechenden Vegetationseinheiten des westlichen Mittelmeergebietes. *Vegetatio* 5, 88–96. <https://doi.org/10.1007/BF00299558>



- Preislerová, Z., Jiménez-Alfaro, B., Mucina, L., Berg, C., Bonari, G., Kuzemko, A., Landucci, F., Marcenò, C., Monteiro-Henriques, T., Novák, P., Vynokurov, D., Bergmeier, E., Dengler, J., Apostolova, I., Bioret, F., Biurrun, I., Campos, J. A., Capelo, J., Čarni, A., Çoban, S., Csiky, J., Čuk, M., Čušterevska, R., Daniëls, F. J. A., De Sanctis, M., Didukh, Ya., Dítě, D., Fanelli, F., Golovanov, Y., Golub, V., Guarino, R., Hájek, M., Iakushenko, D., Indreica, A., Jansen, F., Jašková, A., Jiroušek, M., Kalníková, V., Kavğacı, A., Kucherov, I., Kůzmič, F., Lebedeva, M., Loidi, J., Lososová, Z., Lysenko, T., Milanović, Đ., Onyshchenko, V., Perrin, G., Peterka, T., Rašomavičius, V., Rodríguez-Rojo, M. P., Rodwell, J. S., Růšina, S., Sánchez Mata, D., Schaminée, J. H. J., Semenishchenkov, Y., Shevchenko, N., Šibík, J., Škvorc, Ž., Smagin, V., Stešević, D., Stupar, V., Šumberová, K., Theurillat, J.-P., Tikhonova, E., Tzonev, R., Valachovič, M., Vassilev, K., Willner, W., Yamalov, S., Večeřa, M., Chytrý, M., 2022: Distribution maps of vegetation alliances in Europe. *Applied Vegetation Science* 25(1), e12642. <https://doi.org/10.1111/avsc.12642>
- Raycheva, T., Stoyanov, K., Randelović, V., Uzundzhaliyeva, K., Marinov, J., Trifonov, V. 2021: Overview of the floristic and taxonomic studies on Iridaceae Juss. in Bulgaria. *Thaiszia* 31(1), 87–104. <https://doi.org/10.33542/TJB2021-1-07>
- Rodwell, J. R., Schaminée, J. H. J., Mucina, L., Pignatti, S., Dring, J., Moss, D., 2002: The diversity of European vegetation. An overview of phytosociological alliances and their relationship to EUNIS habitats. Report EC-LNV 2002/054, Wageningen.
- Škvorc, Ž., Jasprica, N., Alegro, A., Kovačić, S., Franjić, J., Krstonošić, D., Vraneša, A., Čarni, A., 2017: Vegetation of Croatia: phytosociological classification of the high-rank syntaxa. *Acta Botanica Croatica* 76(2), 200–224. <https://doi.org/10.1515/botcro-2017-0014>
- Stanišić-Vujačić, M., Stešević, D., Hadžiallahović, S., Šilc, U., 2023: Ecological and syntaxonomical characteristics of early spring therophytic ephemeral grasslands (alliance *Romuleion*) in the northeastern Mediterranean. *Plant Biosystems* 157(3), 540–563. <https://doi.org/10.1080/11263504.2023.2165570>
- Terzi, M., Fernández-González, F., Di Pietro, R., Theurillat, J.-P., 2024: Phytosociological nomenclature of the class names *Helianthemetea guttati*, *Poetea bulbosae* and *Stipo giganteae-Agrostietea castellanae*. *Plant Biosystems* 158(1), 70–83. <https://doi.org/10.1080/11263504.2023.2287539>
- Terzi, M., Jasprica, N., 2024: Changes in grassland vegetation on the island of Plavnik (Croatia) over 100 years. *Acta Botanica Croatica* 83(2), 119–134. <https://doi.org/10.37427/botcro-2024-014>
- Theurillat, J.-P., Willner, W., Fernández-González, F., Bültmann, H., Čarni, A., Gigante, D., Mucina, L., Weber, H., 2021: International code of phytosociological nomenclature. 4th edition. *Applied Vegetation Science* 24(1), e12491. <https://doi.org/10.1111/avsc.12491>
- Weber, H. E., Moravec, J., Theurillat, J.-P., 2000: International Code of Phytosociological Nomenclature. 3rd edition. *Journal of Vegetation Science* 11(5), 739–768. <https://doi.org/10.2307/3236580>

# The taxonomy and distribution of algae in the thermal springs of Türkiye

Sevilay Öztürk\*, Oğuz Kurt

Manisa Celal Bayar University, Faculty of Sciences and Letters, Department of Biology, Manisa 45140, Türkiye

**Abstract** – The algal flora and physio-chemical parameters of seven thermal springs in Denizli were studied for the first time. Samples for algal analyses were taken monthly between May 2013 and June 2014, while the physio-chemical parameters were measured seasonally. The mean pH value of the thermal springs was 6.3, and temperatures varied between 34–60 °C. The significant differences ( $P < 0.001$ ) among the thermal springs were in their temperature and pH, as well as concentration of  $\text{Cl}^-$ ,  $\text{Fe}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Li}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{HCO}_3^-$ , and  $\text{SO}_4^{2-}$  ions. A total of 43 Cyanobacteria and three Bacillariophyta taxa were determined. The most common taxon was *Spirulina subsalsa* Oersted ex Gomont, sampled from five sampling sites. According to the principal component analysis (PCA), the most important determining factor for the algae was temperature, followed by concentration of  $\text{K}^+$  and  $\text{Cl}^-$  ions.

**Keywords:** cyanobacteria, diatoms, ecology, principal component analysis, thermal algae, travertine

## Introduction

Algae are commonly found in seas and fresh or brackish waters. However, they can survive in deserts, moist soils and stones, thermal springs, snow at the poles, and in all conditions with very little moisture. Thermal springs, in particular, are extreme ecosystems for algae. Certain groups of algae are common in thermal springs, such as Cyanobacteria and Bacillariophyta. Examining the species living in thermal springs is important in determining the factors conducive to life in these environments. Thus, the potential economic value of these species is revealed, and their biotechnological use improves.

Because it is located on fault lines, Türkiye has numerous thermal springs, concentrated mainly in the west of the country. The city of Denizli has numerous and varied thermal springs, which are famous worldwide for travertines and historical and cultural features. Travertines are formed by the precipitation of calcium in layers like thick lime deposits (Güner 1970, Pentecost et al. 1997). The most important characteristic of thermal springs in Denizli is that, although they are very close, they show different physio-chemical parameters (Kozak 2020). These thermal springs are in Pamukkale, Karahayıt, Yenicekent, Inalti, Şanlıalp, Umut, and Gölemezli. While Karahayıt, Yenicekent, and Umut have deep circulation and a high temperature, Pamukkale and Gölemezli have a low temperature and shallow water characteristics (Yaman and Özgür 2005). Pamukkale, or

“Hierapolis,” is on the UNESCO list of world heritage sites and is perhaps one of the oldest spa centers in Türkiye. The Hellenistic spa town of Hierapolis was a focus of interest for visitors at the end of the second century BC (UNESCO 2022).

The first study in Türkiye about the biology of thermal springs was from Pamukkale (Regel and Skuja 1937), and there are other studies on the algal flora of the Pamukkale thermal spring (Güner 1966, Pentecost et al. 1997, Altunöz et al. 2016, Öztürk Ulcay et al. 2017). Also, Aysel et al. (1992) examined cyanobacteria in the Ilıksu thermal springs (Zonguldak, Türkiye) and reported 33 Cyanobacteria taxa. Öztürk Ulcay et al. (2007) explored the thermal springs in Dikili (İzmir, Türkiye) and identified 19 Cyanobacteria taxa. Öztürk Ulcay and Kurt (2017) identified a total of 27 taxa (21 Cyanobacteria, five Bacillariophyceae and one Conjugatophyceae) in Alangüllü (Aydın, Türkiye). Öztürk (2020) reported 13 Cyanobacteria taxa in the thermal springs in Kütahya (Türkiye). Thermal algae studies have also intensified in the west of Türkiye where many thermal springs are located. Although there are many thermal springs in Denizli, thermal algae studies in this region have only been carried out in the Pamukkale and Karahayıt thermal springs.

The aim of the present study was to provide for the first time a study of the algal flora in the thermal springs of Denizli and their relationship to the ecological conditions of seven thermal springs in Denizli.

\* Corresponding author e-mail: seviozturk@yahoo.com

## Material and methods

### Sampling

Denizli is one of the important thermal regions of Türkiye for health tourism and the touristic thermal springs there were chosen as the study area (Fig. 1). Gölemezli (D1; 37°59' N 29°02' E), Yenicekent (D2; 38°02' N 28°57' E), İnaltı (D3; 37°54' N 28°45' E), Şanlıalp (D4; 37°57' N 29°04' E), Umut (D5; 37°55' N 28°49' E), Pamukkale (D6; 37°55' N 29°07' E) (Fig. 2), and Karahayıt (D7; 37°58' N 29°06' E) (Fig. 3). Samples were taken from several points (thermal water outlet points, natural and artificial pools, travertines, and thermal water channels) of these thermal springs. Sample sites were scattered over approximately 207 km<sup>2</sup>, ranging from an altitude of 148 m to 420 m a.s.l.

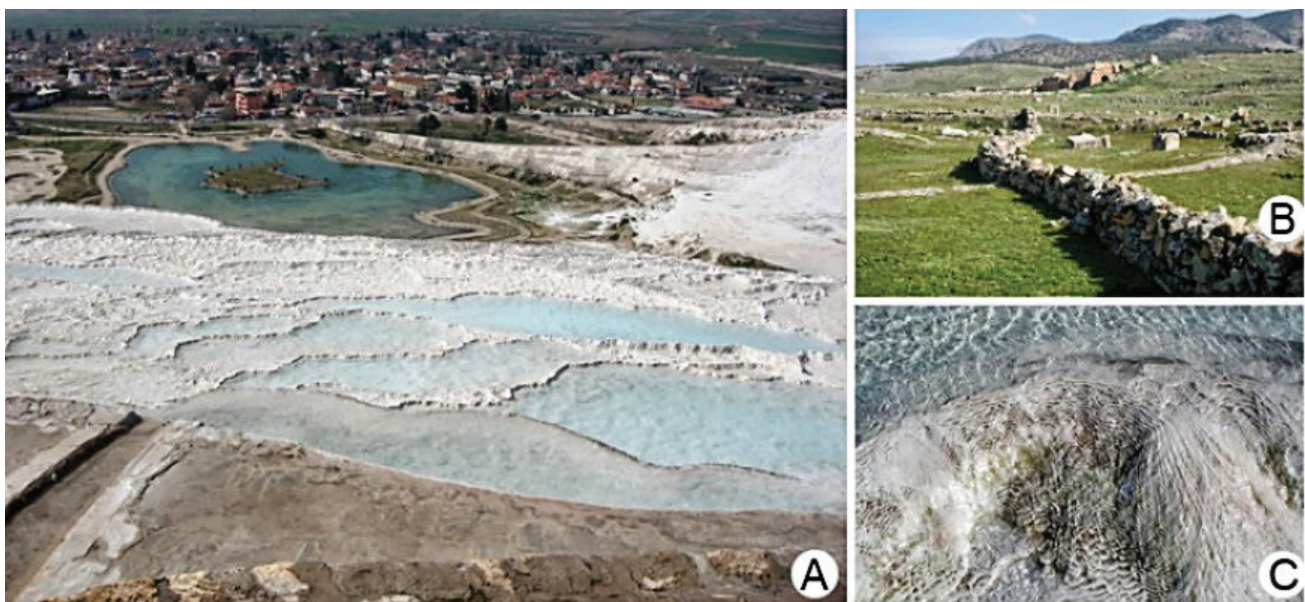
Algae sampling was done monthly between May 2013 and June 2014 using forceps, a spatula, and a plankton net with a 30 µm mesh size. Two algae samples were taken separately from each sample site in 50 mL falcon tubes. Then, a 4% formaldehyde solution was added to one of the samples for fixing. The second algae sample was used for identification. The algae samples were labeled and brought to the laboratory in the dark. Algae samples were examined under an Olympus BX 50 (phase-contrast) microscope and photographed using a Sony DSC-TX7 camera in the laboratory. The literature was used to identify the algae taxa: Komárek and Anagnostidis (1999, 2005), and Komárek (2013) for Cyanobacteria taxa and Cantonati et al. (2017) for Bacillariophyta. The nomenclature was checked on the AlgaeBase database (Guiry and Guiry 2023).

Water samples were taken simultaneously in sterile bottles during the algae sampling. The pH and temperature were measured *in-situ*, with a Hanna HI 9812-5 Portable Meter. Sodium (Na<sup>+</sup>), potassium (K<sup>+</sup>), calcium (Ca<sup>2+</sup>), lithium (Li<sup>+</sup>), magnesium (Mg<sup>2+</sup>), ferrous (Fe<sup>2+</sup>), chloride (Cl<sup>-</sup>), bicar-

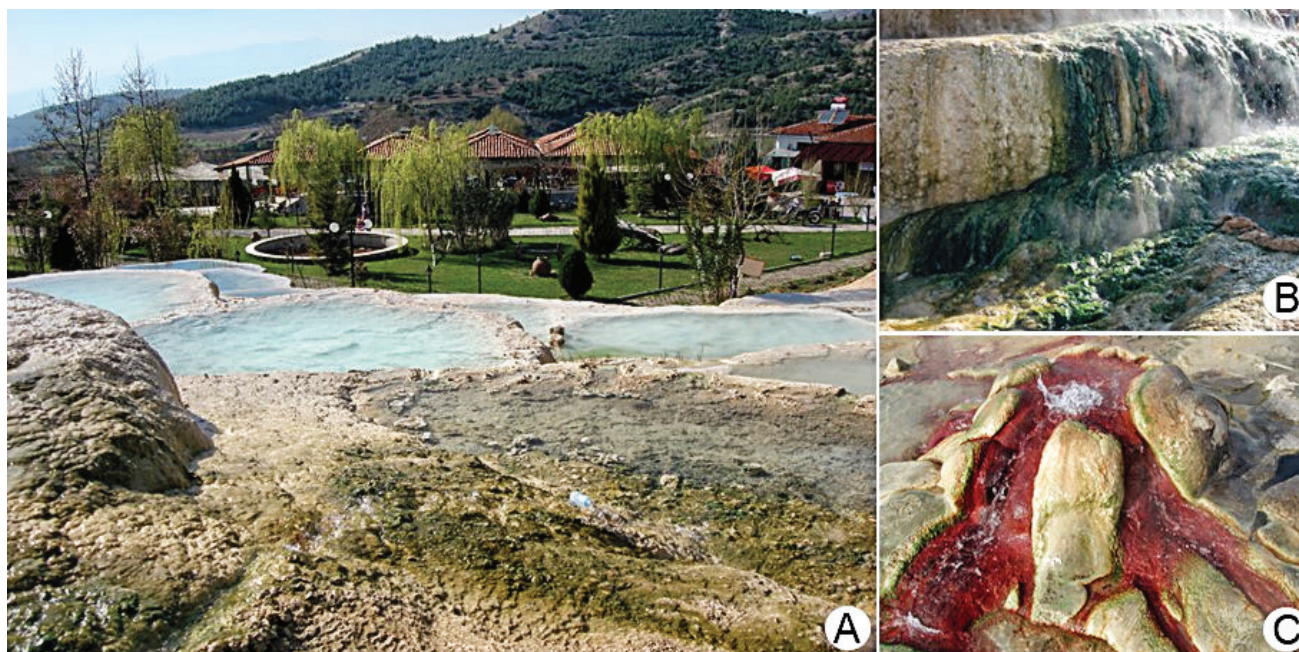


**Fig. 1.** The map of the study area showing the investigated thermal springs, and their locations in Denizli (Türkiye). D1 – Gölemezli, D2 – Yenicekent, D3 – İnaltı, D4 – Şanlıalp, D5 – Umut, D6 – Pamukkale, D7 – Karahayıt.

bonate (HCO<sub>3</sub><sup>-</sup>), and sulphate (SO<sub>4</sub><sup>2-</sup>) analyses were made *ex-situ*. The concentrations of Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Li<sup>+</sup>, Mg<sup>2+</sup>, and Fe<sup>2+</sup> ions were measured by atomic absorption spectrometer, the concentrations of Cl<sup>-</sup> and HCO<sub>3</sub><sup>-</sup> ions were measured by titrimetric analysis, the concentration of SO<sub>4</sub><sup>2-</sup> ion was measured by theoretical methods, and the ion concentration results are given as mg L<sup>-1</sup>. *In-situ* measurements were made every month (between May 2013 and June 2014), while *ex-situ* measurements were made seasonally (in July and October 2013 and in January and April 2014). The results of *in-situ* and *ex-situ* measurements are given as their means.



**Fig. 2.** The Pamukkale thermal spring (D6). A – general view of travertines, B – the Hellenistic spa town of Hierapolis, C – details of the travertines (photo: Sevilay Öztürk).



**Fig. 3.** The Karahayıt thermal spring (D7). A – general view of travertines, B – details of the travertines, C – thermal water outlet points (photo: Sevilay Öztürk).

### Statistical analysis

Canoco 5.0 software for Windows (Microcomputer Power, Ithaca, NY, USA) was used to determine the correlation between the physio-chemical characteristics of the thermal springs and algal flora (Ter Braak and Šmilauer 2012). Firstly, detrended correspondence analysis (DCA) was done to determine the gradient length and whether the studied gradient was suitable for linear or unimodal models. The results of the DCA were ideal for principal component analysis (PCA). The algal flora composition (total of 46 taxa from 7 sample sites on 12 sampling dates) with binary data was used in this analysis. Tests of significance of the first and all canonical axes were performed to statistically assess the relation between algal flora composition and physio-chemical characteristics (Monte Carlo test: 499 permutations under the reduced model).

Additionally, the Monte Carlo permutation test was applied to determine the statistical significance of physio-chemical characteristics in explaining the composition of algal flora. This involved a stepwise "forward selection" of explanatory variables, as available in Canoco. The process was begun by selecting the most effective explanatory variable (the one that best explained the overall variance in the data). Subsequent variables were chosen based on their decreasing importance in explaining the total variance in the dataset, in conjunction with the previously selected variables. The statistical significance of each variable was also assessed. The variation in algal flora composition explained by the physio-chemical characteristics included in the analysis was expressed as a percentage, representing the ratio of the sum of all canonical eigenvalues to the total variance (total inertia).

The Kruskal-Wallis H-test was conducted using SPSS 28.0 software to determine the statistical significance of the difference in values of physio-chemical parameters of thermal springs by sample sites. The Kruskal-Wallis test used the 12-month means of pH and temperature values and the seasonal means of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Li}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Cl}^-$ ,  $\text{HCO}_3^-$ , and  $\text{SO}_4^{2-}$  values of water samples from seven thermal springs. None of these data were transformed. The Kruskal-Wallis test is a non-parametric analysis of variance test used to assess the significance of differences among means of three or more groups in cases where the data does not follow a normal distribution.

## Results

### Physio-chemical characteristics of the thermal springs

Differences were observed among the physio-chemical parameters of seven thermal springs in Denizli (Tab. 1).

The highest temperature value was measured at D3 as 60 °C, the lowest temperature value was measured at D6 as 34 °C, and the mean temperature value of all sampling sites was 47 °C. In terms of calcium values, D7 had the highest value of 377.10 mg L<sup>-1</sup>, while D3 had the lowest value of 8.7 mg L<sup>-1</sup>. With potassium values, D4 had the highest value of 134.0 mg L<sup>-1</sup>, while D6 had the lowest value of 7.5 mg L<sup>-1</sup>. A difference in ferrous values was also seen among sample sites, with D7 having the highest value of 2.45 mg L<sup>-1</sup> and D3 having the lowest value of 0.034 mg L<sup>-1</sup>. In terms of bicarbonate values, D3 (689.53 mg L<sup>-1</sup>) is quite low compared to the other sample sites. It was observed that the lithium values of D6 and D7 were considerably lower than those of the other sample sites. The measured sodium values at D4 (1392.2 mg L<sup>-1</sup>) were twenty times higher than at D6

**Tab. 1.** The physio-chemical parameters of the thermal springs (D1-D7) in Denizli (Türkiye) and the results of the Kruskal-Wallis test (with SPSS 28.0) were used to reveal the statistical significance of the similarities and differences of the physio-chemical parameters of the sample sites ( $P < 0.001$ ). D1 – Gölemezli, D2 – Yenicekent, D3 – İnaltı, D4 – Şanlıalp, D5 – Umut, D6 – Pamukkale, D7 – Karahayıt. T – temperature,  $\text{Ca}^{2+}$  – calcium,  $\text{Mg}^{2+}$  – magnesium,  $\text{Fe}^{2+}$  – ferrous,  $\text{SO}_4^{2-}$  – sulphate,  $\text{K}^+$  – potassium,  $\text{Na}^+$  – sodium,  $\text{Cl}^-$  – chloride, and  $\text{Li}^+$  – lithium ions. The Kruskal-Wallis column contains the test values among the sample sites.

Parameters	Sample sites							Kruskal-Wallis test H	df	P-value
	D1	D2	D3	D4	D5	D6	D7			
$\text{Na}^+$ (mg L <sup>-1</sup> )	Mean ± SD 646.4 ± 12.05	Mean ± SD 725.5 ± 10.69	Mean ± SD 1019.6 ± 23.37	Mean ± SD 1392.2 ± 19.48	Mean ± SD 1235.3 ± 66.93	Mean ± SD 68.6 ± 1.29	Mean ± SD 176.5 ± 2.45	26.483	6	0.001
$\text{K}^+$ (mg L <sup>-1</sup> )	Mean ± SD 80.35 ± 1.32	Mean ± SD 103.3 ± 1.77	Mean ± SD 100.5 ± 2.24	Mean ± SD 134 ± 3.16	Mean ± SD 117.3 ± 4.04	Mean ± SD 7.5 ± 0.16	Mean ± SD 33.5 ± 1.06	26.283	6	0.001
$\text{Ca}^{2+}$ (mg L <sup>-1</sup> )	Mean ± SD 226 ± 10.61	Mean ± SD 197.8 ± 7.73	Mean ± SD 8.7 ± 0.25	Mean ± SD 12 ± 0.84	Mean ± SD 9.9 ± 0.08	Mean ± SD 350.275 ± 5.68	Mean ± SD 377.1 ± 4.4	26.490	6	0.001
$\text{Li}^+$ (mg L <sup>-1</sup> )	Mean ± SD 1.5 ± 0.04	Mean ± SD 1.02 ± 0.00	Mean ± SD 2.35 ± 0.08	Mean ± SD 5.44 ± 0.08	Mean ± SD 4.88 ± 0.08	Mean ± SD 0.16 ± 0	Mean ± SD 0.35 ± 0	26.643	6	0.001
$\text{Mg}^{2+}$ (mg L <sup>-1</sup> )	Mean ± SD 178.4 ± 3.77	Mean ± SD 77.3 ± 1.02	Mean ± SD 38.9 ± 0.62	Mean ± SD 42.3 ± 0.58	Mean ± SD 38.9 ± 0.6	Mean ± SD 150 ± 3.16	Mean ± SD 167 ± 3.65	26.024	6	0.001
$\text{Fe}^{2+}$ (mg L <sup>-1</sup> )	Mean ± SD 0.07 ± 0.00	Mean ± SD 2.19 ± 0.02	Mean ± SD 0.034 ± 0.00	Mean ± SD 0.07 ± 0	Mean ± SD 0.047 ± 0	Mean ± SD 0.06 ± 0	Mean ± SD 2.45 ± 0.02	26.157	6	0.001
$\text{Cl}^-$ (mg L <sup>-1</sup> )	Mean ± SD 86 ± 1.41	Mean ± SD 68 ± 1.41	Mean ± SD 150 ± 2.16	Mean ± SD 150 ± 1.82	Mean ± SD 175 ± 3.16	Mean ± SD 15 ± 0	Mean ± SD 29 ± 0.81	26.117	6	0.001
$\text{HCO}_3^-$ (mg L <sup>-1</sup> )	Mean ± SD 1293.62 ± 60.91	Mean ± SD 2007.56 ± 22.66	Mean ± SD 689.53 ± 8.34	Mean ± SD 2526.23 ± 31.13	Mean ± SD 1629.23 ± 57.47	Mean ± SD 1177.6 ± 10.55	Mean ± SD 1348.54 ± 25.93	26.195	6	0.001
$\text{SO}_4^{2-}$ (mg L <sup>-1</sup> )	Mean ± SD 1740 ± 37.21	Mean ± SD 705 ± 4.54	Mean ± SD 1576 ± 10.42	Mean ± SD 1336 ± 14.98	Mean ± SD 1184 ± 53.75	Mean ± SD 630 ± 7.61	Mean ± SD 889 ± 7.07	26.483	6	0.001
pH	Mean ± SD 6.2 ± 0.05	Mean ± SD 6.7 ± 0.05	Mean ± SD 6.4 ± 0.08	Mean ± SD 6.2 ± 0.05	Mean ± SD 6.2 ± 0.05	Mean ± SD 59 ± 0.08	Mean ± SD 6.025 ± 0.05	25.089	6	0.001
T (°C)	Mean ± SD 46 ± 1.41	Mean ± SD 55 ± 3.16	Mean ± SD 60 ± 1.82	Mean ± SD 54 ± 2.16	Mean ± SD 50 ± 2.44	Mean ± SD 34 ± 1.15	Mean ± SD 54 ± 0.81	23.648	6	0.001

(68.6 mg L<sup>-1</sup>) (Tab. 1). The D6 and D7 thermal springs were clearly distinguished from the others by their high value of  $\text{Ca}^{2+}$  concentration and low  $\text{Cl}^-$ ,  $\text{Na}^+$ , and  $\text{K}^+$  concentrations. In this study, it was determined that the most important physio-chemical parameter was temperature, and considering the temperature differences of thermal springs, it was seen that the most significant difference was found for D6, which has the lowest temperature. Another notable point was the differences in the concentration of  $\text{Fe}^{2+}$  which was higher in D2 and D7 than in other thermal springs (Tab. 1). A significant difference was found when the  $\text{Cl}^-$ ,  $\text{Fe}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Li}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{HCO}_3^-$ ,  $\text{SO}_4^{2-}$ , pH and temperature values of the thermal springs were compared with the Kruskal Wallis test ( $P < 0.001$ ) (Tab. 1).

### Algal flora

A total of 46 algae taxa (43 Cyanobacteria, 3 Bacillariophyta) were identified according to their morphological and ecological characteristics (Tab. 2). Synechococcales was the dominant order with 18 taxa among other Cyanobacteria orders. The D5 sample site had the most biodiversity with 22 taxa. The most common taxon was *Spirulina subsalsa* Oersted ex Gomont, collected from the five sample sites, followed by *Jaaginema geminatum* (Schwabe ex Gomont) Anagnostidis & Komárek, and *Pseudanabaena mucicola* (Naumann & Huber-Pestalozzi) Schwabe from four sample sites. In the present study, three Bacillariophyta taxa were identified from the sample sites of D5 and D6 that had water temperatures of 32–36 °C.

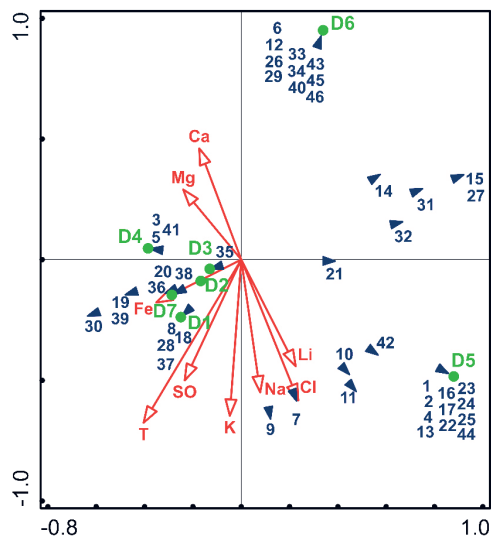
### Statistical analysis

First, the DCA was performed to find a suitable analysis, and then gradient lengths were assessed (Axis 1: 0.7356; Axis 2: 0.4883). According to the gradient lengths obtained from the DCA, the data were found to be suitable for PCA. Among the physio-chemical parameters analyzed, nine were included in the forward selection (temperature,  $\text{Cl}^-$ ,  $\text{SO}_4^{2-}$ ,  $\text{Fe}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Li}^+$ , and  $\text{Ca}^{2+}$ ). In the PCA, the identified 46 taxa, the investigated thermal springs, and the physio-chemical parameters were used (Fig. 4).

The significance effect was supported by a Monte Carlo permutation test (499 permutations, F-ratio < 0.1, P-value = 1). As a result of the PCA, the total variation was 49.71429, and the first two axes explained 58.59% of the variance (Fig. 4). The most determining factor was the temperature (T), followed by  $\text{K}^+$  and  $\text{Cl}^-$ . Although D5 (with 22 taxa) and D6 (with 16 taxa) had the highest number of taxa, they did not closely correlate with all the parameters (Fig. 4). Interestingly, D6 is partially positively correlated to  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  and negatively correlated to the rest. Conversely, D5 is negatively correlated to  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ , while it is partially positively correlated to the rest. Additionally, it can be seen that all the other sample sites are positively correlated to almost all the physio-chemical parameters (Fig. 4).

**Tab. 2.** Taxa, taxa codes and distribution of algal flora determined in thermal springs (D1-D7) in Denizli (Türkiye). D1 – Gölemezli, D2 – Yenicekent, D3 – İnalı, D4 – Şanlıhalp, D5 – Umut, D6 – Pamukkale, D7 – Karahayıt.

Taxa Code	Taxa	Sample Sites
Cyanobacteria		
1	<i>Synechococcus nidulans</i> (Pringsheim) Komárek in Bourrelly	D5
2	<i>Merismopedia</i> sp.	D5
3	<i>Jaaginema</i> sp.	D4
4	<i>Jaaginema angustissimum</i> (West & G.S. West) Anagnostidis & Komárek	D5
5	<i>Jaaginema subtilissimum</i> (Kützing ex De Toni) Anagnostidis & Komárek	D4
6	<i>Jaaginema pseudogeminatum</i> (G.Schmid) Anagnostidis & Komárek	D6
7	<i>Jaaginema geminatum</i> (Schwabe ex Gomont) Anagnostidis & Komárek	D1, D2, D3, D5
8	<i>Pseudanabaena</i> sp.	D1
9	<i>Pseudanabaena mucicola</i> (Naumann & Huber-Pestalozzi) Schwabe	D1, D2, D5, D7
10	<i>Pseudanabaena minima</i> (G.S.An) Anagnostidis	D5, D7
11	<i>Pseudanabaena catenata</i> Lauterbor	D1, D5
12	<i>Leibleinia epiphytica</i> (Hieronymus) Compère	D6
13	<i>Leptolyngbya</i> sp.	D5
14	<i>Leptolyngbya subtilis</i> (West) Anagnostidis	D4, D5, D6
15	<i>Leptolyngbya foveolarum</i> (Gomont) Anagnostidis & Komárek	D5, D6
16	<i>Leptolyngbya boryana</i> (Gomont) Anagnostidis & Komárek	D5
17	<i>Leptolyngbya gelatinosa</i> (Woronichin) Anagnostidis & Komárek	D5
18	<i>Planktolyngbya contorta</i> (Lemmermann) Anagnostidis & Komárek	D7
19	<i>Spirulina subtilissima</i> Kützing ex Gomont	D1, D4
20	<i>Spirulina major</i> Kützing ex Gomont	D3, D7
21	<i>Spirulina subsalsa</i> Oerstedt ex Gomont	D1, D2, D5, D6, D7
22	<i>Spirulina robusta</i> H.Welsh	D5
23	<i>Chroococcus</i> sp.	D5
24	<i>Chroococcus membraninus</i> (Meneghini) Nägeli	D5
25	<i>Cyanosarcina</i> sp.	D5
26	<i>Gloeocapsa gelatinosa</i> Kützing	D6
27	<i>Gloeocapsopsis cyanea</i> (Krieger) Komárek & Anagnostidis	D5, D6
28	<i>Geitlerinema</i> sp.	D1
29	<i>Kamptonema jaszovense</i> (Vouk) Strunecký, Komárek & J.Smarda	D6
30	<i>Kamptonema okenii</i> (C.Agardh ex Gomont) Strunecký, Komárek & J.Smarda	D1, D4, D7
31	<i>Kamptonema animale</i> (C.Agardh ex Gomont) Strunecký, Komárek & J.Smarda	D3, D5, D6
32	<i>Kamptonema cortianum</i> (Meneghini ex Gomont) Strunecký, Komárek & J.Smarda	D1, D5, D6
33	<i>Microcoleus autumnalis</i> (Gomont) Strunecky, Komárek & J.R.Johansen in Strunecky	D6
34	<i>Porphyrosiphon versicolor</i> (Gomont) Anagnostidis & Komárek	D6
35	<i>Symploca</i> sp.	D3
36	<i>Oscillatoria proboscidea</i> Gomont	D7
37	<i>Phormidium</i> sp. 1	D1
38	<i>Phormidium</i> sp. 2	D7
39	<i>Phormidium terebriforme</i> (C.Agardh ex Gomont) Anagnostidis & Komárek	D1, D4
40	<i>Phormidium interruptum</i> Kützing ex Forti	D6
41	<i>Komvophoron minutum</i> (Skuja) Anagnostidis & Komárek	D4
42	<i>Komvophoron skujae</i> Anagnostidis & Komárek	D3, D5
43	<i>Calothrix fusca</i> Bornet & Flahault	D6
Bacillariophyta		
44	<i>Cyclotella</i> sp.	D5
45	<i>Diploneis interrupta</i> (Kützing) Cleve	D6
46	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve	D6



**Fig. 4.** Principal component analysis (PCA) diagram of the algal flora, the physio-chemical parameters, and the sample sites of thermal springs (D1-D7) in Denizli. The graph shows the algal taxa codes (blue triangles), the sample sites (green circles), and the physio-chemical parameters, temperature, and concentration of different ions (red arrows) with the canonical axes. The algal taxa codes are given in Tab. 2. T – temperature, Ca – calcium, Mg – magnesium, Fe – ferrous, SO – sulphate, K – potassium, Na – sodium, Cl – chloride, and Li – lithium.

## Discussion

In this taxonomic study, the algal flora and physico-chemical parameters of seven thermal springs in Denizli were examined. We observed that these thermal springs were different in terms of their physio-chemical properties. As in similar studies in the literature (Ward and Castenholz 2000, Papke et al. 2003, Sompong et al. 2005), the relationship of species diversity in thermal springs and physio-chemical parameters and substrate structure was observed in this study. Accordingly, the correlation between the algal flora of the thermal springs and their physio-chemical parameters was examined statistically by PCA (Fig. 4).

The critical factor determining the distribution of algal flora in the thermal springs was temperature, with diatoms preferring relatively low temperatures. *Pinnularia microstauron* (Ehrenberg) Cleve, which was sampled from sites with low temperatures in this study (D5 and D6), has been reported from similarly low-temperature thermal springs (Fazlutdinova et al. 2020, Kaštovský and Komárek 2001). Kaštovský and Komárek (2001) defined the region below 36 °C in thermal waters as the mesothermophilic diatom region and stated that in this region, diatoms and cyanobacterial taxa form clusters together. Similar to Kaštovský and Komárek (2001), especially at low temperatures in D6, *P. microstauron* and *Diploneis interrupta* (Kützing) Cleve were observed to form mats with cyanobacteria taxa. The results of the PCA (Fig. 4) showed the negative correlation of these two diatom taxa with temperature, confirming diatom preference for lower temperatures in thermal springs.

Cyanobacteria taxa are common in thermal springs because they can survive at higher temperatures than other algae. *Jaaginema angustissimum* (West & G.S.West) Anagnostidis & Komárek is one of the common taxa of thermal and sulfate ( $\text{SO}_4^{2-}$ ) springs (Güner 1970, Komárek and Anagnostidis 2005, Arman et al. 2014, Öztürk Ulcay and Kurt 2017), and it was determined from D5, which is also characterized by a high  $\text{SO}_4^{2-}$  value in this study.

Interestingly, although *Gloeocapsa gelatinosa* Kützing was found to have a low correlation with  $\text{Cl}^-$  in this study (Fig. 4), Roy et al. (2015) reported that the taxon was associated with higher  $\text{Cl}^-$  by canonical correspondence analysis. In this study, *G. gelatinosa* and *Calothrix fusca* Bornet & Flahault were collected together from the same sample site which is consistent with the report of Lukavsky et al. (2011) that they formed a mat together.

The most common taxon in this study was *Spirulina subsalsa*, which did not have a significant correlation with the physio-chemical parameter results of the PCA. Altunöz et al. (2016) identified this taxon in the Pamukkale thermal spring and reported that this taxon did not show a distribution correlated with physio-chemical parameters, which is consistent with the present study. However, Roy et al. (2015) reported that the combined effect of pH,  $\text{K}^+$ , and  $\text{HCO}_3^-$  was effective in the distribution of *S. subsalsa*. As in the study of Kanellopoulos et al. (2016), *S. subsalsa* and *Phormidium terebriforme* (C.Agardh ex Gomont) Anagnostidis & Komárek were sampled together from the travertine substrates in this study (D1). The travertines are frequently colonized with Cyanobacteria mats, commonly composed of more than one taxon (Pentecost 2003, Kanellopoulos et al. 2016). Pentecost (2003) reported that travertines are often covered with a mixture of *Oscillatoria*, *Spirulina*, and *Phormidium* taxa. This result was also observed in this study; *Jaaginema geminatum*, *Spirulina major* Kützing ex Gomont, *S. subsalsa*, and *P. terebriforme* were collected from travertine substrates in D1, D2, D5, and D7. In addition, the D2 and D7 thermal springs had a high value of  $\text{Fe}^{2+}$  concentrations. Pierson and Parenteau (2000) reported well-developed cyanobacterial mats in high  $\text{Fe}^{2+}$  concentrations. As can be seen from the PCA (Fig. 4), *Planktolyngbya contorta* (Lemmermann) Anagnostidis & Komárek and *Oscillatoria proboscidea* Gomont were identified from D7 with the highest  $\text{Fe}^{2+}$  concentration. Also, common taxa of D2 and D7, with high  $\text{Fe}^{2+}$  concentrations, were *Pseudanabaena mucicola* and *S. subsalsa*, and the distribution of these species in this study overlaps with the related data from the literature. However, *S. subsalsa* does not appear to be associated with  $\text{Fe}^+$  concentrations (Fig. 4). The possible reason may be the ecological tolerance of *S. subsalsa*.

## Conclusions

This study enlarged the worldwide knowledge of algal flora inhabiting thermal springs, which helps to improve the ecological data on the physio-chemical parameter preferences of Cyanobacteria and Bacillariophyta taxa and con-

tributes to the understanding of the effects of physio-chemical parameters on the diversity of these ecosystems. Our findings also confirm that algal flora diversity is directly affected by temperature, followed by potassium and chloride ions in the water of thermal springs.

Further ecological, physiological, and biotechnology studies are required for an understanding of the potential usability of these thermophilic species of algae. For all these reasons, determining the species diversity in thermal springs can also underpin new economic and industrial uses.

## Acknowledgments

The authors thank the editors and the anonymous reviewers for their interesting and valuable comments, careful revisions made in the text, and constructive criticism.

## Author contribution statement

S.Ö. collected algae samples, measured thermal springs, and did the statistical analysis. S.Ö. and O.K. identified the algae samples and prepared the manuscript.

## References

- Altunöz, M., Obali, O., Atici, T., Arru, L., 2016: Seasonal monitoring of algal flora in the Pamukkale travertines and thermal springs (Denizli/Turkey). *Biodiversity Conservation* 9(1), 116–127.
- Arman, M., Riahi, H., Yousefzadi, M., Sonboli, A., 2014: Floristic study on cyanophyta of three hot springs of Hormozgan province, Iran. *The Iranian Journal of Botany* 20(2), 240–247. <https://doi.org/10.22092/ijb.2014.11029>
- Aysel, V., Çelik, A., Yayıntaş, A., Şipal-Gezerler, U., 1992: Zonguldak-Ilıksu Kaplıcası alg florası [Algae flora of Zonguldak-Ilıksu Thermal Spring]. *Ege Üniversitesi Su Ürünleri Dergisi* 9, 72–82.
- Cantonati, M., Kelly, M. G., Lange-Bertalot, H., 2017: Freshwater benthic diatoms of Central Europe: over 800 common species used in ecological assessment. Koeltz Botanical Books, Schmitten-Oberreifenberg, Germany.
- Fazlutdinova, A., Gabidullin, Y., Allaguvatova, R., Gaysina, L., 2020: Diatoms in Kamchatka's hot spring soils. *Diversity* 12(11), 435. <https://doi.org/10.3390/d12110435>
- Guiry, M. D., Guiry, G. M., 2023: Algae Base, world wide web electronic publication, National University of Ireland, Galway. Retrieved on May 23, 2023, from <https://www.algaebase.org>.
- Güner, H., 1966: Pamukkale termal suyunun mikroflorası [Microflora of Pamukkale thermal water]. *Ege Üniversitesi Fen Fakültesi İlmî Raporlar Serisi* 3.
- Güner, H., 1970: Ege Bölgesi kaplıca ve maden sularının alg vegetasyonu ile ilgili inceleme [Investigation on algae vegetation of Aegean Region thermal springs and mineral waters] *Ege Üniversitesi Fen Fakültesi İlmî Raporlar Serisi* 99.
- Kanellopoulos, C., Lamprinou, V., Mitropoulos, P., Voudouris, P., 2016: Thermogenic travertine deposits in Thermopylae hot springs (Greece) in association with cyanobacterial microflora. *Carbonates and Evaporites* 31, 239–248. <https://doi.org/10.1007/s13146-015-0255-4>
- Kaštovský, J., Komárek, J., 2001: Phototrophic microvegetation of thermal springs in Karlovy Vary, Czech Republic. In: Elster, J., Seckbach, J., Vincent, W.F., Lhotský, O. (eds.), *Algae and extreme environments*. Nova Hedwigia 123, 107–119.
- Komárek, J., 2013: Cyanoprokaryota, 3. Heterocystous Genera. In: Büdel, B., Gärtner, G., Krienitz, L., Schagerl, M. (ed.), *Süßwasserflora von Mitteleuropa*, Vol 19/3. Berlin, Heidelberg, Springer Spektrum.
- Komárek, J., Anagnostidis, K., 1999: Cyanoprokaryota, 1. Chroococcales. In: Ettl, H., Gärtner, G., Heynig, H., Mollenhauer, D. (eds.), *Süßwasserflora von Mitteleuropa* Vol 19/1. Gustav Fischer Verlag, Stuttgart- New York.
- Komárek, J., Anagnostidis, K., 2005: Cyanoprokaryota, 2. Oscillatoriales. In: Büdel, B., Gärtner, Krienitz, L., Schlagerl, M. (ed.), *Süßwasserflora von Mitteleuropa*, Vol 19/2. Elsevier Gmb H, München.
- Kozak, M., 2020: Denizli ili jeotermal enerji kaynakları ve kullanım alanlarının araştırılması [Investigation of geothermal energy resources and usage areas in Denizli province]. *Journal of Yekarum* 5, 1–11.
- Lukavsky, J., Furnadzhieva, S., Pilarski, P., 2011: Cyanobacteria of the thermal spring at Pancharevo, Sofia, Bulgaria. *Acta Botanica Croatica* 70(2), 191–208. <https://hrcak.srce.hr/file/107339>
- Öztürk, S., 2020: Notes on the thermal habitat: Thirteen new records for the freshwater algal flora of Turkey. *Indian Journal of Geo-Marine Sciences* 49(10), 1661–1668.
- Öztürk Ulçay, S., Kurt, O., 2017: Algae flora of Germencik-Alangüllü (Aydın, Turkey) thermal water. *Celal Bayar University Journal of Science* 13(3), 601–608. <https://doi.org/10.18466/cbayarfbe.339279>
- Öztürk Ulçay, S., Kurt, O., Öztürk, M., Güner, H., 2017: Algae microflora of Pamukkale thermal water (UNESCO, 485) 45 years ago and now. *Fresenius Environmental Bulletin* 26(6), 4153–4157.
- Öztürk Ulçay, S., Öztürk, M., Kurt, O., Taşkın, E., Öztürk, M., 2007: Dikili ilçesi (İzmir) kaplıcalarında yayılış gösteren termal cyanophyceae (mavi-yeşil alg) türleri [Thermal cyanophyceae (blue-green algae) species distributed in the thermal springs of Dikili district (İzmir)]. *Türk Sucul Yaşam Dergisi* 5(8), 371–378.
- Papke, R. T., Ramsing, N. B., Bateson, M. M., Ward, D. M., 2003: Geographical isolation in hot spring cyanobacteria. *Environmental Microbiology* 5(8), 650–659. <https://doi.org/10.1046/j.1462-2920.2003.00460.x>
- Pentecost, A., 2003: Cyanobacteria associated with hot spring travertines. *The Canadian Journal of Earth Sciences* 40(11), 1447–1457. <https://doi.org/10.1139/e03-075>
- Pentecost, A., Bayari, S., Yesertener, C., 1997: Phototrophic microorganisms of the Pamukkale travertine, Turkey: Their distribution and influence on travertine deposition. *Geomicrobiology Journal* 14(4), 269–283. <https://doi.org/10.1080/01490459709378052>
- Pierson, B. K., Parenteau, M. N., 2000: Phototrophs in high iron microbial mats: microstructure of mats in iron-depositing hot springs. *FEMS Microbiology Ecology* 32(3), 181–196.
- Regel, C., Skuja, H., 1937: Süßwasseralgen aus Griechenland und Kleinasien. *Hedwigia* 77, 15–70.
- Roy, S., Bhattacharya, S., Debnath, M., Ray, S., 2015: Diversity of cyanobacterial flora of Bakreswar geothermal spring, West Bengal, India-II. *Algological Studies* 147, 29–44. <https://doi.org/10.1127/1864-1318/2014/0178>
- Sompong, U., Hawkins, P. R., Besley, C., Peerapornpisal, Y., 2005: The distribution of cyanobacteria across physical and chemical gradients in hot springs in northern Thailand. *FEMS Microbiology Ecology* 52(3), 365–376. <https://doi.org/10.1016/j.femsec.2004.12.007>



- Ter Braak, C. J. F., Šmilauer, P., 2012: Canoco reference manual and user's guide: software for ordination, version 5.0. Micro-computer Power, Ithaca, NY, USA.
- UNESCO, 2022: World Heritage Centre, world wide web electronic publication. Retrieved on April 23, 2022, from <http://whc.unesco.org/en/list/485>.
- Ward, D. M., Castenholz, R. W., 2000: Cyanobacteria in geothermal habitats. In: Whitton, B.A., Potts Dordecht, M. (eds.), The ecology of Cyanobacteria. Kulwer Academic Publishers.
- Yaman, D., Özgür, N., 2005: Kızıldere ve çevresi jeotermal alanların hidrojeolojik, hidrojeokimyasal ve izotop jeokimyasal özellikleri [Hydrogeological, hydrogeochemical and isotope geochemical properties of Kızıldere and its surrounding geothermal fields]. II. Ulusal Hidrolojide İzotop Teknikleri Sempozyumu, Ekim 2005, İzmir, 26–30.

# Does palynotaxonomy contribute to the systematics of the genus? The section *Multicaulia* of the genus *Hedysarum* (Fabaceae) example in Türkiye

Burcu Yılmaz Çitak

University of Selçuk, Faculty of Science, Department of Biology, 42130 Konya, Türkiye

**Abstract** – The purpose of the current work was to assess the systematic and taxonomic significance of the pollen morphological characteristics of the *Multicaulia* section of *Hedysarum* species found throughout Türkiye using scanning electron microscopy (SEM) and light microscopy (LM) techniques. For this reason, seven different species currently classified in the *Multicaulia* section were collected from various localities. The pollen grains were nonacetylated, prepared and directly measured according to the Wodehouse method. Pollen grains were found to be subprolate in *H. nitidum* but prolate in others and the exine sculpturing pattern was determined to be microreticulate-perforate in six taxa while only *H. varium* subsp. *syriacum* had reticulate-perforate sculpture. The minimum exine thickness was measured at 0.96 µm in *H. varium* subsp. *varium*, while among the investigated species, *H. nitidum* had an exine measurement of 1.38 µm. In this study, multivariate analysis was used to investigate nine qualitative and quantitative features. Data cluster analysis revealed that the pollen characteristics were important in helping to distinguish the different species from one another. The results demonstrated that the genus *Hedysarum* of the section *Multicaulia* was divided into two major groups based on variations in pollen dimensions. The usefulness of certain pollen morphological traits was assessed, and the congruency of the palynomorphological investigations was examined. The findings demonstrated the significance of pollen micromorphology in precisely identifying and categorizing the *Multicaulia* section.

**Keywords:** *Hedysarum*, Leguminosae, palynology, taxonomy, sweet vetch, systematic

## Introduction

It is a broadly acknowledged theory that the fields around the large Tethys Sea (which covered part of the present Mediterranean zone during the Tertiary) became a center of speciation for many plant families after the Alpine uplift and Oligocene climatic changes during the Pleistocene and Holocene. Only several legume tribes such as *Hedysareae*, *Genisteae*, *Viciae*, *Loteae*, and *Trifoliae*, were able to endure the severe climatic conditions, surviving and multiplying under extreme pressure (summer drought, winter frost). The Fabaceae family of flowering plants, which includes roughly 19,500 species divided into 751 genera, is regarded as the third largest flowering plant family. This enormous family has demonstrated a high degree of species diversity and evolutionary success in a variety of global ecosystems, including stony grasslands, deserts, seashores, alpine and arctic meadows (Choi and

Ohashi 2003, Legume Phylogeny Working Group 2023). The Mediterranean and Irano-Turanic phytogeographical regions have some specific genera such as *Onobrychis*, *Hedysarum*, *Medicago*, *Astragalus*, *Anthyllis*, *Trigonella*, *Coronilla*, *Hippocrepis*, *Ononis*, *Genista*, etc. (Lesins and Lesins 1979, Polhill 1981). Two areas are distinctly bipolar for this speciation: the eastern Mediterranean and Irano-Turanic zone (around the Aegean, Black, and Caspian seas), and the western Mediterranean or Ibero-Maghrebi (around the Tyrrhenian and Alboran seas). Within the genus *Hedysarum*, circum-Mediterranean, mostly eastern Mediterranean, species are outstanding (among the 21 taxa recognized for Europe by Chrtková-Zertova, 1968, 13 are eastern Mediterranean). Among the 45 taxa recognized for the circum-Mediterranean areas by Greuter et al. (1989), 33 are present in the eastern Mediterranean.

In Türkiye alone, 22 taxa (one of which is suspect) of *Hedysarum* were recognized by Hedge (1979), and with the addition of the new taxa, the number of taxa in the genus will reach 24 (Başköse et al. 2018, Aytaç et al. 2020, Hamzaoğlu and Koç 2020, Kandemir et al. 2023). Hedge (1979) separated *Hedysarum* into five sections (*Hedysarum*, *Obscura*, *Multicaulia*, *Subacaulia*, and *Crinifera*) in the Flora of Türkiye. Of these, 11 are endemic to Türkiye, and the endemism ratio of the genus is 52.3%. The classification of sections of *Hedysarum* is also quite problematic. Until recently, it has included four sections: *Hedysarum*, *Membranacea* B. Fedtsch., *Stracheya* (Benth.) B.H. Choi & H. Ohashi, and *Multicaulia* (Boiss.) B. Fedtsch. However, section *Multicaulia* consists of three subsections: *Multicaulia* B.H. Choi & H. Ohashi, *Crinifera* (Boiss.) B.H. Choi & H. Ohashi, and *Subacaulia* (Boiss.) B.H. Choi & H. Ohashi.

Characters such as the size, shape, and number of leaflets, indumentum, flower color, size, shape, and proportions of the standard, wings and keel, and shape and indumentum of the lomentum are extremely variable among the species of the *Hedysarum* genus (Hedge 1979). *Hedysarum varium* is the most common and polymorphic species of the genus in Türkiye. Hedge (1979) also noted that *H. varium* may be extended by hybridization or introgression with *H. pestalozzae*, *H. syriacum*, and *H. nitidum*. The differences between these four species are unsatisfactory, and further work is needed before their interrelationships and taxonomy can be placed on sounder foundations.

The palynomorphology of the many Fabaceae taxa has taxonomic significance (Erdtman 1969, Ohashi 1971, Perveen and Qaiser 1998, Pavlova and Manova 2000, Pınar et al. 2000, Avcı et al. 2013, Çeter et al. 2013, Bagheri et al. 2019). Three apertures, reticulate or supracreticulate ornamentation of exine, oblate-spheroidal or prolate pollen grains were given in those studies. Additionally, the micromorphological (pollen, fruit, and seed) characteristics can contribute to the discrimination of *Hedysarum* species, especially the pollen grains and muri size. However, there has been no taxonomical study conducted on the pollen structures of the subsection *Multicaulia* in Türkiye (Ohashi 1971, Polhill 1981, Faegri et al. 1989, Moore et al. 1991, Choi and Ohashi 1996, Civelek et al. 1999, Pavlova and Manova 2000, Sa et al. 2010, Ghanavati and Amirabadizadeh 2012, Dural and Citak 2015).

In order to enhance the pollen morphological research into the genus *Hedysarum*, the current study was conducted to describe and evaluate in detail, for the first time, the palynomorphological properties of seven taxa distributed in Türkiye using light microscopy (LM) and scanning electron microscopy (SEM). The objectives of this work were to: 1) identify and investigate the features of the pollen grains of species belonging to the *Multicaulia* subsection; and 2) use numerical analyses to clarify the systematic significance of the palynological traits.

## Materials and methods

### Plant material

The specimens of the section *Multicaulia* were collected from various localities, as listed in Table 1, and stored in the herbarium of the Department of Biology of Selçuk University (KNYA). The taxonomical description of the species followed that of Hedge (1979).

### Palynological analysis

Both LM and SEM were used to examine pollen grains of the genus *Hedysarum*. For palynological analysis, pollen samples of specimens were obtained from herbarium materials. To observe and calculate the characteristics of pollen grains, the Wodehouse technique was used (Wodehouse 1935). In this unique and easy technique, pollen grains were separated from the anthers and were stained with glycerin-jelly and covered with coverslips. A Leica DM 1000 light microscope with a Canon 450D camera (Ota City, Tokyo, Japan) and software from the Kameram 21 program (Argenit, Istanbul, Türkiye) was used to measure and observe fifty pollen grains per specimen. The equatorial diameter (E), polar axis (P), colpus length (Clg), colpus width (Clt), apocolpium (t), thickness of the exine and intine, and lumina width (LW) were examined and the P/E ratios were calculated. The mean, standard deviation (SD), minimum–maximum range, and mean values of these pollen characteristics for the taxa under study were given.

The unacetolyzed pollen grains were transferred directly onto aluminum stubs, covered with gold using a Cressington Auto 108 sputter coater (Cressington Scientific Instruments, Watford, Hertfordshire, UK) and photographed

**Tab. 1.** Localities of specimens of *Hedysarum* from the section *Multicaulia* collected from various localities in Türkiye and stored in KNYA herbarium.

Taxa	Locality	Herbarium no
<i>H. laxum</i> (DC.) Spreng	C5 Adana: Tufanbeyli, 1300 m.	B. Çitak-366
<i>H. leucocladum</i> Boiss.	B5 Nevşehir: Zelve, 1100 m.	B. Çitak-150
<i>H. huetii</i> Boiss.	A8 Erzurum: Tortum, 1300 m.	B. Çitak-377
<i>H. nitidum</i> Willd.	A8 Erzincan: Refahiye, 1500 m.	B. Çitak-369
<i>H. pestalozzae</i> Boiss.	C4 Karaman: Ayrancı, 1300 m.	B. Çitak-160
<i>H. varium</i> Willd. subsp. <i>varium</i>	C4 Konya: Konya-Beyşehir road, 1300 m.	B. Çitak-161
	B6 Sivas: Sivas-Erzincan road, 1500 m.	B. Çitak-367
<i>H. varium</i> subsp. <i>syriacum</i> (Boiss.) C.C. Towns.	B5 Aksaray: Aksaray-Nevşehir road, 1150 m.	B. Çitak-151

using a Zeiss Evo Ls10 scanning electron microscope (SEM) (Carl Zeiss NTS GmbH, Oberkochen, Germany) for the SEM analyses (Dural and Citak 2015).

The pollen terminologies of Punt et al. (2007), and Halbritter et al. (2018) were followed.

### Statistical analysis

The determined qualitative and quantitative characters were scored for numerical analysis. Nine pollen characters were used to evaluate *Hedysarum* taxonomic relationships. These characters are equatorial and polar axes, exine, intine, colpus length, colpus width, apocolpium, sculpture and lumen width. Next, the determined qualitative and quantitative characters were turned into a data matrix. For the pollen characters of the seven taxa, the coefficients of correlation were detected and compiled using the clustering analysis method (unweighted pair group method with arithmetic mean (UPGMA), dissimilarity, standardized variables). Using nine characters and seven taxa, a primary matrix was established for the multivariate analysis. The Gower general coefficient similarity (Gower 1971), which can be utilized directly to a variety of character types (binary, qualitative, and quantitative characters), provided a basis for the clustering analysis. The UPGMA method was selected due to its widespread usage, congruence with the classification derived by traditional techniques, and visible accuracy in generating a reflection similarity matrix, as demonstrated by the co-phenetic correlation coefficient of

Sokal and Rohlf (1962) and symmetrical hierarchical structure (Sokal and Rohlf 1962, McNeill 1979). The covariance matrix was created using non-standardized, untransformed, and centered data (Citak 2019). All calculations were performed via MVSP 3.22 software (Kovach Computing Services, Anglesey, Wales, Kovach 2013).

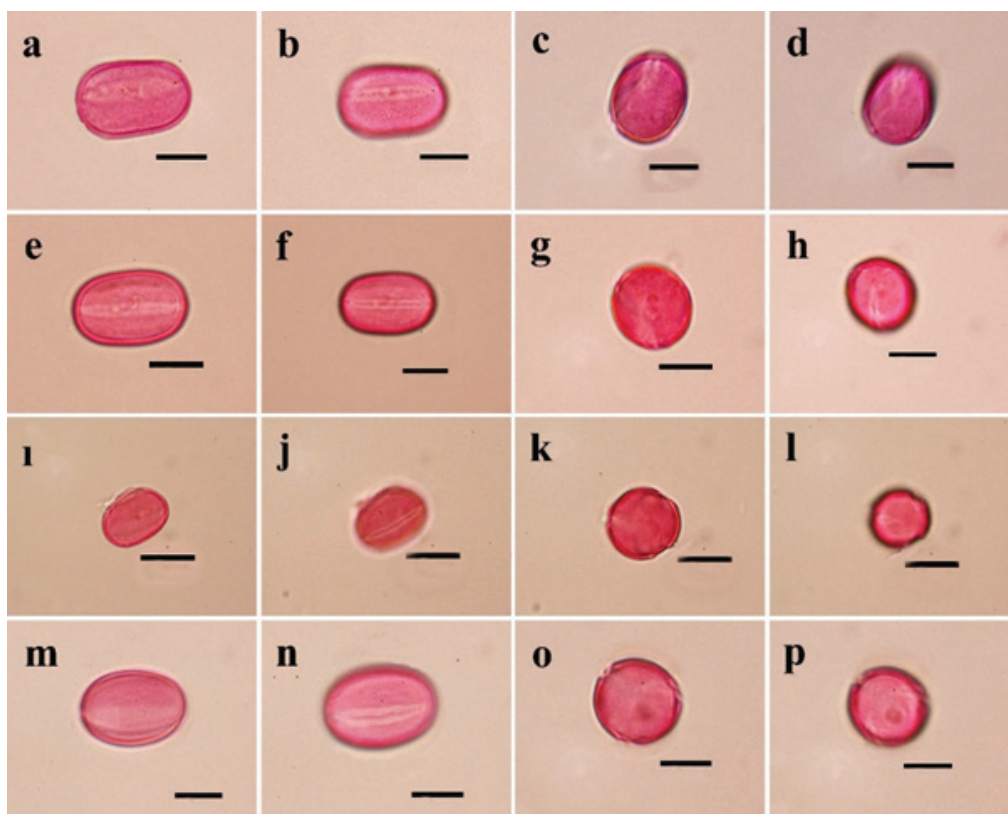
## Results

### Pollen morphology

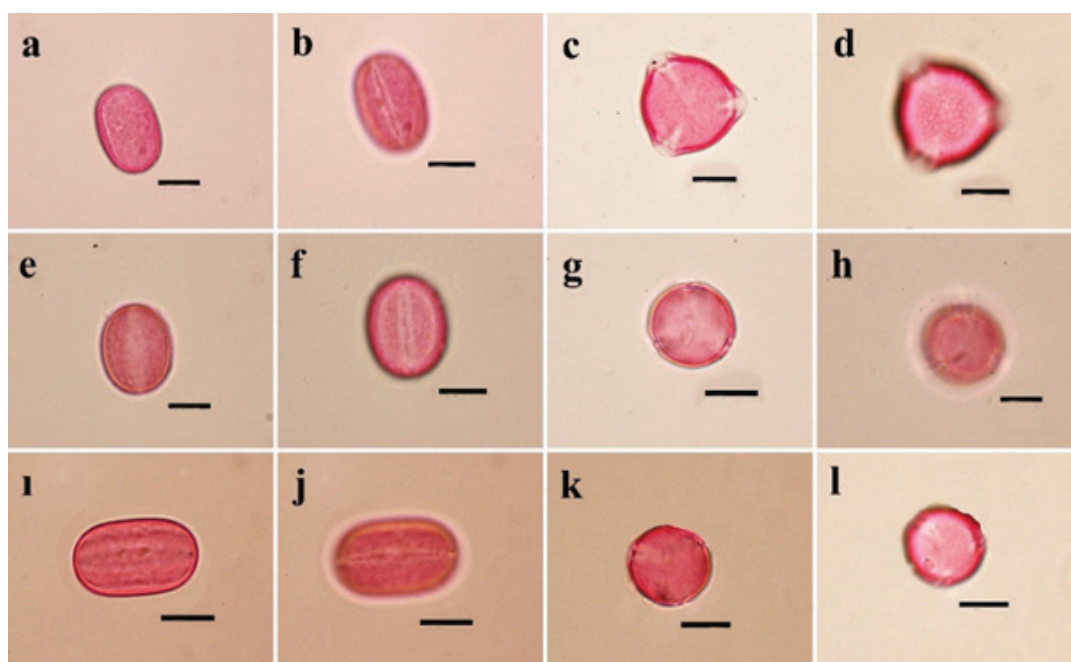
The pollen grains of the taxa of the subsection *Multicaulia* were determined as monad, radially symmetrical and isopolar. The shapes of the pollen grains were elliptic and not compressed at the poles in equatorial view or orbicular in polar view (Fig. 1, Fig. 2).

Quantitative characteristics of pollen grains from the plant samples are summarized in Table 2.

The general pollen shape in this study was determined as prolate; subprolate pollen shape was observed only in *H. nitidum*. The highest average pollen grain diameter on polar axis was observed in *H. laxum* (22.85  $\mu\text{m}$ ), whereas the lowest diameter was observed in *H. huetii* (20.53  $\mu\text{m}$ ). Similarly, the maximum pollen grain diameter on the equatorial axis was observed in *H. nitidum* (16.23  $\mu\text{m}$ ), while the minimum diameter was examined in *H. laxum* (12.68  $\mu\text{m}$ ). The size of the pollen grains was calculated based on the P/E ratio. The highest P/E value was observed in *H. laxum* (1.80), while the lowest value was observed in *H. nitidum* (1.33).



**Fig. 1.** Pollen microphotographs of species of the *Hedysarum* genus of *Multicaulia*: **a-d.** *H. laxum*, **e-h.** *H. leuocladum*, **i-l.** *H. huetii*, **m-p.** *H. nitidum*. Scale bar: 10  $\mu\text{m}$ .



**Fig. 2.** Pollen microphotographs of species of *Hedysarum* genus of *Multicaulia* **a-d.** *H. pestalozzae*, **e-h.** *H. varium* subsp. *varium* **i-l.** *H. varium* subsp. *syriacum*. Scale bar: 10  $\mu$ m.

All of the studied members of the subsection *Multicaulia* had tricolpate apertures. The investigated species had different aperture sizes, with the maximum average colpi length observed in *H. pestalozzae* (19.84  $\mu$ m), while the minimum was found in *H. huetii* (16.83  $\mu$ m). In terms of the width, the maximum colpulus width was found in *H. nitidum* (3.93  $\mu$ m), while the minimum width was observed

had microreticulate-perforate exine sculpturing, except for *H. varium* subsp. *varium*, which had reticulate-perforate exine sculpturing (Fig. 3). The exine thickness ranged from a maximum of 1.38  $\mu$ m in *H. nitidum* to a minimum of 0.96  $\mu$ m in *H. varium* subsp. *varium*. The lumina width also varied among the species, and ranged from a maximum of 1.10  $\mu$ m in *H. varium* subsp. *varium* to a minimum of

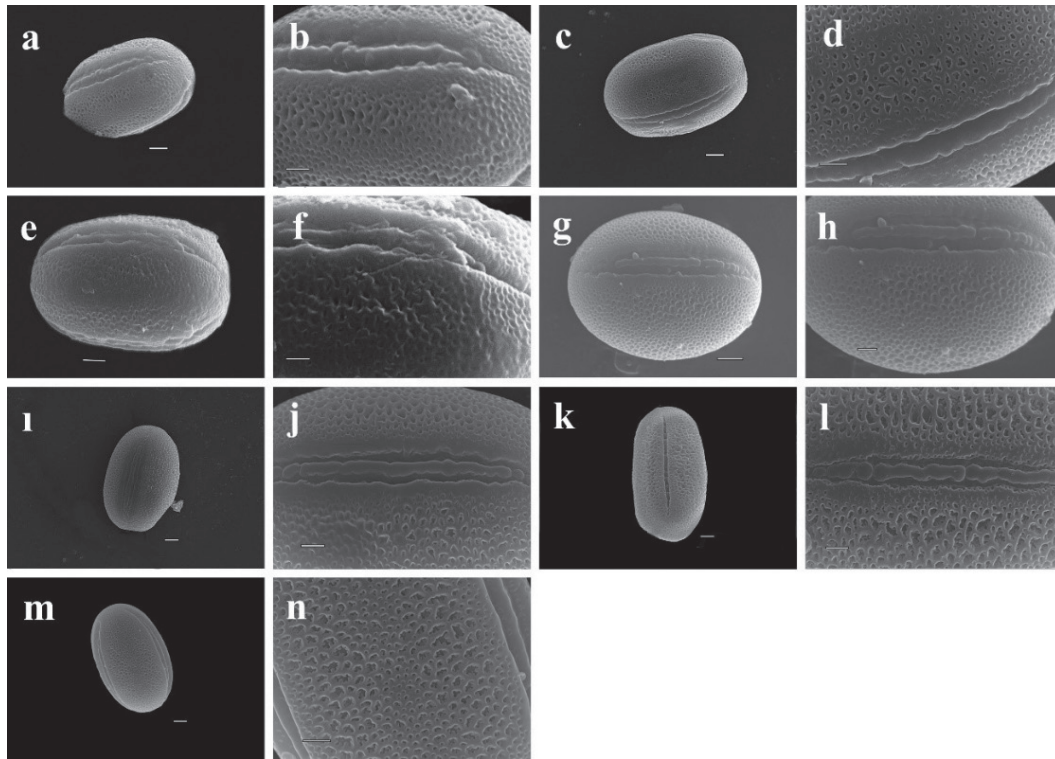
**Tab. 2.** The palynomorphological characters examined in *Multicaulia* subsection (values in  $\mu$ m, except for P/E). PS: pollen shape, P: prolate, SP: subprolate, E: exine, I: intine, Clg: colpulus length, Clt: colpulus width, t: apocolpium, Sc: sculpturing, MP: microreticulate-perforate, RP: reticulate-perforate, LW: lumina width. Fifty pollen grains per specimen were analyzed. \* indicates endemic taxa and \*\* indicates characters used in statistical analysis.

Taxa	Equatorial axes (E)**		Polar axes (P)**		P/E	PS	E**	I**	Clg**	Clt**	T**	Sc**	LW**
	Min – Max	Mean $\pm$ SD	Min – Max	Mean $\pm$ SD									
* <i>H. laxum</i>	10.28 – 15.34	12.68 $\pm$ 1.81	17.67 – 25.26	22.85 $\pm$ 2.37	1.80	P	1.3	0.58	19.23	3.14	3.10	MP	0.57
<i>H. leuocladum</i>	13.26 – 15.76	14.31 $\pm$ 0.78	19 – 22.38	21.03 $\pm$ 1.09	1.46	P	1.27	0.62	19.55	2.97	2.97	MP	0.53
<i>H. huetii</i>	13.55 – 15.28	14.7 $\pm$ 0.7	19.2 – 21.78	20.53 $\pm$ 1.09	1.39	P	1.06	0.54	16.83	2.99	4.39	MP	0.59
* <i>H. nitidum</i>	15.64 – 16.91	16.23 $\pm$ 0.47	19.65 – 23.77	21.67 $\pm$ 1.13	1.33	SP	1.38	0.63	19.71	3.93	3.25	MP	0.74
* <i>H. pestalozzae</i>	9.8 – 16.24	13.6 $\pm$ 2.04	16.37 – 22.65	20.96 $\pm$ 1.54	1.54	P	1.00	0.74	19.84	3.45	3.19	MP	0.50
<i>H. varium</i> subsp. <i>varium</i>	10.41 – 18.13	15.76 $\pm$ 2.44	19.64 – 23.79	21.57 $\pm$ 1.13	1.36	P	0.96	0.66	19.69	3.6	2.74	RP	1.10
<i>H. varium</i> subsp. <i>syriacum</i>	14 – 15.44	14.76 $\pm$ 0.44	21.03 – 23.67	22.34 $\pm$ 0.88	1.51	P	1.16	0.55	19.17	3.82	3.61	MP	0.62

in *H. leuocladum* (2.97  $\mu$ m). An operculum with psilate ornamentation on the colpulus was determined in all pollen grains.

The exine ornamentation was the most distinguishing character of the species. In current study, all of the species

had microreticulate-perforate or reticulate-perforate in equatorial view; however, only perforate ornamentation was observed in polar view. The walls of the muri were sinuous in the examined species.



**Fig. 3.** Electron microphotographs of species of *Hedysarum* genus of *Multicaulia*. **a-b.** *H. laxum*, **c-d.** *H. leucocladum*, **e-f.** *H. huetii*, **g-h.** *H. nitidum*, **i-j.** *H. pestalozzae*, **k-l.** *H. varium* subsp. *varium*, **m-n.** *H. varium* subsp. *syriacum*. Scale bar (**a, c, e, g, i, k, m**): 2  $\mu$ m; scale bar (**b, d, f, h, j, l, n**): 1  $\mu$ m.

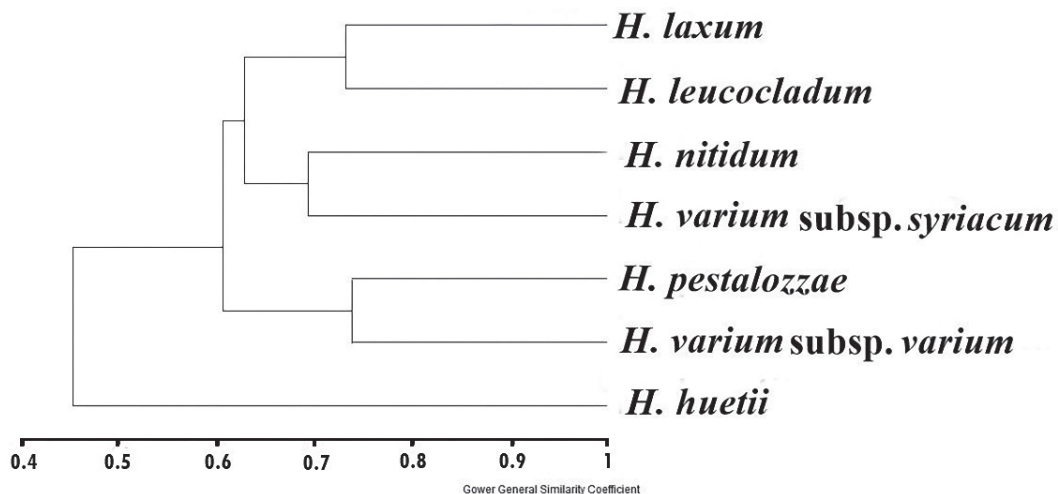
**Numerical analysis of the pollen characters traits**

A dendrogram produced with the use of the UPGMA in the cluster analysis, taking into account the ten palynological variables (Tab. 2) found in the section *Multicaulia*, is shown in Fig. 4. Similarities among the investigated taxa were reflected in the dendrogram, which revealed two main groups: group A (with 45% similarity) contained only *H. huetii*, which was a distinct species, and group B (with 60% similarity) comprised the remaining 6 taxa. Group B comprised two primary clusters, further denoted as clusters C and D. Two species (*H. pestalozzae* and *H. varium* subsp.

*varium*, with a 73% similarity) made up cluster C. Four taxa with two subgroups, clusters D1 and D2, were included in cluster D. Cluster D1 contained *H. laxum* and *H. leucocladum*. Cluster D2 also included 2 species: *H. nitidum* and *H. varium* subsp. *syriacum*.

**Discussion**

The present study examined the usefulness of using the pollen morphological characteristics of species of the section *Multicaulia* of the genus *Hedysarum* in Türkiye. Using



**Fig 4.** The derived dendrogram based on the palynological characters showing the similarity distance of *Hedysarum* species.

these pollen morphological characters, delimitation of taxa was determined.

The findings of current investigation show that the pollen dimensions (E and P/E ratio), the thickness of the exine and intine, the shape of the polar and equatorial views, and the aperture type of the Türkiyeli representatives of *Hedysarum* were comparatively homogeneous (Tab. 2, Fig. 1). Ohashi (1971), Ferguson and Skvarla (1981), Faegri and Iversen (1989), Moore et al. (1991), Pavlova and Manova (2000), and Ghanavati and Amirabadizadeh (2012) confirmed that the pollen morphological characteristics of the different sections of *Hedysarum* are essentially the same. Polar and equatorial axes were useful pollen characters to group species with the aid of numerical analysis as mentioned before in *Astragalus* (Bagheri et al. 2019). Another study declared that *Onobrychis* genus had oblate-spheroidal pollen shape (Avcı et al. 2013).

All of the species investigated in this research had trilcolpate-type pollen grains with microreticulate-perforate exine ornamentation. Only *H. varium* subsp. *varium* was different from others, with reticulate-perforate exine. Reticulate and finely reticulate exine ornamentation were reported previously in the genus *Hedysarum* by Civelek et al. (1999), Pavlova and Manova (2000), and Dural and Citak (2015). The sculpture of exines were determined with three types counted reticulate, suprareticulate and microreticulate in close relatives of *Hedysarum* genus, *Astragalus* and *Onobrychis* (Avcı et al. 2013, Bagheri et al. 2019). Perveen and Qaiser (1998) reported that the ornamentation, and in particular, the shape and size of the muri and lumina, was an important character for Fabaceae (Pavlova and Manova 2000, Ghanavati and Amirabadizadeh 2012, Dural and Citak 2015). The colpus membrane has been identified as being covered with large and small sculptural elements by different researchers in the genus *Hedysarum* (Pavlova and Manova 2000, Dural and Citak, 2015). Moreover, in this study the same sculptural elements were observed in the subsection *Multicaulia* as described for the operculum with psilate sculpture.

The UPGMA dendrogram derived based on the palynological characters discriminated the species of the subsection *Multicaulia*. The positions of the species and the relationships of the subsection *Multicaulia* of the genus *Hedysarum* in the cluster were found to be in agreement with the previous classification made on a large scale (Hedge 1979, Hamzaoğlu and Koç 2020). *H. huetii*, which is a very distinct species, can be easily separated from the others by its fleshy and ovate-orbicular leaflets; hence, it was alone in a different subclade. It was very surprising that *H. pestalozzae* and *H. varium* subsp. *varium* were in the same clade, because of their broad morphological differences, particularly regarding the color of the corolla. In addition, the positions of *H. varium* subsp. *syriacum* and *H. nitidum* in the same clade were surprising due to their morphological differences. This situation raised the question of whether or not new taxonomic regulations for these taxa are required. The last two species, *H. laxum* and *H. leuocladum*, which

are local and endemic, were in the same subclade. The distribution of the species of the subsection *Multicaulia* has raised the question of whether or not the relationships of the taxa should be reviewed one more time based on their pollen characteristics. Furthermore, different lumina dimensions, lumina types, and densities among taxa may be associated to their multiple germination requirements or dispersal strategies, which may have developed as a result of an evolutionary adaptation independent of various habitat types or microclimatic regions, particularly for endemics. More data are needed to solve the systematic problems of this section, especially regarding the phylogenetical relationships of the taxa.

## Conclusion

To sum up, an in-depth investigation of the pollen characters was highly helpful in classifying the studied taxa. In future work, we propose that the systematic problems still unresolved in *Hedysarum* taxa should perhaps be solved by combining morphological with anatomical, micromorphological, karyological, and additional molecular data.

## Acknowledgments

The author thanks Dr. Hüseyin Dural for helping to identify the plant samples of the species of *H. pestalozzae* and *H. varium*.

## References

- Avcı, S., Sancak, C., Can, A., Acar, A., Pınar, N. M., 2013: Pollen morphology of the genus *Onobrychis* (Fabaceae) in Turkey. *Turkish Journal of Botany* 37(4), 669–681. <https://doi.org/10.3906/bot-1207-52>
- Aytaç, Z., İğci, B. K., Körüklü, T. S., 2020: *Hedysarum nallihanse* (Fabaceae): A new species from Turkey. *Phytotaxa* 471(3), 276–282. <https://doi.org/10.11646/phytotaxa.471.3.9>
- Bagheri, A., Roofgar, A. A., Abbasi, S., Maassoumi, A. A., Rutten, T., Blatner, F. R., 2019: Pollen morphology of *Astragalus* section *Hymenostegis* (Fabaceae) and evaluation of its systematic implications. *Grana* 58(5), 328–336. <https://doi.org/10.1080/00173134.2019.1621931>
- Başköse, İ., Yaprak, A. E., Akyıldırım, B., 2018: *Hedysarum kentenoglui* (Fabaceae-Hedysareae), a new species from southern Turkey. *Phytotaxa* 357(4), 291–297. <https://doi.org/10.11646/phytotaxa.357.4.5>
- Çeter, T., Erkul, S. K., Aytaç, Z., Başer, B., 2013: Pollen morphology of the genus *Oxytropis* Dc. in Turkey. *Bangladesh Journal of Botany* 42(1), 167–174. <https://doi.org/10.3329/bjb.v42i1.15908>
- Choi, B., Ohashi, H., 1996: Pollen morphology and taxonomy of *Hedysarum* and its related genera of the tribe Hedysareae (Leguminosae: Papilionoideae). *Journal of Japanese Botany* 71, 191–213.
- Choi, B., Ohashi, H., 2003: Generic criteria and an infrageneric system for *Hedysarum* and related genera (Papilionoideae-Leguminosae). *Taxon* 52, 567–576.
- Citak, B. Y., 2019: A palynological survey of the genus *Iberis* (Brassicaceae), known as candytufts, in Turkey. *Phytotaxa* 397, 213–224.

- Civelek, S., Yaman, A., Sahin, A., Gur, N., 1999: The investigation in point of chromosome number, morphological, anatomical and pollen characteristics of *Hedysarum aucheri* Boiss. Journal of Science and Engineering of Firat University 11, 77–88.
- Dural, H., Citak, B. Y., 2015: Morphology and anatomy of *Hedysarum pannosum* (Boiss.) Boiss. (Fabaceae). Acta Botanica Croatica 74(1), 19–29. <https://doi.org/10.1515/botcro-2015-0009>
- Erdtman, G., 1969: Pollen Morphology and Plant Taxonomy. New York, Hafner Publishing.
- Faegri, K., Iversen, J., 1989: Textbook of pollen analysis. John Wiley & Sons, Chichester.
- Ferguson, J. K., Skvarla, J. J., 1981: The pollen morphology of the subfamily Papilionoideae (Leguminosae). In: Polhill, R.M., Raven, P.H. (eds.), Advances in Legume Systematics. 859–896. Kew Royal Botanic Gardens, London.
- Ghanavati, F., Amirabadizadeh, H., 2012: Pollen grain morphology in Iranian Hedysareae (Fabaceae). Crop Breeding Journal 2(1), 25–33. <http://dx.doi.org/10.22092/cbj.2012.100413>
- Gower, J. C., 1971: A general coefficient of similarity and some of its properties. Biometrics 27, 857–871. <https://doi.org/10.2307/2528823>
- Greuter, W., Burdet, H. M., Long, G., 1989: Med-Checklist. A critical inventory of vascular plants of the circum Mediterranean countries. 4. – Genève: Conservatoire et Jardin botaniques de la Ville de Genève; Berlin: Secrétariat Med-Checklist, Botanischer Garten und Botanisches Museum Berlin-Dahlem.
- Halbritter, H., Ulrich, S., Grimsson, F., Weber, M., Zetter, R., Hesse, M., Buchner, R., Svojtka, M., Frosch-Radivo, A., 2018: Illustrated pollen terminology. Second edition. Springer.
- Hamzaoğlu, E., Koç, M., 2020: *Hedysarum turcicum* (Hedysareae, Fabaceae), a new species from Turkey. Phytotaxa 428(1), 1–6. <https://doi.org/10.11646/phytotaxa.428.1.1>
- Hedge, I. C., 1979: *Hedysarum* L. In: Davis, P. (ed.), Flora of Turkey and the East Aegean Islands. 560–589. Edinburgh University Press, Edinburgh.
- Kandemir, A., Sevindik, E., Yıldız, F., Türkoğlu, H. İ., Yıldız, B., 2023: A new species of *Hedysarum* (Fabaceae) from Turkey. Phytotaxa 587(1), 12–20. <https://doi.org/10.11646/phytotaxa.587.1.2>
- Kovach, W. L., 2013: MVSP – A MultiVariate Statistical Package, MVSP Version 3.22, Kovach Computing Services, Pentraeth, Wales, U.K. <http://www.kovcomp.co.uk/down12.html>
- Legume Phylogeny Working Group, 2013: Legume phylogeny and classification in the 21<sup>st</sup> century: progress, prospects and lessons for other species-rich clades. Taxon 62(2), 217–248. <https://doi.org/10.12705/622.8>
- Lesins, K. A., Lesins, I., 1979: Genus *Medicago* (Leguminosae): a taxogenetic study. Dr. W Junk bv Publishers, Boston.
- McNeill, J. 1979: Structural value: a concept used in the construction of taxonomic classifications. Taxon 28, 481–504. <https://doi.org/10.2307/1219786>
- Moore, P. D., Webb, J. A., Collison, M. E., 1991: Pollen analysis. Blackwell scientific publications, Oxford.
- Ohashi, H., 1971: A taxonomic study of the tribe Coronilleae (Leguminosae), with a special reference to pollen morphology. Journal of the Faculty of Science, University of Tokyo, Section III Botany 3, 25–92.
- Pavlova, D. K., Manova, V. I., 2000: Pollen morphology of the genera *Onobrychis* and *Hedysarum* (Hedysareae, Fabaceae) in Bulgaria. Annales Botanici Fennici 37(3), 207–217. <http://www.jstor.org/stable/23726902>
- Perveen, A., Kaiser, M., 1998: Pollen flora of Pakistan - X. Leguminosae (subfamily: Caesalpinioideae). Turkish Journal of Botany 22(3), 145–150.
- Pınar, N. M., Vural, C., Aytaç, Z., 2000: Pollen morphology of *Ebenus* L. (Leguminosae: subfamily Papilinoideae) in Turkey. Pakistan Journal of Botany 32(2), 303–310.
- Polhill, R. M., 1981: Evolution and systematics of the Leguminosae. Advances in Legume Systematics 1, 1–26.
- Punt, W., Hoen, P. P., Blackmore, S., Nilsson, S., Le Thomas, A., 2007: Glossary of pollen and spore terminology. Review of Palaeobotany and Palynology 143(1–2), 1–81. <https://doi.org/10.1016/j.revpalbo.2006.06.008>
- Sa, R., Su, D., Debreczy, Z., 2010: Taxonomic notes on the *Hedysarum gmelinii* complex (Fabaceae). Annales Botanici Fennici 47(1), 51–58. <http://www.jstor.org/stable/23728274>
- Sokal, R.R., Rohlf, F.J., 1962: The comparison of dendrograms by objective methods. Taxon 11, 33–40. <http://dx.doi.org/10.2307/1217208>.
- Wodehouse, R. P., 1935: Pollen grains. McGraw-Hill Book Company, New York, London.



# Morpho-palynological assessment of the genus *Terminalia* L. (Combretaceae) in Egypt

Wafaa Kamal Taia<sup>1</sup>, Rim Samir Hamdy<sup>2</sup>, Amany Mohamed Abd El-Maged<sup>3</sup>

<sup>1</sup>Alexandria University, Faculty of Science, Botany and Microbiology Department, 21568 Alexandria, Egypt

<sup>2</sup>Cairo University, Faculty of Science, Botany and Microbiology Department, 12613 Cairo, Egypt

<sup>3</sup>Alexandria University, Faculty of Education, Biology and Geology Department, 5429101 Alexandria, Egypt

**Abstract** – Eleven *Terminalia* species growing in streets and gardens in Cairo (Egypt) were investigated for their pollen morphology. The study's objectives were to explore the variation among *Terminalia* species' pollen pheno-characters and to categorize the various species within this genus according to these pollen characteristics. The results showed that there are significant variations in the pollen morphological characteristics in the genus under study, and as a result, these traits can be regarded as eurypalynous. Each of the two main categories of *Terminalia* species mono-morphic and di-morphic includes various pollen morphologies, ranging from spheroidal to prolate with hexa-aperture that may be similar or of two types. For each of the eleven species, an identification key and a clustering phenogram for the fifteen major pollen features have been created. The aperture type, aperture similarity, and exine ornamentation are some of the crucial traits for the differentiation of the species examined while the pollen shape class has a poor diagnostic character. According to the unique characteristics of the pollen morphology of the *Terminalia* species, three groups and two subgroups were identified.

**Keywords:** microscopy, palynology, pollen key, species determination, taxonomy

## Introduction

The family Combretaceae is an advanced flowering plant with a worldwide tropical distribution (Hutchinson 1969). The family has a different number of genera, ranging from 16 (Rendle 1956) to 20 (Mabberley 2017). Cronquist (1981) recognized 400 species, while Willis (1966) recognized 600 species within this family. These variations in the number of genera and species are due to a lot of synonyms and the either clumping or splitting of genera (Scott 1979). Classification of Combretaceae has been faced with many opinions since Exell (1954) as the family was divided into two subfamilies, Strephonematoideae with the monotypic genus *Strephonema* and Combretoideae with the rest of the genera. More recent treatments by Exell and Stace (1966) and Stace (2007), divided the family into two tribes, Laguncularieae and Combreteae. The latter tribe was subdivided into three subtribes: Combretinae, Pteleopsidinae and Terminaliinae. *Terminalia* is the second largest genus after the genus *Combretum* under the subtribe Combritenae, tribe Combreteae, family Combretaceae, suborder Myrtineae, order Myrtales, (Stace 2002, Heywood et al. 2007). Species

under this genus are shrubs, trees, and creepers distributed in tropical and subtropical regions, especially in Africa and savannas (Arundhati et al. 2020). Many of its species have high economic value as they are used widely in commercial applications and Chinese traditional medicine (Zhang et al. 2019). The genus name came from the word terminal in Latin, as their leaves are crowded at the tops of the shoots. Recognition of the species under the genus *Terminalia* was according to fruit and leaf morphological characters (Chakrabarty et al. 2019, Rayane et al. 2022). The species under this genus differ greatly in their internal and external features in addition to their chromosome numbers (Stace 1965, Ohri 1996). Wickens (1973) and Fyhrquist (2007) found that bark, leaves, and fruit morphology can be used as diagnostic features in the identification of *Terminalia* species. Since species under the genus *Terminalia* are traditionally used in the treatment of some illnesses, precise identification and recognition of the closely related species need clarification.

\* Corresponding author email: monytag86@gmail.com

Flowers in the genus *Terminalia* are grouped in terminal simple racemes with duplicates of five anthers alternating with five petals. Despite the numerous stamens, only a few pollen morphological studies have dealt with taxonomical issues of the genus, e.g. Erdtman (1966), Patel et al. (1984), El-Ghazali et al. (1998), Krachai and Pornpongrungrueng (2015). Other palynological articles were mostly focused on the family level. The taxonomic disagreement concerning the species in the genus *Terminalia*, compounded by the challenge of accurately identifying certain species based only on macro-morphological characteristics, is the reason for this investigation. The aims of this work are to investigate the variation in the pollen pheno-characters of *Terminalia* species, and to classify the different species under this genus with respect to their pollen characters. Therefore, the subject of this investigation was the pollen grain classification of eleven *Terminalia* cultivated species, planted in Cairo parks, streets, and botanical gardens as ornamental trees.

## Material and methods

Mature unopened flowers of eleven species were collected from Egyptian botanical gardens and streets during the flowering season (Tab. 1). The flowers were opened carefully using two needles under a stereo-microscope to release the anthers. The obtained anthers were smeared onto cleaned glass slides with a few drops of glycerol, covered, and sealed with paraffin wax. In order to prevent the harmomegathic effects – which occur when chemicals are exposed to aperturate pollen grains during the acetolysis process and the grains fold in response to changes in humidity (Taia 2022), non-acetolysed pollen grains were examined and then measured and photographed using a Motic (B-150D) light microscope (LM) fitted with a USB digital-video camera and computer software with 10× and 40× objective lenses.

All the measurements were based on 20 to 30 pollen grains. A calibrated oculometer was used to obtain the min-

imum and maximum measures and calculate the mean and standard deviation of each parameter and the apocolpium index (polar area index), which was calculated as the ratio of the mean distance between the apices of two colpi in both the main and sub apertures to its equatorial diameter. Values are expressed in micrometers.

For scanning electron microscope (SEM) investigation, non-acetolysed pollen grains were sputtered onto double sticks fixed in cleaned, aluminum labeled stubs, then coated with 20 nm gold in a Polaron JFC-1100 coating unit, finally examined and photographed using JEOL-JSM.I T200 Series Scanning Electron Microscope made available in the electron microscope unit, Faculty of Science, Alexandria University, Egypt. The terminology used for the descriptions is that of Punt et al. (2007).

Fifteen pollen morphological characters have been subjected to cluster analyses using PAST 3 programs V.3. designed by Hammer (1999-2013).

To generate a key, the data obtained from the description of pollen characters were subjected to automated key generation using version 4.12 of the DELTA suite of programs (Dallwitz et al. 2000).

## Results

### General description of *Terminalia* pollen grains

For the accurate identification of members of the *Terminalia* species, the morphological characteristics of pollen – such as size, shape, aperture conditions, and exine ornamentation – are crucial. The morphological properties of pollen, both quantitative and qualitative, derived by LM and SEM analysis are compiled in Tab. 2 and Fig. 1 and Fig. 2. The examined species' pollen grains are eurypalynous, meaning they vary greatly in character, shed in monads, apolar or isopolar and radially symmetric. These species are classified as either monomorphic or dimorphic. *T. arjuna*, *T. bentzoe*, *T. catappa*, *T. mantaly*, and *T. muelleri* are examples of monomorphic plants; on the other hand, the

**Tab. 1.** The localities of collected specimens, geographic coordinates, and the date of collection of the investigated *Terminalia* species, kept at Cairo University Herbarium (CAI).

No	Species	Locality	Date	Collector
1	<i>T. arjuna</i> (Roxb.) Wight & Arn.	Cairo: Zamalek, El-Giza street 30° 02' N 31° 13' E 12	July 2023	Rim Hamdy
2	<i>T. bellirica</i> (Gaertn.) Roxb.	Giza: Zoological garden 30° 01' 27.4" N 31° 12' 52.7" E 7	Nov. 2016	Heba Mostafa
3	<i>T. bentzoe</i> (L.) L. f.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	July 2016	Heba Mostafa
4	<i>T. brownii</i> Fresen.	Giza: Mazhar botanical garden 30° 03' N 31° 08' E 10	Dec. 2016	Heba Mostafa
5	<i>T. catappa</i> L.	Giza: Orman botanical garden 30° 01' N 31° 12' E 7	May 2019	Rim Hamdy
6	<i>T. chebula</i> Retz.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	Sept. 1963	Mohamed El-Mahdi
7	<i>T. laxiflora</i> Engl. & Diels	Cairo: El Zohriya garden 30° 02' N 31° 13' E 16	July 2023	Rim Hamdy
8	<i>T. mantaly</i> H. Perrier	Giza: Mazhar botanical garden 30° 03' N 31° 08' E 10	Feb. 2019	Rim Hamdy
9	<i>T. muelleri</i> Benth.	Giza: Zoological garden 30° 01' 35.2" N 31° 12' 52.4" E 7	Nov. 2016	Heba Mostafa
10	<i>T. myriocarpa</i> Van Heurck & Mull.	Giza Zoological garden 30° 01' 34.8" N 31° 12' 46.0" E 7	Dec. 2019	Heba Mostafa
11	<i>T. sericea</i> Burch. ex DC.	Giza: Zoological garden 30° 01' 34.0" N 31° 12' 46.0" E 7	June 1969	Mohamed El-Mahdi

**Tab. 2.** Pollen morphological data of 11 *Terminalia* species from Cairo, Egypt. AI – apocolpium index, Amb – the outline of pollen grain seen in polar view, Cir – circular, Di – dimorphic, E – equatorial axis diameter ( $\mu\text{m}$ ), Eth – exine thickness ( $\mu\text{m}$ ), EO – exine ornamentation, Gr – group, Hlob – hexalobate, M – mean, Mono – monomorphic, Min–Max – minimum and maximum values, No – number, P – polar axis length ( $\mu\text{m}$ ), P/E – polar axis/equatorial axis, Pr – prolate, RG – rugate granulate, RP – rugate perforate, Sca – scabrate, SD – standard deviation, Sh – shape, SP – subprolate, Sph – spheroidal, St – striate, Str – striate rugate, Tr – triangular.

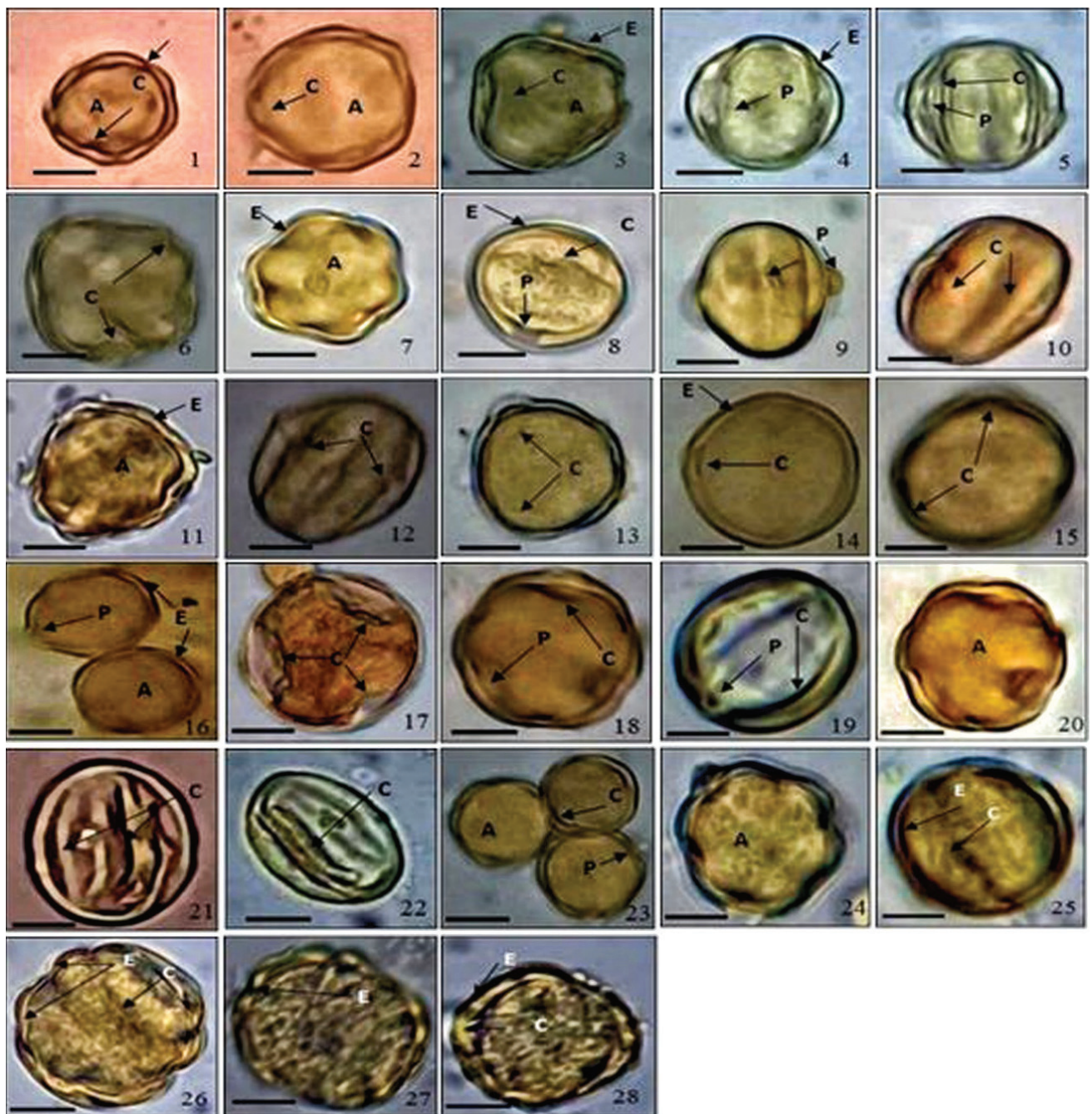
No	Species	State	Min – Max M $\pm$ SD	P	E	P/E	Sh	Amb	Eth	EO	No	Main apertures Type	Length	AI	No	Sub apertures Type	Length	AI
1	<i>T. arjuna</i>	Mono	24.8 – 33.6 30.2 $\pm$ 3.4	24.8 – 33.6 30.2 $\pm$ 3.4	23.8 – 30.2 28.7 $\pm$ 1.5	1.12	SP	Hlob, Cir	2.6	Str	3	Colpate	23.8 – 32.6 29.8 $\pm$ 2.8	0.25	3	Colpate	21.2 – 30.2 28.4 $\pm$ 1.8	0.34
2	<i>T. bellirica</i>	Di	24.8 – 32.2 29.2 $\pm$ 3.0	24.8 – 32.2 29.2 $\pm$ 3.0	21.8 – 28.2 26.5 $\pm$ 1.7	1.15	SP	Tr	2.0	Str	3	Colpate	22.8 – 30.2 29.2 $\pm$ 3.0	0.22	3	Colpate	21.5 – 29.8 26.2 $\pm$ 1.6	0.30
3	<i>T. bentzoe</i>	Mono	25.2 – 30.4 28.2 $\pm$ 1.8	25.2 – 30.4 28.2 $\pm$ 1.8	24.8 – 30.6 29.0 $\pm$ 1.6	1.0	Sph	Tr	2.2	Str	3	Colpate	22.0 – 25.4 24.2 $\pm$ 1.6	0.21	3	Colpate	20.2 – 24.2 22.8 $\pm$ 1.4	0.28
4	<i>T. brownii</i>	Di	23.8 – 28.2 25.6 $\pm$ 2.6	23.8 – 28.2 25.6 $\pm$ 2.6	22.4 – 26.0 25.2 $\pm$ 1.8	1.1	Sph	Hlob	2.2	Sca	3	Colpate	21.2 – 26.4 22.6 $\pm$ 3.8	0.19	3	Colpate	20.4 – 22.6 22.2 $\pm$ 0.4	0.22
5	<i>T. catappa</i>	Mono	24.2 – 31.6 28.4 $\pm$ 3.2	24.2 – 31.6 28.4 $\pm$ 3.2	23.2 – 29.8 27.6 $\pm$ 2.2	1.12	Sp	Tr	2.2	St	3	Colpate	23.2 – 29.6 23.8 $\pm$ 5.8	0.16	3	Colpate	22.8 – 28.2 23.2 $\pm$ 5.0	0.19
6	<i>T. chebula</i>	Di	25.0 – 32.2 29.8 $\pm$ 2.4	25.0 – 32.2 29.8 $\pm$ 2.4	21.8 – 29.8 27.6 $\pm$ 2.2	1.26	Pr	Tr	2.2	St	3	Colpate	24.2 – 31.2 29.4 $\pm$ 1.8	0.14	3	Colpate	22.8 – 30.2 26.6 $\pm$ 3.6	0.20
7	<i>T. laxiflora</i>	Di	25.8 – 34.6 33.6 $\pm$ 1.0	25.8 – 34.6 33.6 $\pm$ 1.0	23.2 – 29.8 27.4 $\pm$ 2.4	1.25	Pr	Tr	2.5	RP	3	Colpate	23.4 – 32.8 30.2 $\pm$ 2.6	0.21	3	Colpate	21.2 – 30.6 28.6 $\pm$ 2.0	0.25
8	<i>T. mantaly</i>	Di	22.4 – 27.6 25.2 $\pm$ 2.4	22.4 – 27.6 25.2 $\pm$ 2.4	20.0 – 24.2 23.2 $\pm$ 1.0	1.12	SP	Cir	2.6	Sca	3	Colpate	20.4 – 24.8 22.8 $\pm$ 2.0	0.24	3	Colpate	16.8 – 18.2 17.0 $\pm$ 1.2	0.36
9	<i>T. muelleri</i>	Mono	22.2 – 27.2 26.2 $\pm$ 1.0	22.2 – 27.2 26.2 $\pm$ 1.0	22.6 – 27.8 26.0 $\pm$ 1.8	1.0	Sph	Cir	2.4	Sca	3	Colpate	19.2 – 24.8 22.0 $\pm$ 2.8	0.28	3	Colpate	15.4 – 18.2 17.2 $\pm$ 1.0	0.41
10	<i>T. myriocarpa</i>	Di	21.8 – 26.8 24.2 $\pm$ 2.6	21.8 – 26.8 24.2 $\pm$ 2.6	20.0 – 24.8 23.8 $\pm$ 1.0	1.1	Sp	Tr	2.2	RG	3	Colpate	19.2 – 24.8 23.6 $\pm$ 1.2	0.25	3	Colporoidate	19.0 – 24.2 23.6 $\pm$ 0.6	0.26
11	<i>T. sericea</i>	Di	24.8 – 33.2 29.2 $\pm$ 4.0	24.8 – 33.2 29.2 $\pm$ 4.0	19.8 – 26.4 24.2 $\pm$ 2.2	1.22	Pr	Hlob	2.2	RG	3	Colpate	21.8 – 30.2 28.2 $\pm$ 2.0	0.26	3	Colporoidate	20.8 – 29.6 26.8 $\pm$ 2.8	0.28
12	<i>T. mantaly</i>	Mono	24.4 – 32.2 30.2 $\pm$ 1.0	24.4 – 32.2 30.2 $\pm$ 1.0	23.8 – 30.4 29.2 $\pm$ 1.2	1.12	Sph	Hlob	2.2	Str	3	Colpate	20.2 – 28.2 27.8 $\pm$ 0.4	0.28	3	Colpate	19.2 – 27.2 25.8 $\pm$ 1.4	0.32
13	<i>T. muelleri</i>	Mono	24.6 – 31.8 29.2 $\pm$ 2.6	24.6 – 31.8 29.2 $\pm$ 2.6	23.4 – 30.8 28.2 $\pm$ 2.6	1.12	Sp	Hlob	2.6	Str	3	Colpate	22.2 – 29.6 27.2 $\pm$ 2.4	0.28	3	Colporoidate	20.2 – 26.4 24.8 $\pm$ 1.6	0.32
14	<i>T. myriocarpa</i>	Di	24.8 – 28.2 25.8 $\pm$ 2.4	24.8 – 28.2 25.8 $\pm$ 2.4	23.2 – 26.4 24.8 $\pm$ 1.6	1.1	SP	Cir	2.4	RP	3	Colpate	22.2 – 26.4 23.6 $\pm$ 2.8	0.34	3	Colpate	20.0 – 24.2 22.6 $\pm$ 1.6	0.38
15	<i>T. myriocarpa</i>	Di	27.8 – 34.5 32.4 $\pm$ 2.1	27.8 – 34.5 32.4 $\pm$ 2.1	24.2 – 30.8 28.6 $\pm$ 2.0	1.25	Pr	Cir	2.4	RP	3	Colpate	24.2 – 30.8 28.2 $\pm$ 2.6	0.32	3	Colpate	20.2 – 27.6 25.8 $\pm$ 1.8	0.36
16	<i>T. sericea</i>	Di	18.6 – 24.4 22.0 $\pm$ 2.2	18.6 – 24.4 22.0 $\pm$ 2.2	17.2 – 23.4 21.2 $\pm$ 2.2	1.12	SP	Hlob	2.2	RP	3	Colpate	18.8 – 20.4 19.2 $\pm$ 1.2	0.32	3	Colporoidate	15.2 – 19.8 18.6 $\pm$ 1.2	0.42
17	<i>T. sericea</i>	Di	20.2 – 24.4 23.2 $\pm$ 1.2	20.2 – 24.4 23.2 $\pm$ 1.2	20.0 – 24.6 23.2 $\pm$ 1.4	1.0	Sph	Cir	2.2	RP	3	Colpate	19.0 – 21.2 20.4 $\pm$ 1.2	0.34	3	Colporoidate	18.2 – 21.0 19.8 $\pm$ 1.6	0.41

dimorphic species were represented by *T. bellirica*, *T. brownii*, *T. chebula*, *T. laxiflora*, *T. myriocarpa* and *T. sericea*; indicating that they have two distinct pollen shapes, sizes, apertures, and even exine ornamentation within the pollen grains that are gathered from the same flower.

**Shape and size**

In their polar view, the pollen grains were triangular, hexa-lobed or circular (Fig. 1 and Fig. 2). In the equatorial view, they were spheroidal (*T. bellirica*, *T. bentzoe*, *T. chebula*, *T. mantaly* and *T. sericea*), subprolate (*T. arjuna*, *T. bellirica*, *T. brownii*, *T. chebula*, *T. laxiflora*, *T. muelleri*, *T. myriocarpa*

and *T. sericea*) and prolate (*T. brownii*, *T. catappa*, *T. laxiflora* and *T. myriocarpa*). *T. catappa* had the largest pollen diameter on the polar axis (P) at 34.6 µm, while *T. sericea* had the smallest (18.6 µm). Similarly, *T. muelleri* and *T. myriocarpa* had the biggest pollen diameter on the equatorial axis (E) (30.8 µm), whereas *T. sericea* had the smallest diameter (17.2 µm). Based on the ratio between the lengths of the polar axis (an imaginary straight line connecting the two poles) to the equatorial diameter (P/E ratio), pollen sizes and shape variations were examined; *T. brownii* had the highest P/E value (1.26), while *T. bellirica*, *T. chebula*, and *T. sericea* had the lowest values (0.1).

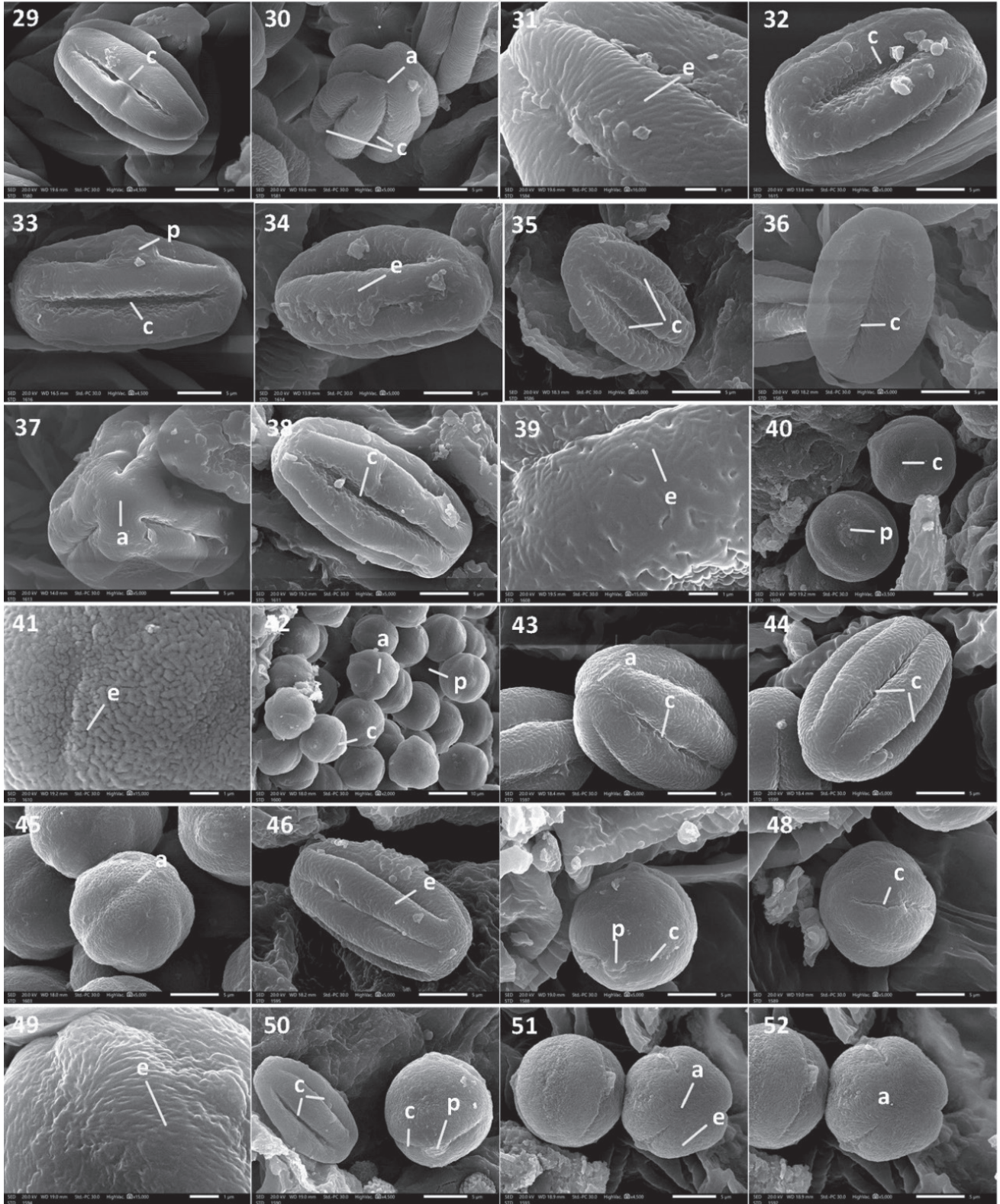


**Fig. 1.** Light microscopy micrographs of the studied *Terminalia* species. (1-2) *T. arjuna*, (3-6) *T. bellirica*, (7-8) *T. bentzoe*, (9-10) *T. brownii*, (11- 12) *T. catappa*, (13- 16) *T. chebula*, (17-19) *T. laxiflora*, (20) *T. mantaly*, (21-22) *T. muelleri*, (23-25) *T. myriocarpa*, (26-28) *T. sericea*. (A – amb, C – colpus, E – exine, P – pore, scale bar = 10 µm).

**Aperture conditions**

Apertures are the first characteristic to take into account when identifying pollen. In *T. arjuna*, *T. brownii*, and *T. catappa*, the apertures are iso-colpate; in the other investi-

gated species, the apertures are hetero-colpate; they have two or more types of colpi, one of which varies in length and/or in whether endoapertures are present. The six apertures seen on the pollen grains of the *Terminalia* species



**Fig. 2.** Scanning electron microscope microphotographs showing the different pollen characters within the studied *Terminalia* species. (29-31) *T. arjuna*, (32-34) *T. bellirica*, (35) *T. bentzoe*, (36-37) *T. brownii*, (38-39) *T. catappa*, (40-41) *T. chebula*, (42-46) *T. laxiflora*, (47-49) *T. mantaly*, (50-52) *T. myriocarpa*. (a – amb, c – colpus, e – exine, p – pore). Scale bar = 1 µm (31, 39, 41, 49), 5 µm (29, 30, 32-38, 40, 43-48, 50-52), 10 µm (42).

under study are composed of three primary, long apertures that alternate with three smaller, possibly distinct, but somewhat shorter apertures.

Pollens of the species *T. arjuna* are hexa-colpate in which the main apertures and the sub-apertures are of the same type. The pollen grains of the species *T. bellirica*, *T. bentzoe*, *T. chebula*, *T. mantaly* and *T. myriocarpa* are hexa-colpate in which the main apertures are colpate and the sub-apertures are colporate. Also, hexa-colpate pollen grains are observed in *T. laxiflora*, *T. muelleri* and *T. sericea* in which the main apertures are colpate and the sub-apertures are colporoidate. In *T. brownii* and *T. catappa* the pollen grains are hexa-colporate in which the main apertures and the sub-apertures are of the same type.

The colpi are not connected at the poles and the apocolpium indices varied between the studied species from 0.14 in *T. brownii* to 0.42 in *T. sericea*. The aperture membranes are either psilate or granulate. In the case of composite apertures, the pores are either small and in the same colpi direction (lolongate) or oval shaped covered by granulated exinous layer (colporoidate). According to the state of the pollen grains, mono- or di- morphic and aperture types the studied species are grouped mainly in two main groups and four subgroups (Tab. 2).

**Exine ornamentation**

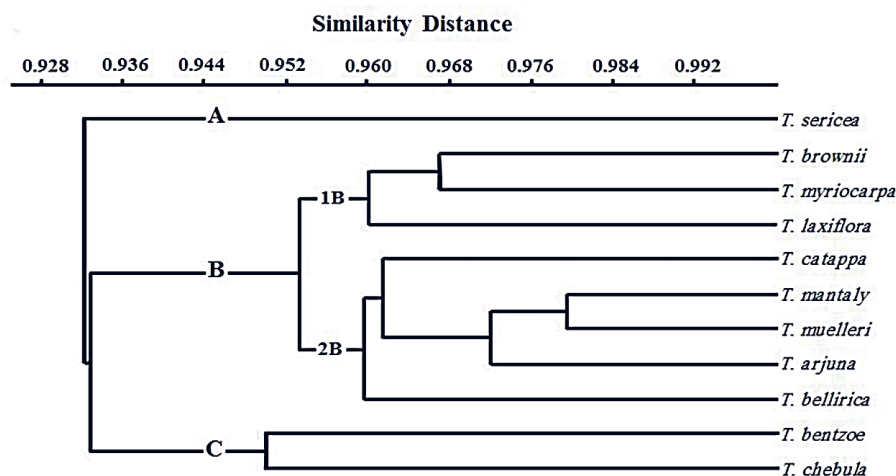
Exine sculpturing is the most distinguishing character of the species; in this study, the species have some variation in exine ornamentation; the exine is generally tectate, scabrate in *T. bentzoe*, and *T. chebula*; rugate in *T. laxiflora*; rugate perforate in *T. catappa*, *T. myriocarpa* and *T. sericea* with thin ectexine and thick endexine and intine layers as observed by light microscope investigation. Striate ornamentation is observed only in *T. brownii* while striate-rugate ornamentation is recorded in *T. arjuna*, *T. bellirica*, *T. mantaly* and *T. muelleri* (Tab. 2). Exine thickness ranges from the minimum (2.0 µm) in *T. bellirica* to the maximum (2.6 µm) in *T. arjuna*, *T. chebula* and *T. muelleri* (Fig. 1).

**Clustering analysis**

The phenogram resulting from the cluster analysis of fifteen palynological characters (On-line Suppl. Tab. 1) after coding according to their states (On-line Suppl. Tab. 2) divided the studied taxa into three groups at similarity distance 0.936. Group 1 has one species only, *T. sericea* (11). Group 2 holds eight species, subdivided into two subgroups at similarity distance 0.952, 2A with three species; *T. brownii* (4), *T. laxiflora* (7), *T. myriocarpa* (10); and subgroup 2B with five species; *T. catappa* (5), *T. mantaly* (8), *T. muelleri* (9), *T. arjuna* (1), *T. bellirica* (2). Group 3 includes two species, *T. bentzoe* (3) and *T. chebula* (6) (Fig. 3).

**Computer-generated key**

1. Pollen shape subprolate or prolate ..... 2  
 Pollen shape subprolate or spheroidal ..... 3  
 Pollen shape prolate ..... ***T. catappa***  
 Pollen shape spheroidal ..... 4  
 Pollen shape subprolate ..... 5
2. Amb shape triangular or hexalobate; Exine ornamentation rugate-granulate ..... ***T. laxiflora***  
 Amb shape triangular; Exine ornamentation striate .....  
 ..... ***T. brownii***  
 Amb shape circular; Exine ornamentation rugate-perforate ..... ***T. myriocarpa***
3. Amb shape circular or hexalobate; Exine ornamentation rugate-perforate ..... ***T. sericea***  
 Amb shape triangular; Exine ornamentation striate-rugate ..... ***T. bellirica***  
 Amb shape circular; Exine ornamentation scabrate . .  
 ..... ***T. chebula***
4. Exine ornamentation scabrate ..... ***T. bentzoe***  
 Exine ornamentation striate rugate ..... ***T. mantaly***
5. Amb shape circular or hexalobate; Sub aperture type colpate ..... ***T. arjuna***  
 Amb shape hexalobate; Sub aperture type colporoidate ..... ***T. muelleri***



**Fig. 3.** Phenogram based on 15 pollen characters showing the possible relationships among the studied *Terminalia* species using percent similarity distances.

## Discussion

Morpho-palynological studies have proved to be of great importance for plant identification and taxonomic classification. In this study, different palynological features such as variations in pollen characters in the same flower (morphism), polar and equatorial outline and their ratio (P/E), type of aperture, presence or absence of pores, beside exine thickness and ornamentation were investigated in eleven *Terminalia* (Combretaceae) species belonging to 7 sections from Egypt using LM and SEM. Based on palynological features, a taxonomic key was created for simple identification and species delimitations.

Mignot et al. (1994) pointed to the importance of aperture type in angiosperm recognition at any taxonomic level, even down to the intra individual level. Based on the results, *Terminalia* species may be divided into two main groups: monomorphic and dimorphic. Each group has a variety of pollen shapes, ranging from spheroidal to prolate, with many shapes seen in the pollen sampled from the same flower. Pollen hydration status and environmental conditions may contribute to heterogeneity in pollen shape and size within the same taxon (Grant-Jacob et al. 2022, Taia et al. 2023). El Ghazali (2022) noted differences in the sizes and forms of pollen among the same taxa within the family Combretaceae. Despite this, he defined their pollens as stenopalynous, meaning that their characteristics are either stable or slightly changed. According to this investigation, the *Terminalia* under study exhibit significant variety in their pollen morphological characteristics, and as a result, they can be classified as eurypalynous.

The highest PA length is seen in *T. myriocarpa* (27.8-34.5 µm), whereas *T. sericea* has the smallest PA length (18.6-23.2 µm). El Ghazali et al. (1998) have documented these differences in pollen size and shape. Given that multiple shape types have been observed in a single flower, shape class is determined to have less diagnostic value within the taxa in the material under examination. Taia et al. (2023) have noted this investigation; as a result, it does not play a significant role in the taxonomic discrimination of the examined taxa, except in the case of *T. sericea*, which is grouped separately in the clustering analysis (group A), this separation being due to it having the smallest PA and ED lengths and the smallest main aperture and sub-aperture lengths. This result supports the classification of this species as it belongs to section Psidioides Exell.

Four monomorphic species (*T. catappa*, *T. mantaly*, *T. muelleri*, and *T. arjuna*) were gathered by the monomorphic character within the same taxa and placed in subgroup 2B in close proximity to the dimorphic taxa (*T. bellerica*). This grouping strengthens the taxonomic status of *T. catappa*, *T. mantaly*, *T. muelleri* as they belong to the same section Eucatappa Engl. & Diels. Despite *T. arjuna* being a separate section Pentaptera (Roxb.) Engl. & Diels, it is grouped with the former three species according to the monomorphic pollen character.

According to El Ghazali and Krzywinski (1989), the apertures can be classified as iso- or hetero-colpate, meaning

that they have two or more types of colpi, with one type differing in length and/or having endoapertures or not. This previously acknowledged observation was discovered in *Rosa sericea* (Ullah et al. 2022). It was also discovered in *Fagonia* species (Taia et al. 2021). El-Ghazali et al. (1998) noted the heterocolpate pollen grains within the taxa under study. This study noted variations in the length and width of the colpi as well as the existence and dimensions of endo-apertures; this result is in line with previous studies mentioned.

Since *T. brownii*, *T. laxiflora*, and *T. myriocarpa* have triangular or circular amb, the triangular amb gathered them in subgroup 1B. Amb shape was determined to be a valuable character in the building of the identification key. This result is partially in agreement with the taxonomy of *T. brownii*, *T. laxiflora* as they belong to the same section, Platycarpae Engl. & Diels, but does not agree with the classification of *T. myriocarpa* as a separate section Myriocarpae. A triangular amb is also present in *T. bellirica*, although it is distinct from the preceding three species in terms of other pollen characteristics and is located in subgroup 2B, albeit remaining closely linked to them. Certain species could be distinguished using the hexagonal, triangular, or circular amb, as the identification key illustrates. The two species; *T. bellirica* and *T. bentzoe* are belonging to section Billiricae Engl. & Diels, however, the study separated them in different groups; this separation is due to the difference between them in pollen state and amb shape.

Exine thickness ranged from 2.0 µm to 2.6 µm; however, there were significant differences in exine ornamentation. The rugate-perforate pattern was seen in *T. myriocarpa* and *T. sericea*, while the striate or striate-rugate pattern was seen in the majority of the investigated species. Only *T. laxiflora* has rugate-granulate ornamentation, but *T. bentzoe* and *T. chebula* have scabrate exine ornamentation; for this reason, the former two species were grouped together in group C although they belong to different sections; *T. bentzoe* sec. Belliricae Engl. & Diels, *T. chebula* sec. Myrobalanus (Gaertn.) Eichl. As a result, exine thickness varies very little among the taxa, whereas exine ornamentation, displayed by the key, is more helpful in identifying the taxa. This observation is consistent with every palynological study conducted since that of Erdtman (1966).

## Conclusion

Based on the study's findings, three groups and two subgroups can be identified based on the unique characteristics of the pollen morphology of the *Terminalia* species under investigation. The pollen morphism, aperture homogeneity, amb form, and exine ornamentation are the most crucial characteristics in classifying the investigated taxa.

## Acknowledgment

The authors are grateful to the Electron Microscope Unit, Faculty of Science, Alexandria University for completing this work.

## References

- Arundhati, D., Akshayakumar, B., Ranjan, R. G., 2020: Molecular characterization and genetic diversity assessment of ten plant species under family Combretaceae using ISSR markers. *Research Journal of Biotechnology* 15(12), 11–18.
- Chakrabarty, T., Krishna, G., Rasingam, L., 2019: Taxonomic notes on Indian *Terminalia* (Combretaceae). *Plant Science Today* 6(3), 281–286. <https://doi.org/10.14719/pst.2019.6.3.539>
- Cronquist, A., 1981: An integrated system of classification of flowering plants. Columbia University Press, New York. <https://doi.org/10.2307/2806386>
- Dallwitz, M. J., Paine, T. A., Zurcher, E. J., 2000: Onwards. Principles of interactive keys. Website <http://delta-intkey.com/www/interactivekeys.htm> [accessed 16 December 2014].
- El Ghazali, G. E., Krzywinski, K., 1989: An attempt to clarify the term heterocolpate. *Grana* 28(3), 179–186. <http://dx.doi.org/10.1080/00173138909427429>.
- El Ghazali, G. E., Tsuji, S., El Ghazaly, G. A., Nilsson, S., 1998: Combretaceae R. Brown. *World Pollen and Spore Flora* 21, 1–40.
- El Ghazali, G. E., 2022: The genus *Buchenavia* Eichl. and its taxonomic affinity to the genus *Terminalia* L. (Combretaceae): Insight from pollen morphology—a review. *Review of Palaeobotany and Palynology* 301, 104644. <https://doi.org/10.1016/j.revpalbo.2022.104644>
- Erdtman, G., 1966: Pollen morphology and plant taxonomy. Angiosperms. (An Introduction to Palynology I). Hafner Publishing Company, New York and London.
- Exell, A. W., 1954: Combretaceae. *Flora Malesiana* 4, 533–589.
- Exell, A. W., Stace, C. A., 1966: Revision of the Combretaceae. *Boletim da Sociedade Broteriana sér. 2*, 40: 5–26.
- Grant-Jacob, J. A., Praeger, M., Eason, R. W., Mills, B., 2022: Generating images of hydrated pollen grains using deep learning. *IOP SciNotes* 3(2), 024001. <https://doi.org/10.1088/2633-1357/ac6780>.
- Heywood, V. H., Brummitt, R. K., Culham, A., Seberg, O., 2007: Flowering Plant Families of the World. Royal Botanic Gardens, Kew. Ontario: Firefly books. <https://doi.org/10.1111/j.1467-8748.2007.00585.x>.
- Hutchinson, J., 1969: Evolution and phylogeny of flowering plants: Dicotyledons: facts and theory, with over 550 illustrations and maps by the author. (No Title). Academic Press, London.
- Fyhrquist, P., 2007: Traditional medicinal uses and biological activities of some plant extracts of African *Combretum* Loeffl., *Terminalia* L. and *Pteleopsis* Engl. Species. Ph.D Thesis, 183p. <http://ethesis.helsinki.fi>.
- Krachai, P., Pornpongrueng, P., 2015: Pollen morphology of Combretaceae from Thailand and its taxonomic significance. *Thai Forest Bulletin (Botany)* 43, 4–14.
- Mabberley, D. J., 2017: Mabberley's plant-book: a portable dictionary of plants, their classification and uses. Cambridge University Press, Cambridge, UK. <https://doi.org/10.1017/9781316335581>
- Mignot, A., Hoss, C., Dajoz, I., Leuret, C., Henry, J. P., Dreuil-laux, J. M., Heberle-Bors, E., Till-Bottraud, I., 1994: Pollen aperture polymorphism in the angiosperms: importance, possible causes and consequences. *Acta Botanica Gallica* 141(2), 109–122. <https://doi.org/10.1080/12538078.1994.10515144>
- Ohri, D., 1996: Genome size and polyploidy variation in the tropical hardwood genus *Terminalia* (Combretaceae). *Plant Systematics and Evolution* 200, 225–232. <https://doi.org/10.1007/BF00984937>
- Patel, V. C., Skvarla, J. J., Raven, P. H., 1984: Pollen characters in relation to the delimitation of Myrtales. *Annals of the Missouri Botanical Garden* 71(3), 858–969. <https://doi.org/10.2307/2399170>
- Hammer, Ø., 1999–2013: PAST paleontological statistics version 3.0: reference manual. University of Oslo.
- Punt, W., Blackmore, S., Nilsson, S., Thomas, A., 2007: Glossary of pollen and spore terminology. *Review of Paleobotany and Palynology* 143(1–2), 1–81. <https://doi.org/10.1016/j.revpalbo.2006.06.008>
- Rayane, T., Ribeiro, M., Maria, I., Loiola, B., Margareth, F. DE. S., 2022: Look at the trees: synopsis of *Terminalia* ss in the Brazilian Amazon with conservation and distribution implications. *Anais da Academia Brasileira de Ciências* 94(4), 1–19. <https://doi.org/10.1590/0001-376520220210265>
- Rendle, A. B., 1956: The Classification of Flowering Plants-Dicotyledons 2. Cambridge University Press, Cambridge, UK. <https://doi.org/10.5962/bhl.title.95518>
- Scott, A. J., 1979: A revision of Anogeissus (Combretaceae). *Kew Bulletin* 1, 555–566. <https://doi.org/10.2307/4109799>
- Stace, C. A., 1965: The significance of the leaf epidermis in the taxonomy of the Combretaceae: I. a general review of tribal, generic and specific characters. *Journal of the Linnean Society of London, Botany* 59(378), 229–252. <https://doi.org/10.1111/j.1095-8339.1965.tb00060.x>
- Stace, C. A., 2002: Proposal to conserve *Terminalia* nom. cons. (Combretaceae) against an additional name *Bucida*. *Taxon* 51(1), 193–194. <https://doi.org/10.2307/1554985>
- Stace, C. A., 2007: Combretaceae. In: K. Kubitzki (ed.), The families and genera of vascular plants, 9, 68–82. Springer Verlag, Berlin, Heidelberg. [https://doi.org/10.1007/978-3-540-32219-1\\_11](https://doi.org/10.1007/978-3-540-32219-1_11)
- Taia, W. K., 2022: Pollen grain diversity and application in taxonomy and evolution. *Taeckholmia* 42(1), 27–42. <https://doi.org/10.21608/taec.2022.258213>
- Taia, W. K., Ibrahim, M., Hassan, S., Asker, A., 2021: Palynological study of the genus *Fagonia* L. (Zygophyllaceae, Zygophylloideae) in Libya. *Libyan Journal of Science and Technology* 13(1), 29–37. <https://doi.org/10.37376/ljst.v13i1.2272>
- Taia, W. K., Hamdy, R. S., Abd El-Maged, A. M., 2023: Morphological diversity of pollen grains within the flowering stages of *Tecoma stans* (L.) Juss. ex Kunth. *GSC Advanced Research and Reviews* 16(1), 280–287. <https://doi.org/10.30574/gscarr.2023.16.1.0320>
- Ullah, F., Gao, Y. D., Zaman, W., Gao, X. F., 2022: Pollen morphology of *Rosa sericea* complex and their taxonomic contribution. *Diversity* 14(9), 705–722. <https://doi.org/10.3390/d14090705>
- Wickens, G. E., 1973: Combretaceae. In: Polhill, R.M. (ed.), *Flora of Tropical East Africa*, 1–99. London, Crown Agents for Oversea Governments and Administrations, United Kingdom.
- Willis, J. C., 1966: A dictionary of the flowering plants and ferns. A dictionary of the flowering plants and ferns 7. Cambridge University Press, Cambridge, UK. <https://doi.org/10.5962/bhl.title.1428>
- Zhang, X. R., Kaunda, J. S., Zhu, H. T., Wang, D., Yang, C. R., Zhang, Y. J., 2019: The genus *Terminalia* (Combretaceae): An ethnopharmacological, phytochemical and pharmacological review. *Natural Products and Bioprospecting* 9, 357–392. <https://doi.org/10.1007/s13659-019-00222-3>



# First record of the woody *Melaleuca williamsii* s.l. (Myrtaceae) out of its native range

Duilio Iamónico<sup>1\*</sup>, Gianluca Nicoletta<sup>2</sup>

<sup>1</sup> University of Rome Sapienza, Department of Environmental Biology, Piazzale Aldo Moro 5, I-00185 Rome, Italy

<sup>2</sup> Via del Gran Paradiso, I-00139 Rome, Italy

**Abstract** – *Melaleuca williamsii* subsp. *synoriensis* (Myrtaceae) has been found in Rome (central Italy) along the banks of the river Tevere, representing the first discovery outside its native range (E-Australia). Description, distribution in Italy, phenology, and original photographs are provided.

**Keywords:** Europe, exotic species, *Melaleuca*, morphology, original material

## Introduction

During the last decade, there has been increasing evidence of the negative impacts of alien species on the environment, the economy, and human well-being in all parts of the world (see e.g. Vilà et al. 2011, Rumlerová et al. 2016, Sohrabi et al. 2023). Provision of data about the distribution and status of the naturalization of alien plants is useful in preventing or reducing these impacts (see e.g. Gallardo et al. 2019, Pyšek et al. 2020.).

In this context, a recent investigation of the genus *Melaleuca* L. by ter Huurne et al. (2023) showed how several species (widely planted around the world mainly for ornamental and pharmaceutical uses) spread rapidly and have significant ecological impacts on autochthonous flora.

*Melaleuca* (Myrtaceae Juss.) is a genus that has been accepted without controversy for over 200 years, but was recently shown to be polyphyletic (Ladiges et al. 1999, Brown et al. 2001, Edwards et al. 2010). Edwards et al. (2010) demonstrated that eight out of the nine genera of the tribe Melaleuceae recognized by Wilson et al. (2005) fall within *Melaleuca* which was accepted as a single species-rich genus including 330–350 taxa (106 nomenclatural changes were proposed by Craven et al. 2014). According to POWO (2024) *Melaleuca* is native to Australia, while some taxa are aliens in Africa, Asia, the Americas and as far as Europe is concerned, only in the former Yugoslavia.

As part of an ongoing study on the alien flora of Italy and the Mediterranean area (see e.g. Iamónico 2010, 2022, Iamónico et al. 2014, Iamónico and Sánchez Del Pino 2016,

Sukhorukov et al. 2016, Iamónico and El Mokni 2017, El Mokni and Iamónico 2018), we found in Rome (central Italy) the woody *Melaleuca williamsii* Craven subsp. *synoriensis* Craven, which represents the first records of the taxon out of its native range. Because of the difficulty we had in the identification of the species of taxon, and with the aim of helping European botanists in understanding this complicated genus, we examined in depth the species found and similar ones by studying also their protologues and associated types and original material.

## Material and methods

The work is based on field surveys, analysis of relevant literature (protologues are included), and examination of specimens preserved in the following herbaria: CANB, GOET, K, MEL, NWS, P, and RO (acronyms according to Thiers 2024+).

The articles cited throughout the text are referred to the Shenzhen Code (hereafter reported as “ICN”; Turland et al. 2018).

## Results

*Melaleuca williamsii* Craven subsp. *synoriensis* Craven, Novon 19: 452–453. 2009 ≡ *Callistemon pungens* subsp. *synoriensis* (Craven) Udovicic & R.D.Spencer, Muelleria 30: 24 (2012)

\* Corresponding author e-mail: duilio.iamonico@uniroma1.it



**Fig. 1.** *Melaleuca williamsii*. A – habitat along the Tevere River (Rome, Italy), B – leaf, C – fruiting branch, D – inflorescence, E – flower (Photo: G. Nicolella).

**Holotype:** Australia, New South Wales, Ca. 200 NE Tom Cabin, NE part of New England Natl. Park, ca. 12 km SE of Ebor, 04 December 1993, P. J. Lepschi & J. Mowatt 1411 (CANB467657!, image of the holotype available at <https://plants.jstor.org/stable/viewer/10.5555/al.ap.specimen.canb467657>); isotypes: BRI (*non vidi fide* Craven 2009: 452), MEL2456739! (<https://plants.jstor.org/stable/viewer/10.5555/al.ap.specimen.mel2456739>), NE (*non vidi fide* Craven 2009: 452), NSW (*non vidi fide* Craven 2009: 452).

**Native distribution area:** Eastern Australia, from Queensland to New South Wales (Craven 2006, POWO 2024).

**Occurrence in Italy:** the plants found in Italy (February 2023) grow among the cracks in the quay along the right bank of the river Tevere in Rome (Latium region, central Italy) (Fig. 1).

Flowering and fruiting times: late winter to late spring (February–June). The discovery is not only the first one for Italy and Europe, but also the first out of the native range of the species (see Craven 2006, Uotila 2011, POWO 2024 and literature therein). We consider *Melaleuca williamsii* subsp. *synoriensis* as casual alien in Italy.

**Notes:** currently, three subspecies are accepted for *Melaleuca williamsii*, i.e. subsp. *williamsii*, subsp. *fletcheri* Craven, and subsp. *synoriensis* Craven (POWO 2024, WFO 2024). These subspecies can be distinguished from each other based on the bark texture, hairiness of leaves and hypanthium, color of filaments, and shape of cotyledons

(Craven 2009). The Roman plants are identifiable as *M. williamsii* subsp. *synoriensis*, having leaves and hypanthium pubescent and staminal filament red [no data about the bark was reported by Craven (2009) for subsp. *synoriensis*, whereas the author distinguished subsp. *williamsii* and subsp. *fletcheri* in having barks with, respectively, papery and fibrous texture], whereas the other two subspecies display leaves lanuginose (subsp. *williamsii*) and filaments white, pink, or mauve (subsp. *fletcheri*).

## Discussion

Although several taxonomic issues on *Melaleuca* were solved, some others, concerning the species rank, still need to be addressed. Indeed, we had difficulties in identifying the *Melaleuca* plants found in Rome even when we referred to the important diagnostic key to Australian *Melaleuca* taxa prepared by Craven et al. (2016). Issues arise for the leaf features, i.e. shape (some statements are reported in both the key phrases of step no. 5 of Australian *Melaleuca*'s key) and width. Roman plants have leaves 0.3–0.6 cm wide, a range that overlaps the choice of the step no. 5 in diagnostic key of Australian *Melaleuca* (“Leaves less than 4 mm wide” vs. “Leaves 4 mm or more wide”). By choosing both the phrases of step no. 5, we reached the following candidates: *M. linearis*, *M. subulata*, and *M. williamsii*. However, to attain a certain identification, we decided to examine carefully these three species, especially by studying the protologues, types, and original material.

The results follow:

- 1) *Melaleuca linearis* [described by Wendland and Schrader 1796: 19, Tab. XI" (image available at [http://www.plant-illustrations.org/illustration.php?id\\_illustration=246556](http://www.plant-illustrations.org/illustration.php?id_illustration=246556)); lectotype designated by Dowe et al. (2019) on a specimen preserved at GOET]: leaf blade has a ratio length/width of at least 10 (up to 15).
- 2) *Melaleuca subulata* [ $\equiv$  *Callistemon subulatus* (described by Cheel 1925: 259); holotype: NSW139989 (image available at <https://plants.jstor.org/stable/viewer/10.5555/al.ap.specimen.nsw139989?loggedin=true>); a further Cheel's specimen (not isotype, found by us) at K (barcode K000793339, image available at <https://plants.jstor.org/stable/viewer/10.5555/al.ap.specimen.k000793339>): ratio length/width of the leaf blade ranging from 10 to 16.
- 3) *Melaleuca williamsii* [proposed by Craven (2006: 474) as *nomen novum pro Callistemon pungens* Lumley & R.D.Spencer *non Melaleuca pungens* Schauer (see Arts. 6.9 and 53.1 of ICN); holotype at MEL; isotypes at K (found by us), NE, NWS, and CANB; a good illustration by Lumley and Spencer (1990: 254, Fig. 1)]: ratio length/width of the leaf blade ranges from 3 to 7.

Based on the ratio length/width of the leaf blade, the Roman plants (ratio 5.5–10.0) overlap *Melaleuca williamsii* (3–7), whereas *M. linearis* and *M. subulata* have higher ratios (10–15 and 10–20, respectively).

In addition to the spontaneous Australian species, we also considered the *Melaleuca* species that are currently used as ornamentals in Italy (personal observations made in plant nurseries), i.e. *M. citrina* (native to E-Australia and alien in India, Kenya and Tanzania, former Yugoslavia, U.S.A., and Mexico; POWO 2024) and *M. viminalis* (native to E-Australia and alien in India, Kenya and Tanzania, U.S.A., and Mexico; POWO 2024):

- 1) *Melaleuca viminalis*: it has stamens fused (*vs.* free and inserted just in a ring on the hypanthium rim in the Roman plants) and hypanthium glabrous (*vs.* pubescent) (Craven et al. 2016).
- 2) *Melaleuca citrina*: this species cannot be ascribed to Roman plants (based on Craven et al. 2016) because of the length of the longest stamens [up to 25 mm long (range 17–25 mm) in *M. citrina vs.* up to 20 mm long (range 13–20 mm) in our plants] and the width of the inflorescences (45–70 mm in *M. citrina vs.* 38–47 mm wide). Concerning the ratio length/width of the blades, we studied the protologue of *M. citrina* and examined the original material. Curtis (1794: [260] validly published the name *Metrosideros citrina* (basionym of *Melaleuca citrina*) also providing a coloured illustration (Plate no. 260, original material). Curtis' Botanical Magazine is an illustrated publication (first issue published on 1 February 1787), the longest-running botanical magazine (currently referred to as the journal Curtis's Botanical Magazine) and including drawings

of ornamental and exotic plants cultivated at Kew Gardens. According to the HUH-Index of Botanists (2013), Curtis' herbarium is unknown. No specimen useful for lectotypification was traced. Therefore, Curtis' Plate no. 260 is the only extant original material for *Metrosideros citrina* and the ratio length/width of the blades ranges from 2 to 6 (*vs.* 5.5–10.0 in our plants). Since the length of the stamens and the width of the inflorescence cannot be measured on Curtis' Plate no. 260 (a scale bar is lacking), we examined specimens collected by L. A. Craven and/or B. J. Lepschi (the experts in the genus *Melaleuca* and authors of the Australian key) and verifying that longer stamens are 21–23 mm long (*vs.* up to 20 mm long), whereas inflorescence are 53–55 mm wide (*vs.* 38–47 mm).

## Acknowledgments

Thanks are due to Directors and Curators of all quoted Herbaria for their support during our visits, loan of specimens/photograph or requests.

Project funded under the National Recovery and Resilience Plan (NRRP), Mission 4 Component 2 Investment 1.4 - Call for tender No. 3138 of 16 December 2021, rectified by Decree n.3175 of 18 December 2021 of Italian Ministry of University and Research funded by the European Union – NextGenerationEU. Project code CN\_00000033, Concession Decree No. 1034 of 17 June 2022 adopted by the Italian Ministry of University and Research, CUP B83C22002950007, Project title "National Biodiversity Future Center - NBFC".

## References

- Brown, G. K., Udovicic, F., Ladiges, P. Y., 2001: Molecular phylogeny and biogeography of *Melaleuca*, *Callistemon* and related genera (Myrtaceae). *Australian Systematic Botany* 14(4), 565–585. <https://doi.org/10.1071/SB00029>
- Craven, L. A., 2006: New combinations in *Melaleuca* for Australian species of *Callistemon* (Myrtaceae). *Novon* 16(4), 468–475. [https://doi.org/10.3417/1055-3177\(2006\)16\[468:NCIM-FA\]2.0.CO;2](https://doi.org/10.3417/1055-3177(2006)16[468:NCIM-FA]2.0.CO;2)
- Craven, L. A., 2009: *Melaleuca* (Myrtaceae) from Australia. *Novon* 19(4), 444–460. <https://doi.org/10.3417/2007137>
- Craven, L. A., Edwards, R. D., Cowley, K. J., 2014: New combinations and names in *Melaleuca* (Myrtaceae). *Taxon* 63(3), 663–670. <https://doi.org/10.12705/633.38>
- Craven, L. A., Lepschi, B. J., Barlow, B. A., Cowley, K. J. B., 2016: Flora of Australia: vascular plants: Species of *Melaleuca*. Retrieved November 29, 2023 from <https://keybase.rbq.vic.gov.au/keys/show/6135>
- Curtis, W., 1794: The Botanical magazine, or, flower-garden displayed: in which the most ornamental foreign plants, cultivated in the open ground, the green-house, and the stove, are accurately represented in their natural Colours 8. Stephen Couchman, London.
- Dowe, J. L., Reiner-Drehwald, M. E., Appelhans, M. S., Hörandl, E., 2019: The Wendlands of Herrenhausen Gardens (Hannover, Germany): a study of their Australian plant specimens in the herbarium of the University of Goettingen (GOET).

- Australian Systematic Botany 32(3), 177–215. <https://doi.org/10.1071/SB18020>
- Edwards, R. D., Craven, L. A., Crisp, M. D., Cook, L. G., 2010: *Melaleuca* revisited: cpDNA and morphological data confirm that *Melaleuca* L. (Myrtaceae) is not monophyletic. *Taxon* 59(3), 744–754. <https://doi.org/10.1002/tax.593007>
- El Mokni, R., Iamónico, D., 2018: A new record for the non-native flora of Tunisia, *Eclipta prostrata* (Ecliptinae, Asteraceae), and a note on the national status of *Erigeron bonariensis*, *Lepidium didymum*, and *Symphytotrichum squamatum*. *Flora Mediterranea* 28, 145–153. <https://doi.org/10.7320/FlMedit28.145>
- Gallardo, B., Bacher, S., Bradley, B., Comín, F. A., Gallien, L., Jeschke, J. M., Sorte, C. J. B., Vilà, M., 2019: InvasiBES: understanding and managing the impacts of invasive alien species on biodiversity and ecosystem services. *NeoBiota* 50, 109–122. <https://doi.org/10.3897/neobiota.50.35466>
- HUH Index of botanists, 2013-onwards: Index of botanists, Harvard University Herbaria & Libraries. Curtis, William. Retrieved February 4, 2024 from [https://kiki.huh.harvard.edu/databases/botanist\\_search.php?mode=details&id=981](https://kiki.huh.harvard.edu/databases/botanist_search.php?mode=details&id=981)
- Iamónico, D., 2010: On the presence of *Amaranthus polygonoides* L. (*Amaranthaceae*) in Europe. *Phyton* (Horn, Austria) 50(2), 205–219.
- Iamónico, D., 2022: Biodiversity in urban areas: the extraordinary case of the Appia Antica Regional Park (Rome, Italy). *Plants* 11(16), 2122. <https://doi.org/10.3390/plants11162122>
- Iamónico, D., El Mokni, R., 2017: *Amaranthus palmeri* (*Amaranthaceae*) in Tunisia, a second record for the continental African flora and nomenclatural notes on *A. sonoriensis* *nom. nov. pro A. palmeri* var. *glomeratus*. *Bothalia* 47(1), a2100. <https://doi.org/10.4102/abc.v47i1.2100>
- Iamónico, D., Iberite, M., Nicoletta, G., 2014: Aggiornamento alla flora esotica del Lazio (Italia centrale). II. *Informatore Botanico Italiano* 46(2), 215–220.
- Iamónico, D., Sánchez Del Pino, I., 2016: Taxonomic revision of the genus *Alternanthera* (*Amaranthaceae*) in Italy. *Plant Biosystems* 150(2), 333–342. <https://doi.org/10.1080/11263504.2015.1019588>
- Ladiges, P. Y., McFadden, G. I., Middleton, N., Orlovich, D. A., Treloar, N., Udovicic, F., 1999: Phylogeny of *Melaleuca*, *Callistemon*, and related genera of the *Beaufortia* suballiance (Myrtaceae) based on 5S and ITS-1 spacer regions of nrDNA. *Cladistics* 15(2), 151–172. <https://doi.org/10.1111/j.1096-0031.1999.tb00257.x>
- Lumley, P. F., Spencer, R. D., 1990: Two new species of *Callistemon* R.Br. (Myrtaceae). *Muelleria* 7(2), 253–257.
- POWO, 2024: Plant of the World Online. *Melaleuca* L. Retrieved February 4, 2024 from <https://powo.science.kew.org/taxon/urn:lsid:ipni.org:names:27624-1>
- Pyšek, P., Hulme, P. E., Simberloff, D., Bacher, S., Blackburn, T. M., Carlton, J. T., Dawson, W., Essl, F., Foxcroft, L. C., Genovesi, P., Jeschke, J. M., Kühn, I., Liebhold, A. M., Mandrak, N. E., Meyerson, L. A., Pauchard, A., Pergl, J., Roy, H. E., Seebens, H., Kleunen, M. van, Vilà, M., Wingfield, M. J., Richardson, D. M., 2020: Scientists' warning on invasive alien species. *Biological Review* 95(6), 1511–1534. <https://doi.org/10.1111/brv.12627>
- Rumlerová, Z., Vilà, M., Pergl, J., Nentwig, W., Pyšek, P., 2016: Scoring environmental and socioeconomic impacts of alien plants invasive in Europe. *Biological Invasions* 18(12), 3697–3711. <https://doi.org/10.1007/s10530-016-1259-2>
- Sohrabi, S., Vilà, M., Zand, E., Gherekhloo, J., Hassanpour-Bourkheili, S., 2023: Alien plants of Iran: impacts, distribution and managements. *Biological Invasions* 25(1), 97–114. <https://doi.org/10.1007/s10530-022-02884-6>
- Sukhorukov, A. P., Martín-Bravo, S., Verloove, F., Maroyi, A., Iamónico, D., Catarino, L., El Mokni, R., Daniel, T. F., Belyaeva, I. V., Kushunina, M., 2016: Chorological and taxonomic notes on African plants. *Botany Letters* 163(4), 417–428. <https://doi.org/10.1080/23818107.2016.1224731>
- ter Huurne, M. B., Potgieter, L. J., Botella, C., Richardson, D. M., 2023: *Melaleuca* (Myrtaceae): Biogeography of an important genus of trees and shrubs in a changing world. *South African Journal of Botany* 162, 230–244. <https://doi.org/10.1016/j.sajb.2023.08.052>
- Thiers, B., 2024+: Index Herbariorum: A global directory of public herbaria and associated staff. New York Botanical Garden's Virtual Herbarium. Retrieved February 4, 2024 from <http://sweetgum.nybg.org/ih/>
- Turland, N. J., Wiersema, J. H., Barrie, F. R., Greuter, W., Hawksworth, D. L., Herendeen, P. S., Knapp, S., Kusber, W.-H., Li, D.-Z., Marhold, K., May, T. W., McNeill, J., Monro, A. M., Prado, J., Price, M. J., Smith, G. F., (eds.) 2018: International Code of Nomenclature for algae, fungi, and plants (Shenzhen Code) adopted by the Nineteenth International Botanical Congress Shenzhen, China, July 2017. *Regnum Vegetabile* 159, 1–254. Koeltz Botanical Books, Glashütten. <https://doi.org/10.12705/Code.2018>
- Uotila, P., 2011+: Juglandaceae. In: Euro+Med Plantbase – the information resource for Euro-Mediterranean plant diversity. Retrieved February 4, 2024 from <https://www.europlusmed.org>
- Vilà, M., Espinar, J. L., Hejda, M., Hulme, P. E., Jarošik, V., Maron, J. L., Pergl, J., Shaffner, U., Sun, Y., Pyšek, P., 2011: Ecological impacts of invasive alien plants: a meta-analysis of their effects on species, communities and ecosystems. *Ecology Letter* 14(7), 702–708. <https://doi.org/10.1111/j.1461-0248.2011.01628.x>
- Wendland, J. C., Schrader, H. A., 1796: *Sertum Hannoveranum seu plantae rariores quae in horticis regni Hannoverae vicinis coluntur* 1(2). Vandenhoeck et Ruprecht, Goettingae.
- Wilson, P. G., O'Brien, M. M., Heslewood, M. M., Quinn, C. J., 2005: Relationships within Myrtaceae *sensu lato* based on a matK phylogeny. *Plant Systematic and Evolution* 251(1), 3–19. <https://doi.org/10.1007/s00606-004-0162-y>
- WFO, 2024: The World Flora Online. *Melaleuca williamsii* Craven. Retrieved February 4, 2024 from <https://www.world-floraonline.org/taxon/wfo-0000809160>

# Changes in grassland vegetation on the island of Plavnik (Croatia) over 100 years

Massimo Terzi<sup>1\*</sup>, Nenad Jasprica<sup>2</sup>

<sup>1</sup>Institute of Bioscience and Bioresources (IBBR), National Council of Research (CNR), via Amendola 165/A, IT-70126, Bari, Italy

<sup>2</sup>University of Dubrovnik, Institute for Marine and Coastal Research, Kneza Damjana Jude 12, HR-20000 Dubrovnik, Croatia

**Abstract** – The changes in the grassland vegetation that have occurred over the last almost 100 years on the northeastern Adriatic island of Plavnik (Croatia) were studied. For this purpose, 29 phytosociological relevés of standard plot size were collected using the Braun-Blanquet phytosociological approach and compared with those of 100 years ago. With the aim of defining the differences among the communities and identifying the main environmental gradients, cluster analysis (Ward's method on a Chord distance matrix) and non-metric multidimensional scaling ordination were applied. Taxa score-weighted means of the Ellenberg-type indicator values of main environmental variables were used to assess ecological differences among communities, and disturbance-related indicator values were calculated to assess the disturbance regime in shaping plant communities. According to the syntaxonomic framework for the grassland associations, three associations and two subassociations were found. Among them the *Festuco valesiacae-Poetum bulbosae* (*Poetalia bulbosae*, *Poetea bulbosae*), the *Helichrysetum italici salvietosum officinalis* (*Scorzoneretalia villosae*, *Festuco-Brometea*), and the *Hedypnoido rhagadioloidis-Asphodeletum ramosi* were proposed as new. The *Festuco valesiacae-Poetum bulbosae* documents for the first time, based on phytosociological data, the occurrence of the *Poetea bulbosae* in Croatia. Three associations were considered extinct, possibly as a result of changes in grazing pressure and type.

**Keywords:** eastern Adriatic, grazing pressure, *Helichrysum italicum*, Mediterranean Islands, *Poetea bulbosae*, *Scorzoneretalia villosae*, syntaxonomy, vegetation changes

## Introduction

The Mediterranean basin is considered a biodiversity hotspot on account of its exceptional concentration of endemic species and the loss of habitats suitable for them (Myers et al. 2000). A significant component of this biodiversity is preserved in the Mediterranean islands due to the presence of steno-endemic taxa and distinctive plant assemblages (Médail 2017, Vargas 2020). The islands of the eastern Adriatic, with its multitude of archipelagos, are particularly important from this point of view, and they are characterised by a very high plant richness, comprising almost 1800 vascular taxa (Nikolić et al. 2008).

The phytosociological investigation of the north-eastern Adriatic using the approach of Braun-Blanquet (Westhoff and van der Maarel 1980) began in the 1930s, with the pioneering studies of Croatian botanist Stjepan Horvatić on some islands, such as Pag and Rab (Trinajstić 2001 and

references therein). On those islands, one of the most widespread aspects of vegetation was represented by open Mediterranean-sub-Mediterranean grasslands the particular floristic composition of which justified their attribution to an endemic alliance (*Chrysopogono grylli-Saturejion subspicatae* Horvat et Horvatić 1934 *nom. inval.*). Subsequently, that alliance was classified in an order (*Scorzonero villosae-Chrysopogonetalia grylli* Horvatić et Horvat in Horvatić 1963) and class also endemic to the Western Balkans (*Brachypodio-Chrysopogonetea* Horvatić 1963) (see Terzi 2011).

According to the current European syntaxonomic framework, the so-called EuroVegChecklist (EVC, Mucina et al. 2016), those grasslands mostly belong to the western Balkan alliances *Chrysopogono grylli-Koelerion splendidis* and *Scorzonerion villosae*, and to the trans-Adriatic order

\* Corresponding author e-mail: massimo.terzi@cnr.it

*Scorzoneretalia villosae*, included in turn in the class *Festuco-Brometea* (but see also Terzi et al. 2023).

The first survey – the subject of Horvatić's doctoral thesis (Trinajstić 2001) – concerned the vegetation of the island of Plavnik, in the Kvarner area (Croatia), where Horvatić (1927) described five grassland syntaxa, namely *Festucetum valesiacae*, *Asphodeletum microcarpi*, *Helichrysetum italici*, “*Helichrysetum salviosum*” and *Andropogonetum grylli*. For each of them, a diagnosis was provided consisting of a variable number (from 5 to 10) of quadrats of 1 square metre each, with a complete list of taxa accompanied by quantitative scores. Although each vegetation type was sampled on only a small area, Horvatić (1927) gave a fairly precise idea of the main dominant and codominant taxa present at that time.

Quite surprisingly, neither Horvatić's later papers (e.g. Horvatić 1963, 1973) nor the subsequent syntaxonomic revisions of Croatian grasslands of the order *Scorzoneretalia villosae* (e.g. Horvat et al. 1974, Trinajstić 2008, Terzi 2015) mentioned these five syntaxa. Moreover, the herbaceous vegetation of the island of Plavnik has never been studied subsequently.

This island presents a rather interesting situation from a vegetation point of view because it has remained uninhabited for a long period of time, being used as a hunting reserve and only occasionally visited by hunters and tourists (along the coastal areas). The conspicuous presence of animals, sheep and fallow deer, left in the wild, has presumably significantly increased the grazing pressure on the grassland ecosystems.

The present work therefore aims to (i) provide new relevés for the herbaceous vegetation of the island of Plavnik, using a more reliable plot size than that used by Horvatić (1927), (ii) assess the changes in vegetation that have occurred over almost 100 years, (iii) provide a syntaxonomic framework for the grassland associations occurring on the island, and (iv) identify the main ecological drivers of grassland diversity.

## Materials and methods

### Study area

The island of Plavnik (surface area 8.63 km<sup>2</sup>, coastline length 18.47 km) belongs to the north-eastern Adriatic group of islands in the Kvarner Bay, Croatia (Fig. 1). It is located between the islands of Cres and Krk and it stretches in a northwest-southeast direction for a length of 6.3 km. The coastline of the island is not indented: the northern coast is very steep and poorly accessible, while the southern is flat, gentle, accessible and with several coves.

The island has high terrain ruggedness. The northern and central part is a karstic plateau with an elevation of approximately 90–180 m a.s.l., bordered by steep rocky slopes exceeding 30°. The highest peak reaches an elevation of 193.6 m a.s.l. To the south-east, the karstic plateau gradually passes into lowlands with altitudes lower than 30 m a.s.l. (Fig. 2).

The bedrock consists mainly of carbonate rocks (Fuček et al. 2015). The Upper Cretaceous carbonate deposits can be recognized in different types of vertically alternating lithotypes. Multi-layered and massive recrystallized limestones with radiolithid-chondrodotic lithosomes are developed on both western and eastern coasts, while in the central part there are medium-thick layered shallow-sea mudstones and granular limestones with benthic foraminifera, radiolithid rudists and chondrodonts. The brown soils on limestone are predominant on the island (Vukadinović 2021).

The area is influenced by both Mediterranean and continental climates and characterized by intrusions of cold air during the winter and convective precipitation in the summer. These specific conditions result in the lack of a distinct dry period typical of the Mediterranean climate (Šegota and Filipčić 2003). The average annual air temperature is 15.3 °C and precipitation averages 1253.7 mm yr<sup>-1</sup> (data from the town of Krk station for 1998-2022, Croatian Meteorological and Hydrological Service). The highest monthly average



**Fig. 1.** The island of Plavnik is located between the islands of Cres and Krk in the Kvarner Bay (Croatia). The square on the map in the lower right corner shows the study area in the NE Adriatic context.



**Fig. 2.** Physiognomy of the plant associations on the island of Plavnik in May 2023: A – *Helichrysetum italici* Horvatić 1927 on the island's eastern coast, B – *Helichrysetum italici salvietosum officinalis subass. nov.* covers the open areas on the island's western slopes, C – *Hedypnoido rhagadioloidis-Asphodeletum ramosi ass. nov.*, in the southernmost part of the island, D – *Festuco valesiacae-Poetum bulbosae ass. nov.* occurs on the karstic plateau in the north-central part of the island.

temperature is 30.5 °C in July, and the lowest 2.5 °C in January. The absolute maximum of 38.8 °C was recorded on 21 July 2015, and the absolute minimum (-9.0 °C) on 24 January 2006. The greatest rainfall comes in September, October and November with averages of 155.6, 162.0 and 178.0 mm, respectively. In the period from June to August the total rainfall is 184.2 mm. Northern winds (NW, ENE, NE) prevail throughout the year. This area has 2521 hours of sunshine per year (data for the island of Rab station for 1998-2022). On average the relative air humidity is 70%.

Phytoclimatic indices were calculated according to Rivas-Martínez et al. (2011): annual positive temperature ( $T_p$ ) is 1,834; continentality index ( $I_c$ ) is 18.9; thermicity index ( $I_t$ ) is 238; ombrothermic index ( $I_o$ ) is 0.68. According to these indices, the neighbouring Krk station lies within the temperate macrobioclimate, meso-temperate thermotype and semiarid ombrotype.

The observed global and regional warming has been unequivocal since the middle of the 20<sup>th</sup> century (IPCC 2014) and is present along the eastern Adriatic coast (Branković et al. 2013). Climate change shows a tendency to manifest in drier conditions for the Adriatic (Patarčić et al. 2014, WBG 2021), which is a common feature for the broader European part of the Mediterranean (EEA 2019). However, for example, the precipitation change is regionally highly variable and further analyses are needed to define the assessment of climate change impacts at the regional and local scale (Gajić-Čapka et al. 2018).

From the phytogeographic point of view, the eastern and southwestern part of the island is covered by stands of the *Fraxino orni-Ostryion* alliance i.e. *Quercus pubescenti-Carpinetum orientalis* Horvatić 1939 association. The meso-Mediterranean evergreen vegetation of the *Fraxino orni-Quercion ilicis* alliance is developed only in the north-

western part of the island. However, woodlands have been significantly changed due to the long-term anthropogenic influences and nowadays only degraded vegetation types, i.e. maquis, garrigue and grasslands, may be found. Large areas of trees were felled between the two world wars to obtain open pastures, intended for sheep rearing. The island is classified among Croatia's Important Plant Areas (IPAs) and is part of the Natura 2000 ecological network (Topić and Vuković 2010, Official Gazette 2019).

The island has had no permanent residents or settlements during the last centuries. Between the two world wars, only a few herding groups lived on the island to provide living conditions for livestock, mainly sheep, while goats made up only 10-20% of the total number of head (Dumančić 1988). After the Second World War, such a simple nomadic way of life gradually faded away. The sheep population has drastically decreased, while goats, in general, have practically disappeared on the eastern coast of the Adriatic. There are no available data on the number of sheep in Plavnik over last century. In addition, the non-native European fallow deer (*Dama dama*) was introduced in 1995. According to the data of the local hunting association Farma Plavnik d.o.o. (personal communication), a number of the fallow deer, with regular annual shooting of ca. 50 nap, are stable and amounts to about 160 nap. The maximum nutritional requirements for females are during lactation, and this period coincides with the maximum development of vegetation.

#### Vegetation sampling and data analysis

The map of the distribution areas of the five types of plant communities on the island of Plavnik provided by Horvatić (1927: p. 44) was compared with the most recent satellite images available on Google Earth Engine, to

identify areas still covered by herbaceous vegetation. Those areas, and in particular the locations mentioned in that paper, were surveyed. The vegetation types were identified in the field based on their dominant and codominant taxa. Only for the “*Andropogonetum grylli*”, whose dominant taxon should have been *Chrysopogon gryllus*, we were unable to find any phytocoenosis clearly dominated by this plant, which is nowadays rather rare on the island.

The remaining four vegetation types were sampled through 29 relevés, made in May 2023 according to the phytosociological approach (see Westhoff and van der Maarel 1980). The relevés were sampled in quadrats of 25 m<sup>2</sup> (5 m × 5 m). All vascular plants in the plots were recorded and their cover-abundance values were visually estimated using the Braun-Blanquet cover-abundance scale with the subdivisions 2m, 2a and 2b as proposed by Barkman, Doing and Segal (in: van der Maarel 1979). This extended Braun-Blanquet scale (i.e. r, +, 1, 2m, 2a, 2b, 3, 4, 5) was replaced by the 9-point ordinal scale (i.e. 1-9, see Westhoff and van der Maarel 1980), before the statistical analyses. The main physical features of the plots (e.g., geographic coordinates, slope, exposure, vegetation cover, rockiness, and stoniness) were also recorded. Stoniness is defined here as the percentage of plot coverage by stones (incoherent material), regardless of their size. Rockiness, on the other hand, indicates the percentage of coverage of the plot determined by the outcropping rocky substrate.

The data matrix (29 relevés × 115 taxa) was subjected to hierarchical clustering using Ward's method on a Chord distance matrix (i.e., Euclidean distance on pre-transformed data through normalization by sites). Four interpretable clusters were subjectively defined by visual examination of the dendrogram.

To visualize the floristic and ecological relationships among the four clusters of relevés, nonmetric multidimensional scale (NMS) ordination was performed by using the Chord distance. Both cluster analysis and ordination were performed through the PcOrd 6.22 software package (McCune and Mefford 2011), the NMS with the "slow and thorough" option of the autopilot mode.

To assess ecological differences among the four plant communities, taxa score-weighted means of the Ellenberg-type Indicator Values (IVs) of temperature (T), soil reaction (R), moisture, and nutrient content were calculated for each relevé, based on the EIVs provided for Italy – which is the closest country geographically and ecologically to the study area – in Tichý et al. (2023).

In addition, given the presence of high numbers of sheep and fallow deer on the island of Plavnik and the possible importance of the disturbance regime in shaping the different plant communities, three additional disturbance-related IVs, taken from Midolo et al. (2023), were considered. In more detail, the following IVs, were calculated on taxa score-weighted means for each relevé: Disturbance severity (Ds), Disturbance frequency (Df), and Grazing pressure

(Gr). Joint plots were used to represent linear correlations between IVs and ordination scores, with an  $r^2$  cut-off of 0.3.

Indicator species analysis (ISA) (Dufrêne and Legendre 1997) was used to identify the indicator taxa of the four main clusters of relevés obtained by cluster analysis, and their combinations in larger groups (see De Cáceres et al. 2010). The indicator values were calculated for all taxa present in at least three relevés. The statistical significance ( $P < 0.01$ ) of the indicator values was tested by a Monte Carlo test with 10000 permutations. The ISA was run in R software (R Core Team 2023) through the package *indicpecies* 1.7.12 (De Cáceres and Legendre 2009).

In order to verify whether the relevés from the island of Plavnik in which *Asphodelus ramosus* was dominant (i.e. cluster 'He' in Figs. 3 and 4) could represent an association independent of the others described so far in the eastern Adriatic, we compared these relevés with the associations *Bromo erecti-Chrysopogonetum grylli* Horvatić 1934 and *Narcisso tazettae-Asphodeletum microcarpi* Šegulja 1969. In more detail, a second data matrix (55 relevés × 283 taxa) was obtained by adding the seven relevés reported in Tab. 3 to the other 48 relevés carried out by i) Horvatić (1934, rels. 11-23 of table XXVIII) on the island of Pag, Croatia; ii) Stanišić-Vujačić et al. (2022, rels. 1-17 of table 1) at Čemovsko polje, Montenegro, iii) Šegulja (1970, rels. 1-8 of table II) in Labin, Istria, Croatia, and iv) Hećimović (1984, *Narcisso tazettae-Asphodeletum microcarpi sisymbriosum officinalis* Hecimović 1984, rels. 1-10 of table 5) on the islets of Bobara and Mrkan, in southern Croatia.

Using the same methods as described above, an NMS ordination was performed on that matrix to visualise the floristic relationships between the relevés, and then an indicator species analysis was run to identify the taxa that distinguish the *Asphodelus ramosus* community on Plavnik Island from those elsewhere.

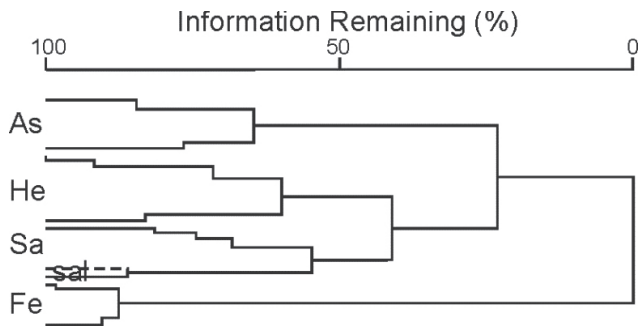
The taxonomic nomenclature follows Euro+Med (2006-2023). *Bromopsis erecta* and *B. condensata* were treated under the same tag, the latter being more abundant and frequent on the island of Plavnik.

Syntaxonomic nomenclature of alliances, orders and classes follows the EuroVegChecklist (<https://floraveg.eu/vegetation/>, retrieved on 10 October 2023, as modified by Terzi et al. 2024a), whose correct names are given without author citations. Syntaxon names not included in the EVC, but are instead provided with author citations. Names reported exactly as mentioned in the original papers are reported in quotation marks.

## Results

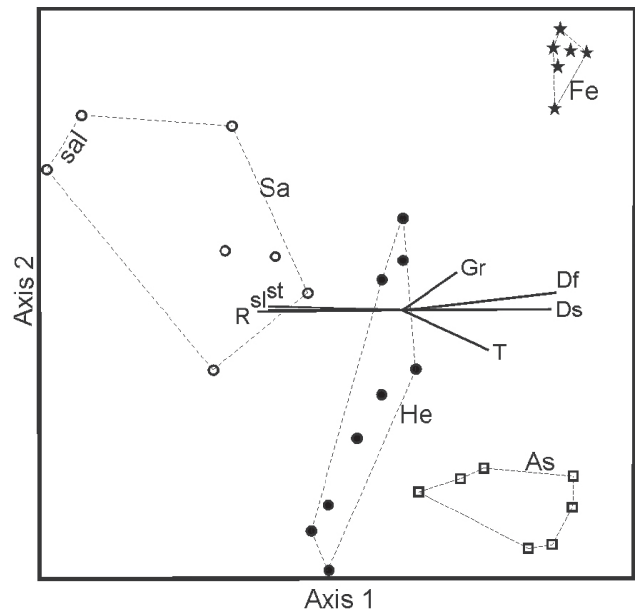
The dendrogram (Fig. 3) and further NMS (Fig. 4) showed four main clusters of relevés corresponding to the plant communities dominated by (Fe) *Festuca valesiaca* and *Poa bulbosa*, (As) *Asphodelus ramosus*, (He) *Helichrysum italicum*, and (Sa) *Salvia officinalis* and *Helichrysum italicum*.





**Fig. 3.** Hierarchical clustering (Ward's method, Chord distance) shows four main groups of relevés corresponding to the plant communities: As – *Hedypnoido rhagadioloidis-Asphodeletum ramosi*, He – *Helichrysetum italici*, Sa – *Helichrysetum italici salvietosum officinalis*, “sal” indicates a group of relevés clearly dominated by *Salvia officinalis*, Fe – *Festuco valesiaca-Poetum bulbosae*.

The cluster Fe groups together relevés from the summit plateau of the island, at more than 150 m a.s.l., where Horvatić (1927) had previously described the association *Festucetum valesiaca*. The mean number of taxa is 33 whereas in all the relevés of this cluster 53 taxa were found. In addition to *F. valesiaca* and *P. bulbosa* many species were recorded with high frequency (see Tab. 1).



**Fig. 4.** Non-metric multidimensional scaling of relevés in the dataset. Abbreviations: As – *Hedypnoido rhagadioloidis-Asphodeletum ramosi*, He – *Helichrysetum italici*, Sa – *Helichrysetum italici salvietosum officinalis*, “sal” indicates a group of relevés clearly dominated by *Salvia officinalis*, Fe – *Festuco valesiaca-Poetum bulbosae*. Ds – disturbance severity, Df – disturbance frequency, Gr – grazing pressure, T – temperature, R – soil reaction, st – stoniness, sl – slope.

**Tab. 1.** *Festuco valesiaca-Poetum bulbosae* ass. nov. on the island of Plavnik. Life-forms (LF): Ch – chamaephytes, G – geophytes, H – hemicryptophytes, P – phanerophytes, T – therophytes. Asterisk (\*) indicates holotypus (rel. 1).

Rel. number	LF	1*	2	3	4	5	6
Exposition (°)		0	0	0	0	0	0
Slope (%)		0	0	0	0	0	0
Rockiness (%)		30	40	30	60	40	20
Stoniness (%)		10	30	10	10	20	30
Vegetation cover (%)		70	60	65	50	60	70
<b><i>Festuco valesiaca-Poetum bulbosae</i></b>							
<i>Poa bulbosa</i>	H	6	6	6	5	6	4
<i>Festuca valesiaca</i>	H	5	6	6	5	6	6
<b><i>Poetalia bulbosae, Poetea bulbosae</i></b>							
<i>Trifolium suffocatum</i>	T	6	4	4	4	3	3
<i>Plantago lanceolata</i>	H	3	2	2	2	2	3
<i>Erodium cicutarium</i>	T	2	2	2	2	2	2
<i>Trifolium subterraneum</i>	T	3	2	.	2	3	2
<i>Trifolium scabrum</i>	T	2	2	2	2	.	2
<b><i>Stipo-Trachynietea distachyae</i></b>							
<i>Trifolium campestre</i>	T	4	4	5	5	2	5
<i>Filago germanica</i>	T	4	4	3	3	4	2
<i>Galium murale</i>	T	3	3	3	2	3	2
<i>Sherardia arvensis</i>	T	3	2	3	2	2	3
<i>Gastridium ventricosum</i>	T	2	2	3	2	2	3
<i>Sideritis romana</i>	T	2	2	2	3	3	2
<i>Catapodium rigidum</i>	T	2	2	2	2	2	2
<i>Ornithogalum collinum</i>	G	2	2	.	2	.	.
<i>Ononis reclinata</i>	T	2	.	.	2	.	.

Tab. 1. continued

Rel. number	LF	1*	2	3	4	5	6
<b>Tuberarietea guttatae</b>							
<i>Crepis neglecta</i> subsp. <i>neglecta</i>	T	2	3	2	2	2	2
<i>Vulpia ciliata</i>	T	2	2	3	3	2	2
<i>Medicago minima</i>	T	3	2	2	2	2	2
<i>Galium divaricatum</i>	T	.	2	3	2	2	2
<i>Aira elegantissima</i>	T	2	2	2	2	.	2
<i>Asterolinon linum-stellatum</i>	T	.	2	.	2	2	2
<i>Plantago bellardii</i>	T	.	.	3	3	2	.
<i>Minuartia hybrida</i>	T	4	2	.	2	.	.
<i>Lotus hispidus</i>	T	.	2	.	.	2	.
<b>Festuco-Brometea</b>							
<i>Koeleria splendens</i>	H	3	4	2	2	3	3
<i>Thymus longicaulis</i>	Ch	2	2	3	.	2	3
<i>Teucrium capitatum</i> subsp. <i>capitatum</i>	Ch	2	2	2	.	2	2
<i>Carduus nutans</i> subsp. <i>micropterus</i>	H	.	.	.	2	.	2
<i>Eryngium amethystinum</i>	H	.	.	.	2	.	2
<i>Salvia pratensis</i>	H	2	2	.	.	.	.
<b>Other taxa</b>							
<i>Petrorhagia saxifraga</i>	H	3	2	2	2	2	2
<i>Anagallis arvensis</i>	T	2	.	2	2	2	2
<i>Scorzoneroideis autumnalis</i>	H	2	2	2	.	.	2
<i>Cerastium pumilum</i> subsp. <i>glutinosum</i>	T	2	2	2	2	.	.
<i>Torilis nodosa</i>	T	2	2	.	.	2	.
<i>Geranium molle</i>	T	2	.	2	.	2	.
<i>Marrubium incanum</i>	H	.	.	2	2	2	.
<i>Medicago monspeliaca</i>	T	.	.	2	.	2	.
<i>Sedum acre</i>	Ch	2	.	2	.	.	.
<i>Euphorbia helioscopia</i>	T	2	.	.	.	2	.

Date, coordinates and sporadic species (with 2) of the relevés: Rel. 1 (holotypus): 28.V.2023, lat. 44° 58' 41", long. 14° 31' 27", *Arenaria serpyllifolia*, *Linum tenuifolium*; Rel. 2: 28.V.2023, lat. 44° 58' 42.5", long. 14° 31' 23", *Asparagus acutifolius*, *Brachypodium sylvaticum*, *Euphorbia exigua*, *Veronica arvensis*; Rel. 3: 28.V.2023, lat. 44° 58' 41", long. 14° 31' 21", *Euphorbia cyparissias*, *Helichrysum italicum*; Rel. 4: 28.V.2023, lat. 44° 58' 39", long. 14° 31' 23", *Valantia muralis*, *Achnatherum bromoides*, Rel. 5: 28.V.2023, lat. 44° 58' 38", long. 14° 31' 27.5", *Ficaria verna*; Rel. 6: 28.V.2023, lat. 44° 58' 35", long. 14° 31' 32", *Carex halleriana*.

The indicator species associated to this cluster are *Trifolium suffocatum*, *Erodium cicutarium*, *Crepis neglecta* subsp. *neglecta*, *Petrorhagia saxifraga*, and *Marrubium incanum*.

The second partitioning level separates the cluster As. This cluster represents the vegetation clearly dominated by *Asphodelus ramosus*, the latter being always recorded with a cover greater than 75%, except in one relevé. This vegetation type is well represented in the southern part of the island. The indicator taxa of this cluster are *Asphodelus ramosus*, *Hedypnois rhagadioloides*, *Crepis zacintha*, and *Centaureum maritimum*. The mean number of taxa per relevé is 32.

The third cluster He represents the most widespread type on the island: open vegetation dominated by *Helichrysum italicum* and *Koeleria splendens*, and with other taxa typical of the *Festuco-Brometea* class occurring with high frequency (e.g. *Bromopsis erecta*, *Euphorbia cyparissias*, Tab. 2).

The indicator species of this cluster are *Reichardia picroides*, *Bupleurum veronense*, *Plantago holosteum*, and *Hippocrepis comosa*. This vegetation type usually develops on rocky and stony ground whereas on steep slopes on the eastern part of the island, in some places, it gives way to a *Salvia officinalis*-*Helichrysum italicum* community, represented by the cluster Sa. The latter corresponds to the "*Helichrysetum salviosum*" described by Horvatić (1927) and includes seven relevés, with a mean number of taxa of 21. This cluster is actually heterogeneous with some relevés which represent a variant of the *Helichrysetum italicum* due to the presence and co-dominance of both *Helichrysum italicum* and *Salvia officinalis*, whereas two relevés (indicated as "sal" in Figs. 3 and 4) are clearly dominated by the latter. *Salvia officinalis* is the only indicator species of this cluster.

The NMS (Fig. 4) resulted in a two-axis solution, with a final stress 8.96, indicating a fairly good ordination.

**Tab. 2.** *Helichrysetum italici typicum* (rels. 1-10) and *Helichrysetum italici salvietosum officinalis* (rels. 11-14); transition to *Stipo-Salvietum officinalis* (rels. 15-16) on the island of Plavnik. For abbreviations of Life-forms [LF], see caption of Tab. 1. Asterisk (\*) indicates neotypus (rel. 3), and holotypus (rel. 13), respectively.

Rel. number	LF	1	2	3*	4	5	6	7	8	9	10	11	12	13*	14	15	16
Exposition (°)		20	50	0	130	230	50	140	0	160	70	160	150	270	50	270	60
Slope (%)		5	10	10	2	3	10	2	0	2	2	15	20	10	20	30	25
Rockiness (%)		30	40	40	10	70	30	20	10	40	40	40	40	30	30	30	30
Stoniness (%)		40	50	30	40	20	70	20	40	40	40	70	40	40	50	70	70
Vegetation cover (%)		60	60	70	60	60	70	60	70	70	75	50	55	60	60	70	60
<b><i>Helichrysetum italici</i></b>																	
<i>Helichrysum italicum</i>	Ch	7	7	8	6	6	7	7	8	7	7	6	6	6	5	2	2
<i>Salvia officinalis</i>	Ch	.	.	.	.	.	.	.	.	.	6	5	4	7	7	8	8
<b><i>Chrysopogono grylli-Koelerion splendidis, Scorzoneretalia villosae</i></b>																	
<i>Bromopsis condensata</i> (+ <i>Br. erecta</i> )	H	4	4	3	2	4	3	2	4	3	5	2	2	5	3	4	3
<i>Koeleria splendens</i>	H	5	5	5	4	5	4	3	4	5	4	.	.	2	2	.	.
<i>Festuca valesiaca</i>	H	.	.	2	2	.	.	2	2	4	3	.	.	3	2	.	.
<i>Bupleurum veronense</i>	T	2	2	2	2	2	2	.	.	.	.	.	.	.	.	.	.
<i>Carduus nutans</i> subsp. <i>micropterus</i>	H	.	.	.	.	2	.	.	.	.	.	.	.	2	2	2	.
<i>Centaurea tommasinii</i>	H	2	2	2	.	.	.	.	.	.	.	.	2	.	.	.	.
<i>Teucrium montanum</i>	Ch	2	.	.	.	.	.	.	.	.	2	2	.	2	.	.	.
<i>Aethionema saxatile</i>	Ch	.	.	.	.	.	.	.	.	.	.	2	2	2	.	.	.
<i>Medicago prostrata</i>	H	.	.	2	2	.	.	.	.	2	.	.	.	.	.	.	.
<i>Chrysopogon gryllus</i>	H	.	.	.	.	.	.	.	2	2	.	.	.	.	.	.	.
<i>Eryngium amethystinum</i>	H	.	.	.	.	.	.	.	.	.	2	.	.	.	2	.	.
<i>Genista sylvestris</i> subsp. <i>dalmatica</i>	Ch	2	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>Stipa eriocalis</i>	H	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	2
<b><i>Festuco-Brometea</i></b>																	
<i>Euphorbia cyparissias</i>	H	4	3	2	2	3	5	2	.	2	2	.	.	.	.	.	.
<i>Linum tenuifolium</i>	Ch	2	2	.	2	2	2	.	.	2	2	2	.	2	.	.	.
<i>Melica ciliata</i>	H	.	.	2	.	.	.	2	2	2	2	2	2	.	.	2	2
<i>Salvia pratensis</i>	H	2	.	3	2	3	2	2	2	.	2	.	.	.	.	.	.
<i>Sanguisorba minor</i>	H	2	2	2	.	2	3	2	2	.	2	.	.	.	.	.	.
<i>Sedum acre</i>	Ch	2	.	2	2	2	2	.	.	.	.	.	.	.	.	.	.
<i>Thymus longicaulis</i>	Ch	2	.	2	2	2	.	.	2	.	.	.	.	.	.	.	.
<i>Convolvulus cantabrica</i>	H	.	.	.	2	.	2	2	.	.	.	.	2	.	.	.	.
<i>Hippocrepis comosa</i>	H	2	2	.	2	.	2	.	.	.	.	.	.	.	.	.	.
<i>Plantago holosteum</i>	H	4	4	4	.	.	4	.	.	.	.	.	.	.	.	.	.
<i>Achnatherum bromoides</i>	H	.	.	.	.	.	.	2	2	.	.	.	.	.	.	.	.
<i>Carex halleriana</i>	H	.	.	.	.	.	.	.	.	.	.	.	2	4	.	.	.
<i>Centaureum erythraea</i>	H	.	.	2	2	.	.	.	.	.	.	.	.	.	.	.	.
<i>Dorycnium pentaphyllum</i> subsp. <i>herbaceum</i>	H	2	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>Galium corrudifolium</i>	H	.	2	.	.	.	.	.	.	.	.	.	.	.	.	.	.
<i>Knautia purpurea</i>	H	.	.	.	.	.	.	.	.	.	2	.	.	.	.	.	.
<i>Pilosella piloselloides</i>	H	.	.	.	.	.	.	.	.	.	.	.	.	2	.	.	.
<i>Teucrium chamaedrys</i>	Ch	.	.	.	.	.	.	.	2	.	.	.	.	.	.	.	.
<b><i>Stipo-Trachynietea distachyae</i></b>																	
<i>Catapodium rigidum</i>	T	3	2	2	2	2	3	2	3	3	2	2	2	2	3	.	.
<i>Trifolium campestre</i>	T	2	2	3	3	3	2	4	4	5	3	2	3	2	2	.	.
<i>Trachynia distachya</i>	T	3	2	3	3	3	2	2	2	3	2	4	3	2	.	.	.

Tab. 2. continued

Rel. number	LF	1	2	3*	4	5	6	7	8	9	10	11	12	13*	14	15	16
<i>Euphorbia exigua</i>	T	2	2	2	2	2	.	2	2	2	2	3	3	2	.	.	.
<i>Gastridium ventricosum</i>	T	2	2	2	2	2	2	3	2	2	2	.	2	2	.	.	.
<i>Sideritis romana</i>	T	2	.	2	5	2	2	2	2	2	2	2	2	.	2	.	.
<i>Medicago minima</i>	T	2	2	2	3	2	.	3	3	3	2	2	3	.	.	.	.
<i>Valantia muralis</i>	T	3	3	2	3	2	4	2	.	2	.	2	2	.	2	.	.
<i>Trifolium scabrum</i>	T	2	2	2	3	.	.	3	3	3	2	.	2	2	.	.	.
<i>Filago germanica</i>	T	.	.	2	2	2	.	2	2	.	.	.	.	.	.	2	.
<i>Sherardia arvensis</i>	T	.	.	2	2	.	.	2	3	2	.	.	.	.	2	.	.
<i>Asterolinon linum-stellatum</i>	T	.	2	2	2	.	.	.	.	2	.	.	2	.	.	.	.
<i>Ononis reclinata</i>	T	.	.	2	2	2	.	.	.	2	2	.	.	.	.	.	.
<i>Plantago bellardii</i>	T	.	.	2	3	3	.	.	2	.	.	.	.	.	.	.	.
<i>Crepis neglecta</i> subsp. <i>neglecta</i>	T	.	.	2	.	.	.	.	.	.	.	.	.	.	2	2	.
<i>Galium murale</i>	T	.	.	.	.	.	.	2	2	.	2	.	.	.	.	.	.
<i>Ornithogalum collinum</i>	G	2	2	.	.	.	.	.	2	.	.	.	.	.	.	.	.
<i>Valerianella eriocarpa</i>	T	.	.	2	2	.	.	2	.	.	.	.	.	.	.	.	.
<b>Other taxa</b>																	
<i>Dactylis glomerata</i> subsp. <i>hispanica</i>	H	2	2	2	2	.	3	2	3	2	3	.	3	2	2	3	.
<i>Linum trigynum</i>	T	3	2	2	2	2	2	2	2	2	3	2	.	3	2	.	.
<i>Anagallis arvensis</i>	T	3	2	2	2	2	2	2	2	2	2	2	2	.	.	.	.
<i>Carlina corymbosa</i>	H	2	.	2	2	2	2	.	2	2	2	.	.	2	.	.	.
<i>Reichardia picroides</i>	H	2	2	2	2	2	2	2	.	.	2	.	.	2	.	.	.
<i>Juniperus oxycedrus</i>	P	2	2	2	2	.	.	.	.	2	2	.	.	.	4	2	.
<i>Plantago lanceolata</i>	H	3	2	2	2	2	2	2	.	2	.	.	.	.	.	.	.
<i>Sonchus bulbosus</i> subsp. <i>bulbosus</i>	G	.	.	.	.	.	.	3	4	2	2	.	2	2	2	2	.
<i>Teucrium capitatum</i> subsp. <i>capitatum</i>	Ch	2	.	2	.	2	.	2	2	2	2	.	2	.	.	.	.
<i>Geranium purpureum</i>	T	.	.	2	.	.	.	2	2	2	2	.	.	2	2	.	.
<i>Orobanche minor</i>	T	.	.	.	2	2	.	2	2	2	.	.	.	2	.	.	.
<i>Torilis nodosa</i>	T	.	.	.	.	.	.	2	2	2	2	.	2	.	2	.	.
<i>Petrorhagia saxifraga</i>	H	.	.	.	2	.	.	2	2	2	.	.	2	.	.	.	.
<i>Poa bulbosa</i>	H	.	.	2	2	.	.	3	3	2	.	.	.	.	.	.	.
<i>Scorpiurus muricatus</i>	T	.	2	2	.	3	3	.	.	.	.	.	2	.	.	.	.
<i>Aira elegantissima</i>	T	.	.	2	.	.	.	2	2	.	.	.	.	.	.	.	.
<i>Allium</i> sp.	G	2	.	.	.	.	.	.	.	.	.	2	.	.	2	.	.
<i>Asparagus acutifolius</i>	P	.	.	.	.	.	.	2	.	2	.	.	.	2	.	.	.
<i>Cerastium pumilum</i> subsp. <i>glutinosum</i>	T	.	.	.	.	.	.	.	2	.	.	.	2	.	2	.	.
<i>Galium divaricatum</i>	T	.	.	.	2	.	.	.	2	2	.	.	.	.	.	.	.
<i>Lotus corniculatus</i>	H	2	2	2	.	.	.	.	.	.	.	.	.	.	.	.	.

Dates, coordinates and sporadic species (with 2): Rel. 1: 25.V.2023, lat. 44° 58' 28", long. 14° 32' 25", *Euphorbia peplus*, *Thesium divaricatum*; Rel. 2: 25.V.2023, lat. 44° 58' 13", long. 14° 32' 36", *Thesium divaricatum*, *Anthyllis vulneraria* subsp. *rubriflora*; Rel. 3 (neotypus): 25.V.2023, lat. 44° 58' 00", long. 14° 32' 42", *Asphodelus ramosus*, *Hedypnois rhagadioloides*, *Leontodon crispus*, *Pilosella officinarum*, *Convolvulus althaeoides* subsp. *tenuissimus*; Rel. 4: 25.V.2023, lat. 44° 57' 54", long. 14° 32' 47", *Asphodelus ramosus*, *Crepis zacintha*, *Hedypnois rhagadioloides*, *Leontodon tuberosus*, *Crepis foetida*; Rel. 5: 25.V.2023, lat. 44° 57' 50", long. 14° 32' 56", *Pilosella officinarum*; Rel. 6: 25.V.2023, lat. 44° 57' 55", long. 14° 32' 54", *Lotus hispidus*, *Rubus ulmifolius*; Rel. 7: 27.V.2023, lat. 44° 58' 04", long. 14° 31' 30", *Clematis vitalba*, *Vulpia ciliata*, *Oxalis corniculata*, *Quercus ilex*, *Stachys officinalis*; Rel. 8: 27.V.2023, lat. 44° 58' 09.5", long. 14° 31' 15", *Clematis vitalba*, *Cynosurus echinatus*, *Euphorbia peplus*, *Geranium molle*, *Minuartia hybrida*, *Vulpia ciliata*, *Trifolium suffocatum*, *Ficaria verna*; Rel. 9: 27.V.2023, lat. 44° 58' 13.5", long. 14° 31' 02", *Paliurus spina-christi*, *Scorzoneroideis autumnalis*; Rel. 10: 28.V.2023, lat. 44° 58' 49", long. 14° 31' 42", *Crepis zacintha*, *Minuartia hybrida*, *Paliurus spina-christi*, *Sonchus asper*, *Carthamus lanatus*; Rel. 11: 26.V.2023, lat. 44° 57' 27.5", long. 14° 31' 59", *Urospermum picroides*; Rel. 12: 26.V.2023, lat. 44° 57' 28", long. 14° 31' 55", *Cynosurus echinatus*; Rel. 13 (holotypus): 27.V.2023, lat. 44° 58' 11", long. 14° 31' 13", Rel. 14: 28.V.2023, lat. 44° 58' 42", long. 14° 31' 38", *Geranium molle*; Rel. 15: 27.V.2023, lat. 44° 58' 06.5", long. 14° 31' 07", *Sesleria autumnalis*, *Sonchus asper*, *Myosotis arvensis*, *Phillyrea latifolia*; Rel. 16: 28.V.2023, lat. 44° 58' 39", long. 14° 31' 39", *Clematis flammula*.

The two axes accounted for 79.5 of the total variation (axis 1 = 65.3%, axis 2 = 14.2%). The four clusters occupy different parts of the ordination space without overlapping each other. Axis 1 separates the clusters He and Sa among them and from Fe and As. This axis is positively correlated with Df, Ds, Gr and temperature whereas it is negatively correlated with slope, stoniness and reaction. The correlation strengths of the EIVs of moisture and nutrient content with the ordination axes were lower than the selected  $r^2$  cut-off and we will not comment on those variables further. Therefore, the Sa cluster groups relevés taken on steeper slopes than the other clusters and with high percentages of stones. The Fe and As clusters have the highest values of disturbance, in terms of both severity and frequency. The lowest reaction values for these two clusters are probably related to disturbance. Axis 2 clearly separates Fe from As, the former with greater values of grazing, the second with greater values of temperature, being in the southern part of the island at the lowest altitude and mostly on the south-facing side.

The NMS ordination of the relevés of the As cluster together with those of the associations *Bromo erecti-Chrysopogonatum grylli* and *Narcisso tazettae-Asphodeletum microcarpi* resulted in a three-axis solution with a final stress of 7.5 (Fig. 5).

Most of the variance is represented by axis 1 (55.1 %) whereas axes 2 and 3 account for 22.6 % and 2.5 %, respectively. The relevés of the two associations *Bromo erecti-Chrysopogonatum grylli* (BC1 and BC2 in Fig. 5) and *Narcisso tazettae-Asphodeletum microcarpi* (NA1 and NA2) are separated on axis 1, whereas the relevés of the cluster As are in the middle. The latter are clearly separated from the others on axis 3 (Fig. 5). Interestingly, the relevés of the two phytosociological tables of the *Bromo erecti-Chrysopogonatum*

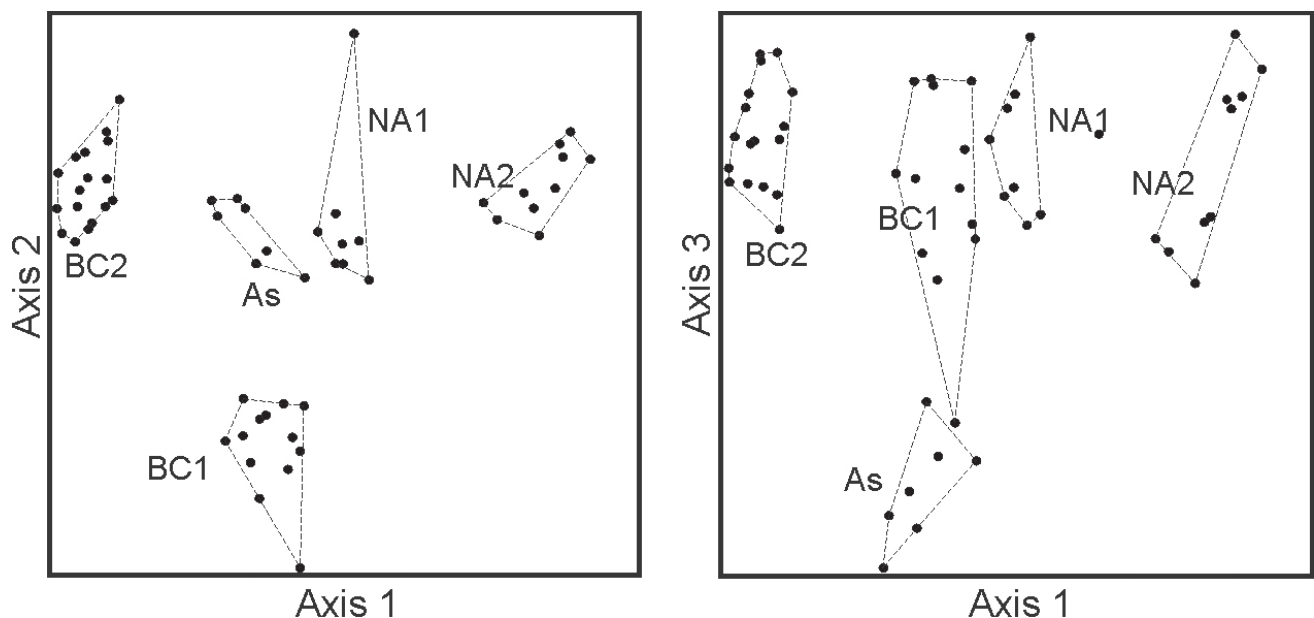
*grylli* (i.e. BC1 and BC2) and those of the two tables of the *Narcisso tazettae-Asphodeletum microcarpi* (i.e. NA1 and NA2) form four well separated groups of relevés.

The cluster As was found to be well distinguished in the species-space from the other associations with which it has no overlap. The indicator species analysis indicated the following 11 taxa as diagnostic for the As cluster (indicator values in brackets): *Hedypnois rhagadioloides* (74), *Linum trigynum* (72.3), *Galium murale* (70.6), *Euphorbia cyparissias* (68.9), *Geranium purpureum* (67.2), *Sideritis romana* (62.4), *Vulpia ciliata* (60.8), *Centaureum maritimum* (56.3), *Plantago bellardii* (56.3), *Scorpiurus muricatus* (56.3), and *Lotus hispidus* (42.3).

## Discussion

A direct quantitative comparison of Horvatić's data with the new data is not completely possible because the plot sizes used in the two surveys are too different (see also Dengler et al. 2009). For example, the five plots used by Horvatić (1927) to represent the *Festucetum valesiaca* sample a total area of 5 m<sup>2</sup> and include 17 taxa. The new dataset comprises six relevés of the same area – each with a plot size of 25 m<sup>2</sup> and containing an average of 33 taxa – for a total number of taxa of 53. Furthermore, the quantitative values of the taxa in Horvatić's article are estimated on a scale comprising at least 10 (undefined) values (-, -1, 1, 1-2, 2, 2-3, 3, 3-4, 4, 5), which are not directly comparable with the 9-points scale used in this paper. However, on the basis of qualitative considerations, it can be deduced whether the plant communities remained fairly similar or not during the 100 years.

As written above, although Horvatić (1927) described five grassland syntaxa on the island of Plavnik, we were on-



**Fig. 5.** Non-metric multidimensional scaling (with axes 1, 2 and 3) of eastern Adriatic associations with *Asphodelus ramosus*. Abbreviations: HA – *Hedypnois rhagadioloides-Asphodeletum ramosi*, BC – *Bromo erecti-Chrysopogonatum grylli* (BC1 from island of Pag, Croatia, and BC2 from Montenegro), NA – *Narcisso tazettae-Asphodeletum microcarpi* (NA1 from Istria and NA2 from islands of Bobara and Mrkan, Croatia).

ly able to find four grassland community types. In fact, the area originally covered by *Andropogonetum grylli* is completely occupied by a plant community dominated by *Asphodelus ramosus*, whereas *Chrysopogon gryllus* – i.e. the dominant taxon of the *Andropogonetum grylli* – is quite rare on the island.

### *Festuco valesiaca*-*Poetum bulbosae*

The *Festucetum valesiaca* described by Horvatić (1927) for the continuous herbaceous expanses on the summit of the island is no longer present. This association was dominated by *Festuca valesiaca*, followed as cover by *Bromopsis erecta* and *Plantago holostium*, and included several other species of the class *Festuco-Brometea* class (e.g. *Thymus longicaulis*, *Chrysopogon gryllus*, *Koeleria splendens*), to which it clearly belonged. In that area, the physiognomic situation appears quite different nowadays, with herbaceous phytocoenoses interspersed with shrubs and trees of *Juniperus oxycedrus*, generally on flat surfaces, where sheep and fallow deer find shelter (Fig. 2D). The herbaceous vegetation is dominated by *Poa bulbosa* and *F. valesiaca*, followed by some *Trifolium* species (*T. campestre*, *T. suffocatum*, *T. subterraneum*, *T. scabrum*), and *Koeleria splendens*. The results show that this plant community is clearly distinct from the others in terms of floristic composition and ecology. Indeed, it is the most disturbed in terms of intensity and frequency of disturbance (Ds and Df in Fig. 3) together with *Asphodelum ramosi* but, unlike the latter, it has a higher grazing pressure (Gr). Therefore, the plant community dominated by *Poa bulbosa* and *Festuca valesiaca* (Fe in Fig. 3) is here considered to be a new association, the *Festuco valesiaca*-*Poetum bulbosae* *ass. nov. hoc loco* (holotypus rel. 1 in Tab. 1), possibly derived from *Festucetum valesiaca* – of which *Festuca valesiaca*, *Koeleria splendens*, and other taxa are still present – as a consequence of the prolonged stay of sheep and deer and intensive grazing.

The presence with high cover and/or frequency of *Poa bulbosa*, *Trifolium suffocatum*, *T. subterraneum*, *Erodium cicutarium*, *Plantago lanceolata*, and other species, indicates that the *Festuco valesiaca*-*Poetum bulbosae* belongs to the *Poetea bulbosae*, i.e. a class representing heavily grazed pastures, which are mainly present in the western Mediterranean basin (e.g. Rivas Goday and Rivas-Martínez 1963). This class comprises a single order, *Poetalia bulbosae*, and several alliances, none of which occur in Croatia (Preislerová et al. 2022), with the exception of the *Romuleion* (Škvorc et al. 2017). This alliance was originally invalidly described by Oberdorfer (1954, see Terzi et al. 2024b) for the south and southeast of the Balkan Peninsula, and was identified by 20 character species, none of which are present in our relevés (at least in the phenological stage of vegetation at the time of sampling). The alliance was originally classified in the class “*Thero-Brachypodietea*” (Oberdorfer 1954), and then moved to the “*Brachypodio-Chrysopogonetea*” (Horvat et al. 1974), “*Saginetea maritimae*” (Rodwell et al. 2002), and *Tuberarietea guttatae* (Čarni et al. 2014). The EVC has instead classified the *Romuleion* in the *Poetea*

*bulbosae* (EVC) to represent “Macedonian seasonal perennial pastures on acidic substrates”. The *Poetea bulbosae*, indeed, are most frequently found on acid substrates (Oberdorfer 1954, Čarni et al. 2014).

The numerous taxa of the *Stipo-Trachynietea distachyae* found in the *Festuco valesiaca*-*Poetum bulbosae* are linked to the basic substratum of the island of Plavnik, while those of the *Tuberarietea guttatae* highlight the dynamic relationships with the oligotrophic grasslands of this class, where the *Romuleion* was framed by some authors (e.g. Čarni et al. 2014). Two associations of the *Poetalia bulbosae* were recently described near Podgorica, Montenegro, on fluvio-glacial deposits and calcareous rocky substrate and framed in the *Romuleion* (Stanišić-Vujačić et al. 2023). The floristic composition of those associations, however, includes only three character species of this alliance (*Romulea linairesii* subsp. *graeca*, and the very rare *Hypochaeris cretensis* and *Hedypnois rhagadioloides*). The associations from Montenegro were found in an area where rocky grasslands of the alliance *Chrysopogono grylli-Koelerion splendentis* are widespread and indeed at least one of those associations (*Romuleo bulbocodii-Poetum bulbosae* Stanišić-Vujačić et al. 2023) is characterized by the presence of numerous *Festuco-Brometea* species (Stanišić-Vujačić et al. 2022, 2023). The *Festuco valesiaca*-*Poetum bulbosae*, derived from the *Festucetum valesiaca*, also has dynamic relationships with that alliance and in fact shares several species (e.g., *Koeleria splendens*, *Thymus longicaulis*) with the Montenegrin associations. However, the natural potential vegetation of the Plavnik area is different from that of the surrounding of Podgorica, in Montenegro.

The floristic relationships of the *Festuco valesiaca*-*Poetum bulbosae* with the *Romuleion* are, however, even weaker, given that none of the character species of this alliance were detected in our relevés. For this reason, the *Festuco valesiaca*-*Poetum bulbosae* is only provisionally classified in the *Romuleion*, pending the expansion of knowledge about the variability of the *Poetalia bulbosae* in the Balkans by new phytosociological studies. It is indeed plausible to admit that a second alliance of the *Poetalia bulbosae*, in addition to the *Romuleion*, is present in the Western Balkans and that it is dynamically linked to a degradation of the grasslands of the *Scorzoneretalia villosae* (not present in Greece, where the *Romuleion* was originally described), but a large-scale comparison is needed to confirm this hypothesis, which is beyond the scope of this paper. In any case, the *Festuco valesiaca*-*Poetum bulbosae* documents for the first time, based on phytosociological data, the presence of *Poetalia bulbosae* and *Poetea bulbosae* in Croatia.

### *Helichrysetum italicum*

The most frequent vegetation type on the island is represented by the *Helichrysum italicum* dominated plant community (“He” in Figs. 3 and 4), which grows on rocky and stony substrates throughout the island. *Helichrysum italicum* is widespread in southern Europe with several subspecies and the nominal one, *Helichrysum italicum* subsp. *italicum*,

which is the only one found on the island of Plavnik, has a range in Europe that includes southern France, Italy and the Balkans (Euro+Med 2023). This subspecies grows in different types of vegetation, such as thermophilic scree communities of the *Scrophulario-Helichrysetalia* (Brullo and Spampinato 1991), and coastal sub-aerohaline dwarf scrub vegetation of the *Helichrysetalia italici* (Foucault 2020). In the Balkans, it has been considered a character taxon of the “*Brachypodio-Chrysopogonetea*” (Horvatić 1963, Horvat et al 1974). *Helichrysum italicum* is a suffruticose chamaephyte and, where it is dominant, the vegetation physiognomy takes on the appearance of a chamaephytic garrigue (Horvatić 1927). In the Dinaric Arc mountain range, this type of vegetation is often rich in many species typical of grasslands, so that it is not always possible to distinguish chamaephytic garrigues from 'pure' grasslands from a syntaxonomic point of view. In fact, the alliance *Chrysopogono grylli-Koelerion splendentis*, of the order *Scorzoneretalia villosae*, includes vegetation types dominated by both hemicryptophytes and dwarf chamaephytes, such as *Helichrysum italicum* and *Salvia officinalis*.

The same situation can be observed on the island of Plavnik where the *Helichrysetum italici* includes *Helichrysum italicum*, as the dominant, together with many grassland taxa, such as *Bromopsis condensata*, *Trifolium campestre*, *Dactylis glomerata* subsp. *hispanica*, *Salvia pratensis*, *Euphorbia cyparissias*, *Koeleria splendens*, *Hippocrepis comosa*. This plant community grows on incoherent substrata with a floristic composition that resembles that sampled by Horvatić (1927) a hundred years ago, except for the absence of *Chrysopogon gryllus*, which has evidently become rather rare on the island (Fig. 2A). Although the total number of species found in the new relevés is far higher than that recorded by Horvatić (1927), this difference is almost certainly due to the different sizes of the plots used in the two sampling surveys. The species that shape the physiognomy of the *Helichrysetum italici* are *Helichrysum italicum*, *Bromopsis condensata*, *Koeleria splendens*, and *Trifolium campestre*. The association clearly belongs to the *Chrysopogono grylli-Koelerion splendentis* due to the presence of characteristic species of the class *Festuco-Brometea* (e.g. *Bromopsis condensata*, *Euphorbia cyparissias*, *Linum tenuifolium*, *Festuca valesiaca*, *Hippocrepis comosa*), of the order *Scorzoneretalia villosae* and of the alliance (e.g. *Thymus longicaulis*, *Centaurea tommasinii*, *Plantago holosteum*, *Bupleurum veronense*) (see Terzi 2015). Although *Helichrysum italicum* is quite common in other associations of this alliance, the specific floristic composition of the *Helichrysetum italici* is different from that of all the other associations already described. In any case, even if this association was considered as a syntaxonomic synonym of another association belonging to the alliance, the name *Helichrysetum italici* would retain priority over names described later.

The *Helichrysetum italici* occurs in its typical aspect (subass. *typicum*, represented by rels. 1-9, in Tab. 2) on most of the island, and only on some spots does it associate with *Salvia officinalis* to give rise to the subassociation described

in the next section. The typical aspect corresponds to that described by Horvatić (1927) with 10 plots of 1 square meter each. The size of these individual plots thus appears to be much smaller than that normally used to sample the plant communities of the *Scorzoneretalia villosae*, which varies in 90% of cases between 10 and 150 square meters (Terzi 2015). Because of the small area of the plots sampled by Horvatić (1927), and perhaps also because of the omission of some species, each of those plots counts only a very limited number of taxa. Moreover, the quantitative values associated with these taxa are on a scale hardly comparable with those routinely used in modern phytosociological studies.

Although none of the 10 relevés originally published by Horvatić (1927), if taken alone, can be considered a representative relevé of the *Helichrysetum italici* for the reasons expressed above, the association is nevertheless validly described because at least the synoptic column of the association, i.e. the last column of table IV (“*Skrizaljka IV*”, on p. 33, in Horvatić 1927), constitutes a sufficient original diagnosis according to Article 7 of the International Code of Phytosociological Nomenclature (ICPN, Theurillat et al. 2021).

Therefore, we do not designate a lectotype, which would be incomplete and unrepresentative of the typical aspect of the association. Thus, not having a suitable relevé of the original diagnosis to be selected as lectotype of the association (see also Def. VIII of the ICPN), we find it more correct and useful to designate the relevé 3 in Tab. 2 as neotypus hoc loco of the *Helichrysetum italici* and of its subassociation *typicum*.

#### ***Helichrysetum italici salvietosum officinalis***

Along some steep, mainly north-facing slopes on incoherent substrates or on limestone outcrops, the typical aspect of the *Helichrysetum italici* gives way to another type of vegetation where *Salvia officinalis* becomes dominant or co-dominant with *Helichrysum italicum* (Fig. 2B). This aspect was erroneously described by Horvatić (1927) as “*Helichrysetum salviosum*” (Art. 3e of the ICPN, Theurillat et al. 2021), to indicate a subtype of the *Helichrysetum italici*. Its floristic composition indeed retains many species of the *Helichrysetum italici* with the addition of a few others, including the dominant *Salvia officinalis*, thus forming the subassociation *Helichrysetum italici salvietosum officinalis subass. nov. hoc loco* (holotypus rel. 13 of Tab. 2). This subassociation has fewer species than the typical one.

The grassland vegetation of the Western Balkans dominated by *Salvia officinalis* belongs to different vegetation types, mainly framed in the alliances *Chrysopogono grylli-Koelerion splendentis* and *Saturejion subspicatae* (Terzi et al. 2023). In our case, the *Helichrysetum italici salvietosum officinalis* definitely belongs to the first alliance and is perhaps transitional to the *Stipo-Salvietum officinalis*, whose nomenclatural type was recorded on the island of Unije, also located in Kvarner, about 50 km (as the crow flies) from Plavnik. In particular, two relevés could belong to this association (subcluster 'sal' in Fig. 3), which show the lowest

number of taxa, the clear dominance of *Salvia officinalis*, and the presence of *Helichrysum italicum* with very low coverage values.

**Hedypnoido rhagadioloidis-Asphodeletum ramosi**

The vegetation with *Asphodelus ramosus* develops mainly in the southern part of the island, with large and surprising expanses absolutely dominated by this species, with a cover usually greater than 90% (Fig. 2C). Two relevés recorded in the vicinity of the *Helichrysetum italicum* distribution area represent a transition towards this association and indeed there *Helichrysum italicum* is the second dominant species with a cover of 25-50%. A very different situation is described by Horvatić (1927) who reports local patches of *Asphodeletum microcarpi* interspersed in a matrix of other associations (*Andropogonetum grylli* and *Helichrysetum italicum*) and sharing several species with them. He notes that this sharing of species between the *Asphodeletum microcarpi* and the other associations is even more evident during the winter, when *Asphodelus ramosus*, the main dominant species, survives belowground. In the plots sampled by Horvatić (1927), *Asphodelus ramosus* was recorded with a score of “2” (or “3” in only one case) together with other species with

similar or greater cover, so that some of its plots look like local variants of the *Andropogonetum grylli*.

On the other hand, as written above, during our surveys on the island we found few plants of *Chrysopogon gryllus* for which it was not possible to identify the *Andropogonetum grylli* nor transitions between this association and the *Asphodeletum microcarpi*. In the most typical situation, *Asphodelus ramosus* is by far the dominant species, and with the exception of the cases representing a transition with the *Helichrysetum italicum* (rels. 5 and 6 in Tab. 3), in all other relevés the second dominant species never scored higher than 2a (i.e. up to 12% cover).

This plant community comprises numerous annual taxa of the classes *Stipo-Trachynietea distachyae* and *Tuberarietea guttatae*. The presence of so many annual plants is not a novelty for this kind of vegetation and has been observed in many other areas of the Mediterranean (see Terzi 2023 and references therein). The plant community dominated by *Asphodelus ramosus* was found to be the most disturbed grassland type on the island, together with the *Festuco valesiacae-Poetum bulbosae*. The thriving of *Asphodelus ramosus* is usually associated with intense grazing pressure and frequent fires. These factors might also be the ecologi-

**Tab. 3.** *Hedypnoido rhagadioloidis-Asphodeletum ramosi* ass. nov. on the island of Plavnik. For abbreviations of Life-forms [LF], see caption of Tab. 1. Asterisk (\*) indicates holotypus (rel. 2).

Rel. number	LF	1	2*	3	4	5	6	7
Exposition (°)		90	0	0	40	0	0	0
Slope (%)		5	0	0	2	0	0	0
Rockiness (%)		3	10	0	0	20	20	10
Stoniness (%)		3	20	5	0	40	30	10
Vegetation cover (%)		90	100	100	95	98	85	90
<b><i>Hedypnoido rhagadioloidis-Asphodeletum ramosi</i></b>								
<i>Asphodelus ramosus</i>	G	9	9	9	9	9	7	9
<i>Helichrysum italicum</i>	Ch	2	3	2	3	7	7	5
<i>Crepis zacintha</i>	T	.	4	3	3	2	.	2
<i>Hedypnois rhagadioloides</i>	T	2	2	2	2	2	2	.
<i>Centaureum maritimum</i>	T	.	.	2	3	.	2	2
<b><i>Stipo-Trachynietea distachyae</i></b>								
<i>Gastridium ventricosum</i>	T	2	3	4	3	4	2	3
<i>Trifolium campestre</i>	T	2	2	3	3	3	2	3
<i>Trifolium scabrum</i>	T	3	3	2	2	2	.	3
<i>Sherardia arvensis</i>	T	2	2	2	2	2	2	2
<i>Catapodium rigidum</i>	T	3	2	2	.	2	3	2
<i>Sideritis romana</i>	T	2	.	2	.	2	5	2
<i>Filago germanica</i>	T	2	2	2	3	.	.	2
<i>Galium murale</i>	T	2	2	3	2	.	.	2
<i>Trachynia distachya</i>	T	.	2	.	.	3	3	3
<i>Medicago minima</i>	T	2	.	.	.	2	3	2
<i>Euphorbia exigua</i>	T	.	.	2	2	2	.	2
<i>Trifolium subterraneum</i>	T	2	.	2	2	.	.	.
<i>Valantia muralis</i>	T	.	2	.	.	.	3	.



Tab. 3. continued

Rel. number	LF	1	2*	3	4	5	6	7
<b>Tuberarietea guttatae</b>								
<i>Linum trigynum</i>	T	2	3	3	2	2	2	2
<i>Galium divaricatum</i>	T	2	2	3	3	2	.	2
<i>Plantago bellardii</i>	T	2	2	2	2	2	.	2
<i>Vulpia ciliata</i>	T	5	2	4	3	.	.	2
<i>Scorpiurus muricatus</i>	T	2	2	2	.	2	2	.
<i>Aira elegantissima</i>	T	.	3	3	3	.	.	.
<i>Lotus hispidus</i>	T	4	2	3	.	.	.	.
<b>Festuco-Brometea</b>								
<i>Euphorbia cyparissias</i>	H	5	4	4	2	2	.	3
<i>Sanguisorba minor</i>	H	2	2	2	2	2	2	2
<i>Festuca valesiaca</i>	H	3	2	2	.	2	2	2
<i>Linum tenuifolium</i>	Ch	2	2	.	2	2	2	2
<i>Bromopsis condensata</i> (+ <i>Br. erecta</i> )	H	.	.	.	2	2	4	2
<i>Convolvulus cantabrica</i>	H	.	2	2	.	2	2	.
<i>Koeleria splendens</i>	H	.	2	.	.	3	2	.
<i>Salvia pratensis</i>	H	.	2	.	3	.	2	.
<i>Thymus longicaulis</i>	Ch	.	.	.	.	2	2	2
<i>Galium corrudifolium</i>	H	.	.	.	.	2	4	.
<i>Bupleurum veronense</i>	T	.	.	.	.	.	2	.
<i>Medicago prostrata</i>	H	.	.	.	.	.	.	2
<i>Chrysopogon gryllus</i>	H	.	.	.	.	2	.	.
<i>Melica ciliata</i>	H	.	.	.	.	.	.	2
<b>Other taxa</b>								
<i>Dactylis glomerata</i> subsp. <i>hispanica</i>	H	4	.	2	2	2	3	4
<i>Anagallis arvensis</i>	T	2	3	2	2	2	2	2
<i>Poa bulbosa</i>	H	3	.	2	2	2	2	2
<i>Geranium purpureum</i>	T	.	2	2	2	2	2	2
<i>Teucrium capitatum</i> subsp. <i>capitatum</i>	Ch	2	2	.	.	2	2	2
<i>Plantago lanceolata</i>	H	.	.	.	2	.	2	.
<i>Orobancha minor</i>	T	.	.	.	.	.	2	2

Date, coordinates and sporadic species (with 2, except where indicated) of the relevés: Rel. 1: 25.V.2023, lat. 44° 58' 02", long. 14° 32' 41", *Daucus carota*; Rel. 2 (holotypus): 26.V.2023, lat. 44° 57' 26", long. 14° 33' 01", *Carlina corymbosa*; Rel. 3: 26.V.2023, lat. 44° 57' 28.5", long. 14° 32' 59"; Rel. 4: 26.V.23, lat. 44° 57' 36", long. 14° 32' 58", *Scorzoneroides autumnalis* (3); Rel. 5: 26.V.2023, lat. 44° 57' 31", long. 14° 32' 51"; Rel. 6: 25.V.2023, lat. 44° 57' 51", long. 14° 32' 52", *Euphorbia peplus*, *Leontodon tuberosus*, *Ononis reclinata*, *Anisantha madritensis*, *Asterolinon linum-stellatum*, *Sedum acre*; Rel. 7: 26.V.2023; lat. 44° 57' 33", long. 14° 32' 42", *Urospermum dalechampii*.

cal drivers on the island of Plavnik, where we hypothesise that at least one of them (grazing or fires) has presumably occurred on the island with greater intensity and/or frequency than was the case 100 years ago. Increased grazing pressure is plausible due to the high number of sheep easily observed on that side of the island, while it is more difficult to assess the role of fires as data on their intensity and frequency are not available.

Thus, the present plant community dominated by *Asphodelus ramosus* on Plavnik Island differs both floristically and ecologically from the *Asphodeletum microcarpi* described by Horvatić (1927). It is also clearly different from the *Bromo erecti-Chrysopogonetum grylli* (i.e. *Asphodelo-Chrysopogonetum grylli* Horvatić 1963, cf. Terzi 2015) and *Narcisso tazettae-Asphodeletum microcarpi*. In this case, the Plavnik plant community differs in physiognomy, given the

clear dominance of *Asphodelus ramosus*, and floristic composition (Fig. 5). The indicator species that distinguish the plant community of Plavnik Island from those of other locations in the eastern Adriatic are mainly annual species, some of which indicate a slightly acidic to almost neutral substrate (e.g. *Centaureum maritimum*, *Linum trigynum*), as is to be expected in heavily grazed karst areas.

In view of the above, the plant community dominated by *Asphodelus ramosus* on the island of Plavnik is considered here a new association: *Hedypnoido rhagadioloidis-Asphodeletum ramosi* ass. nov. hoc loco (holotypus rel. 2 of Tab. 3). However, its classification at a higher syntaxonomic rank is rather difficult (see Terzi 2023 and references therein). Biondi et al. (2016) proposed a new class, order and alliance to classify the vegetation dominated by *Asphodelus ramosus* (and other geophytes) but they have not been ac-

cepted because a comprehensive syntaxonomic review on the subject is still lacking (Biurrun and Willner 2020, Terzi 2023). In Montenegro, *Asphodelus ramosus* is included within communities of the *Artemisietea vulgaris*, although it is most abundant in the *Bromo erecti-Chrysopogonetum grylli* (Stanišić-Vujačić et al. 2022). As a result of this as yet undefined syntaxonomic framework, we refrain from classifying the new association into one of the accepted classes.

With the exception of *Festucetum valesiacaе*, *Andropogonetum grylli* and *Asphodeletum microcarpi*, which can be considered extinct, the other plant communities on Plavnik Island can be classified as in the following syntaxonomic scheme:

*Festuco-Brometea* Br.-Bl. et Tx. ex Soó 1947

*Scorzoneretalia villosae* Kovačević 1959

*Chrysopogono grylli-Koelerion splendidis*  
Horvatić 1973

*Helichrysetum italici* Horvatić 1927 *typicum*

*Helichrysetum italici salvietosum officinalis*  
*subass. nov. hoc loco*

*Poetea bulbosae* Rivas Goday et Rivas-Mart. in  
Rivas-Mart. 1978

*Poetalia bulbosae* Rivas Goday et Rivas-Mart. in  
Rivas Goday et Ladero 1970

*Romuleion* Oberd. 1954 *nom. inval. (?)*

*Festuco valesiacaе-Poetum bulbosae ass. nov.*  
*hoc loco*

Class: ?

*Hedypnoido rhagadioloidis-Asphodeletum ramosi*  
*ass. nov. hoc loco*

## Acknowledgments

The authors thank Steve Latham (United Kingdom) for improving the English. We thank the Croatian Geological Survey for providing the Geological Map of the Republic of Croatia (sheet Cres and Lošinj). We also thank the Croatian Meteorological and Hydrological Service for providing climatological data from the meteorological station in the town of Krk, island of Krk, for the period 1998-2022. We would also like to thank two anonymous reviewers for their helpful suggestions.

The study was financed with funds from the Multi-purpose Institutional Financing Program supported by the Ministry of Science and Education of the Republic of Croatia.

## Author contribution statement

MT and NJ conceived the study, sampled the vegetation, made the statistical analysis, and share first authorship.

## References

Biondi, E., Pesaresi, S., Galdenzi, D., Gasparri, R., Biscotti, N., Del Viscio, G., Casavecchia, S., 2016: Post-abandonment dynamic on Mediterranean and sub-Mediterranean peren-

nial grasslands: the edge vegetation of the new class *Charyb-dido pancratii-Asphodeletea ramosi*. *Plant Sociology* 53(2), 3–18. <https://doi.org/10.7338/pls2016532/01>

- Biurrun, I., Willner, W., 2020: First report of the European Vegetation Classification Committee (EVCC). *Vegetation Classification and Survey* 1, 145–147. <https://doi.org/10.3897/VCS/2020/60352>
- Branković, Č., Güttler, I., Gajić-Čapka, M., 2013: Evaluating climate change at the Croatian Adriatic from observations and regional climate models' simulations. *Climate Dynamics* 41(9), 2353–2373. <https://doi.org/10.1007/s00382-012-1646-z>
- Brullo, S., Spampinato, G., 1991: La vegetazione dei corsi d'acqua della Sicilia. *Bollettino dell'Accademia Gioenia di Scienze Naturali* 23, 119–252.
- Čarni, A., Matevski, V., Šilc, U., Čušterevska, R., 2014: Early spring ephemeral therophytic non-nitrophilous grasslands as a habitat of various species of *Romulea* in the southern Balkans. *Acta Botanica Croatica* 73(1), 155–177. <https://doi.org/10.2478/botcro-2013-0017>
- De Cáceres, M., Legendre, P., 2009: Associations between species and groups of sites: indices and statistical inference. *Ecology* 90(12), 3566–3574. <https://doi.org/10.1890/08-1823.1>
- De Cáceres, M., Legendre, P., Moretti, M., 2010: Improving indicator species analysis by combining groups of sites. *Oikos* 119(10), 1674–1684. <https://doi.org/10.1111/j.1600-0706.2010.18334.x>
- Dengler, J., Löbel, S., Dolnik, C., 2009: Species constancy depends on plot size – a problem for vegetation classification and how it can be solved. *Journal of Vegetation Science* 20(4), 754–766. <https://doi.org/10.1111/j.1654-1103.2009.01073.x>
- Dufrêne, M., Legendre, P., 1997: Species assemblages and indicator species: the need for a flexible asymmetrical approach. *Ecological Monographs* 67(3), 345–366. [https://doi.org/10.1890/0012-9615\(1997\)067\[0345:SAAI\]2.0.CO;2](https://doi.org/10.1890/0012-9615(1997)067[0345:SAAI]2.0.CO;2)
- Dumančić, D., 1988: Unapređenje proizvodnosti travnjaka mediteranskog područja kao preduvjet razvoja ovčarstva i kozarstva [Improving the productivity of grasslands in the Mediterranean area as a prerequisite for the development of sheep and goat farming]. *Agronomski glasnik* 1, 105–111.
- EEA (European Environment Agency), 2019: The European environment - state and outlook 2020. Knowledge for transition to a sustainable Europe. Retrieved October 8, 2023 from <https://www.eea.europa.eu/soer/publications/soer-2020>
- Euro+Med. 2006-2023: Euro+Med PlantBase - the information resource for Euro-Mediterranean plant diversity. Retrieved September 1, 2023 from <http://www.europlusmed.org/>
- Foucault, B. (de), 2020: Sur la synsystème des *Rosmarinetea officinalis* ouest-méditerranéens. *Carnets Botaniques* 4, 1–27. <https://doi.org/10.34971/s9d8-qj43>
- Fuček, L., Matičec, D., Vlahović, I., Oštrić, N., Prtoljan, B., Kroljija, B., Korbar, T., Husinec, A., Palenik, D., 2015: Osnovna geološka karta Republike Hrvatske M 1:50 000: list Cres i Lošinj [Basic geological map of the Republic of Croatia, scale 1:50 000: sheet Cres and Lošinj]. Hrvatski geološki institut, Zagreb.
- Gajić-Čapka, M., Güttler, I., Cindrić, K., Branković, Č., 2018: Observed and simulated climate and climate change in the lower Neretva river basin. *Journal of Water and Climate Change* 9(1), 124–136. <https://doi.org/10.2166/wcc.2017.034>
- Hećimović, S., 1984: Vegetation der Inseln Bobara und Mrkan. *Acta Botanica Croatica* 43(1), 109–118.
- Horvat, I., Horvatić, S., 1934: *Chrysopogoneto-Satureion subspicatae*: ein neuer Verband der *Brometalia erecti* Braun-Blanquet. *Acta Botanica Instituti Botanici Universitatis Zagrebensis* 9(1), 8–12.

- Horvat, I., Glavaš, V., Ellenberg, H., 1974: Vegetation Südosteuropas. Geobotanica selecta 4. Gustav Fischer Verlag, Stuttgart.
- Horvatić, S., 1927: Flora i vegetacija otoka Plavnika [Flora and vegetation of the island of Plavnik]. Acta Botanica Instituti Botanici Universitatis Zagrebensis 2(1), 1–56.
- Horvatić, S., 1963: Vegetacijska karta otoka Paga s općim pregledom vegetacijskih jedinica Hrvatskog primorja [Vegetation map of the island of Pag with survey of the plant communities of Hrvatsko Primorje]. Prirodoslovna Istraživanja 33. Acta Biologica IV, 3–187.
- Horvatić, S., 1973: Syntaxonomic analysis of the vegetation of dry grassland and stony meadows in Eastern Adriatic coastal Karts district based on the latest phytocoenological research. Fragmenta Herbologica Jugoslavica 32, 1–15.
- IPCC (Intergovernmental Panel on Climate Change), 2014: Climate Change 2014: Synthesis Report. In: Core Writing Team, Pachauri, R. K., Meyer, L. A., (eds.), Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. IPCC, Geneva, Switzerland.
- McCune, B., Mefford, M. J., 2011: PC-ORD. Multivariate analysis of ecological data. Version 6.0 MjM Software, Gleneden Beach, Oregon, USA.
- Médail, F., 2017: The specific vulnerability of plant biodiversity and vegetation on Mediterranean islands in the face of global change. Regional Environmental Change 17(6), 1775–1790. <https://doi.org/10.1007/s10113-017-1123-7>
- Midolo, G., Herben, T., Axmanová, I., Marcenò, C., Pätsch, R., Bruelheide, H., Karger, D. N., Ačić, S., Bergamini, A., Bergmeier, E., Biurrun, I., Bonari, G., Čarni, A., Chiarucci, A., De Sanctis, M., Demina, O., Dengler, J., Dziuba, T., Fanelli, G., Garbolino, E., Giusso del Galdo, G., Goral, F., Güler, B., Hinojos-Mendoza, G., Jansen, F., Jiménez-Alfaro, B., Lengyel, A., Lenoir, J., Pérez-Haase, A., Pielech, R., Prokhorov, V., Rašomavičius, V., Ruprecht, E., Rüşina, S., Šilc, U., Škvorc, Ž., Stančić, Z., Tatarenko, I., Chytrý, M., 2023: Disturbance indicator values for European plants. Global Ecology and Biogeography 32(1), 24–34. <https://doi.org/10.1111/geb.13603>
- Mucina, L., Bültman, H., Dierssen, K., Theurillat, J.-P., Dengler, J., Čarni, A., Šumberová, K., Raus, T., Di Pietro, R., Gavilán García, R., Chytrý, M., Iakushenko, D., Schaminée, J. H. J., Bergmeier, E., Santos Guerra, A., Daniëls, F. J. A., Ermakov, N., Valachovič, M., Pigantti, S., Rodwell, J. S., Pallas, J., Capelo, J., Weber, H. E., Lysenko, T., Solomeshch, A., Dimopoulos, P., Aguiar, C., Freitag, H., Hennekens, S. M., Tichý, L., 2016: Vegetation of Europe: Hierarchical floristic classification system of plant, lichen, and algal communities. Applied Vegetation Science 19(S1), 3–264. <https://doi.org/10.1111/avsc.12257>
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A., Kent, J., 2000: Biodiversity hotspots for conservation priorities. Nature 403, 853–858. <https://doi.org/10.1038/35002501>
- Nikolić, T., Antonić, O., Alegro, A., Dobrović, I., Bogdanović, S., Liber, Z., Rešetnik, I., 2008: Plant species diversity of Adriatic islands: an introductory survey. Plant Biosystems 142(3), 435–445. <https://doi.org/10.1080/11263500802410769>
- Oberdorfer, E., 1954: Nordägyptische kraut- und zwergrsträucher im vergleich mit den entsprechenden Vegetationseinheiten des westlichen Mittelmeergebietes. Vegetatio 5(1), 88–96. <https://doi.org/10.1007/BF00299558>
- Official Gazette, 2019: Uredba o ekološkoj mreži i nadležnostima javnih ustanova za upravljanje područjima ekološke mreže [Regulation on the ecological network and the competences of public institutions for the management of ecological network areas]. 80. Croatian Government, Zagreb.
- Patarčić, M., Gajić-Čapka, M., Cindrić, K., Branković, Č., 2014: Recent and near-future changes in precipitation-extremes indices over the Croatian Adriatic coast. Climate Research 61(2), 157–176. <https://doi.org/10.3354/cr01250>
- Preislerová, Z., Jiménez-Alfaro, B., Mucina, L., Berg, C., Bonari, G., Kuzemko, A., Landucci, F., Marcenò, C., Monteiro-Henriques, T., Novák, P., Vynokurov, D., Bergmeier, E., Dengler, J., Apostolova, I., Bioret, F., Biurrun, I., Campos, J. A., Capelo, J., Čarni, A., Čoban, S., Csiky, J., Čuk, M., Čušterevska, R., Daniëls, F. J. A., De Sanctis, M., Didukh, Ya., Dítě, D., Fanelli, F., Golovanov, Y., Golub, V., Guarino, R., Hájek, M., Iakushenko, D., Indreica, A., Jansen, F., Jašková, A., Jiroušek, M., Kalníková, V., Kavğacı, A., Kucherov, I., Kuzmich, F., Lebedeva, M., Loidi, J., Lososová, Z., Lysenko, T., Milanović, Đ., Onyshchenko, V., Perrin, G., Peterka, T., Rašomavičius, V., Rodríguez-Rojo, M. P., Rodwell, J. S., Rüşina, S., Sánchez Mata, D., Schaminée, J. H. J., Semenishchenkov, Y., Shevchenko, N., Šibík, J., Škvorc, Ž., Smagin, V., Stešević, D., Stupar, V., Šumberová, K., Theurillat, J.-P., Tikhonova, E., Tzonev, R., Valachovič, M., Vassilev, K., Willner, W., Yamalov, S., Večeřa, M., Chytrý, M., 2022: Distribution maps of vegetation alliances in Europe. Applied Vegetation Science 25(1), e12642. <https://doi.org/10.1111/avsc.12642>
- R Core Team, 2023: R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.
- Rivas Goday, S., Rivas-Martínez, S., 1963: Estudio y clasificación de los pastizales españoles. Publicaciones del Ministerio de Agricultura, Madrid.
- Rivas-Martínez, S., Sáenz, S. R., Penas, A., 2011: Worldwide bioclimatic classification system. Global Geobotany 1, 1–634.
- Rodwell, J. S., Schaminée, J. H. J., Mucina, L., Pignatti, S., Dring, J., Moss, D., 2002: The diversity of European vegetation. an overview of phytosociological alliances and their relationships to EUNIS habitats. National reference centre for agriculture, nature and fisheries, Wageningen.
- Stanišić-Vujačić, M., Stešević, D., Hadžiablahović, S., Čaković, D., Šilc, U., 2022: An *Asphodelus ramosus* dominated plant community in Montenegro: fringe or grassland? Acta Botanica Croatica 81(1), 12–22. <https://doi.org/10.37427/botcro-2022-027>
- Stanišić-Vujačić, M., Stešević, D., Hadžiablahović, S., Šilc, U., 2023: Ecological and syntaxonomical characteristics of early spring therophytic ephemeral grasslands (alliance *Romuleion*) in the northeastern Mediterranean. Plant Biosystems 157(3), 540–563. <https://doi.org/10.1080/11263504.2023.2165570>
- Šegota, T., Filipčić, A., 2003: Köppenova podjela klime i hrvatsko nazivlje [Köppen climate division and Croatian nomenclature]. Geoadria 8(1), 17–37. <https://doi.org/10.15291/geoadria.93>
- Šegulja, N., 1970: Vegetacija sjeveroistočnog dijela Labinštine u Istri [Vegetation of the north-eastern part of Labin in Istria]. Acta Botanica Croatica 29, 157–172.
- Škvorc, Ž., Jasprica, N., Alegro, A., Kovačić, S., Franjić, J., Krstonošić, D., Vraneša, A., Čarni, A., 2017: Vegetation of Croatia: phytosociological classification of the high-rank syntaxa. Acta Botanica Croatica 76(2), 200–224. <https://doi.org/10.1515/botcro-2017-0014>
- Terzi, M., 2011: Nomenclatural revision for the order *Scorzonero-Chrysopogonetalia*. Folia Geobotanica 46(4), 411–444. <https://doi.org/10.1007/s12224-011-9100-2>

- Terzi, M., 2015: Numerical analysis of the order *Scorzoneretalia villosae*. *Phytocoenologia* 45(1–2), 11–32. <https://doi.org/10.1127/phyto/2015/0009>
- Terzi, M., 2023: A new *Asphodelus ramosus*-dominated association from the Murge Plateau (SE Italy). *Hacquetia* 22(2), 179–195. <https://doi.org/10.2478/hacq-2022-0020>
- Terzi, M., Jasprica, N., Pandža, M., Milović, M., Caković, D., 2023: Diversity and ecology of *Salvia officinalis* communities in the Western Balkans. *Plant Biosystems* 157(1), 175–187. <https://doi.org/10.1080/11263504.2022.2098868>
- Terzi, M., Fernández-González, F., Di Pietro, R., Theurillat, J.-P., 2024a: Phytosociological nomenclature of the class names *Helianthemetea guttati*, *Poetea bulbosae* and *Stipo giganteae-Agrostietea castellananae*. *Plant Biosystems* 158(1), 70–83. <https://doi.org/10.1080/11263504.2023.2287539>
- Terzi, M., Jasprica, N., Čarni, A., Matevski, V., Bergmeier, E., Theurillat, J.-P., 2024b: Nomenclature of the Balkan alliance *Romuleion graecae* (*Poetea bulbosae*). *Acta Botanica Croatica* 83(2), 87–91. <https://doi.org/10.37427/botcro-2024-020>
- Theurillat, J. P., Willner, W., Fernández-González, F., Bültmann, H., Čarni, A., Gigante, D., Mucina, L., Weber, H., 2021: International code of phytosociological nomenclature. 4th edition. *Applied Vegetation Science* 24(1), e1249. <https://doi.org/10.1111/avsc.12491>
- Tichý, L., Axmanová, I., Dengler, J., Guarino, R., Jansen, F., Mídolo, G., Nobis, M. P., Van Meerbeek, K., Ačić, S., Attorre, F., Bergmeier, E., Biurrun, I., Bonari, G., Bruelheide, H., Campos, J. A., Čarni, A., Chiarucci, A., Čuk, M., Čušterevska, R., Didukh, Y., Dítě, D., Dítě, Z., Dziuba, T., Fanelli, G., Fernández-Pascual, E., Garbolino, E., Gavilán, R.G., Gégout, J.-C., Graf, U., Güler, B., Hájek, M., Hennekens, S. M., Jandt, U., Jašková, A., Jiménez-Alfaro, B., Julve, P., Kambach, S., Karger, D. N., Karrer, G., Kavgaci, A., Knollová, I., Kuzemko, A., Kůzmič, F., Landucci, F., Lengyel, A., Lenoir, J., Marcenò, C., Moeslund, J.E., Novák, P., Pérez-Haase, A., Peterka, T., Pielech, R., Pignatti, A., Rašomavičius, V., Rüşüña, S., Saatkamp, A., Šilc, U., Škvorc, Ž., Theurillat, J.-P., Wohlgemuth, T., Chytrý, M., 2023: Ellenberg-type indicator values for European vascular plant species. *Journal of Vegetation Science* 34(1), e13168. <https://doi.org/10.1111/jvs.13168>
- Topić, J., Vuković, N., 2010: Krk i Plavnik [Krk and Plavnik]. In: Nikolić, T., Topić, J., Vuković N. (eds.), *Botanički važna područja Hrvatske*, 217–224. Prirodoslovno-matematički fakultet Sveučilišta u Zagrebu, Školska knjiga d.o.o., Zagreb.
- Trinajstić, I., 2001: Stjepan Horvatić i Gabrijel Tomažič - pioniri fitocenoloških istraživanja travnjaka u Sloveniji [Stjepan Horvatić and Gabrijel Tomažič - the pioneers of phytosociological investigations of grasslands in Slovenia]. *Hladnikia* 12–13, 23–29.
- Trinajstić, I., 2008: Biljne zajednice republike Hrvatske [Plant communities of Croatia]. Akademija Šumarskih Znanosti, Zagreb.
- van der Maarel, E., 1979: Transformation of cover-abundance values in phytosociology and its effects on community similarity. *Vegetatio* 39(2), 97–114. <https://doi.org/10.1007/BF00052021>
- Vargas, P., 2020: The Mediterranean floristic region: High diversity of plants and vegetation types. In: Goldstein, M. I., DellaSala, D. A. (eds.), *Encyclopedia of the world's biomes*, vol. 3, 602–616. Elsevier, Oxford. <https://doi.org/10.1016/B978-0-12-409548-9.12097-4>
- Vukadinović, V., 2021: Pedološka karta Hrvatske [Pedological map of Croatia]. Retrieved October 8, 2023 from [http://pedologija.com.hr/iBaza/DPKH\\_QGIS\\_VV.webp](http://pedologija.com.hr/iBaza/DPKH_QGIS_VV.webp). Josip Juraj Strossmayer University, Osijek.
- WBG (World Bank Group), 2021: Climate Risk Country Profile - Croatia. Retrieved October 8, 2023 from <https://reliefweb.int/report/croatia/climate-riskcountry-profile-croatia>
- Westhoff, V., van der Maarel, E., 1980: The Braun-Blanquet approach. In: Whittaker, R. H. (ed.), *Classification of plant communities*, 287–399. 2nd edition. Dr. W. Junk Publishers, The Hague, Boston, London. [https://doi.org/10.1007/978-94-009-9183-5\\_9](https://doi.org/10.1007/978-94-009-9183-5_9)

# Effects of hormopriming and pretreatment with gibberellic acid on fenugreek (*Trigonella foenum-graecum* L.) seed germination

Sabrina Gueridi, Lilya Boucelha, Ouzna Abrous-Belbachir, Réda Djebbar\*

University of Sciences and Technology Houari Boumediene (USTHB), Faculty of Biological Sciences, Laboratory of Biology and Physiology of Organisms, BP 32, El Alia, Bab Ezzouar, 16111 Algiers, Algeria.

**Abstract** – Various approaches are used to improve crop production. Seed priming is one of the simplest and least expensive methods currently used to ensure rapid and uniform yields. Our study highlights the role of priming and imbibition in improving seed germination. The objective of this study was to investigate the effect of seed imbibition and hormopriming with 0.1 mM gibberellic acid (GA<sub>3</sub>) on germination performance and biochemical changes in fenugreek (*Trigonella foenum-graecum* L.) radicles. The results showed that hydropriming and imbibition with GA<sub>3</sub> significantly improved germination performance and radicle growth. Concurrently, treatments induced stimulation of the antioxidant activities of superoxide dismutase, ascorbic peroxidase, catalase and guaiacol peroxidase, and decreased lipid peroxidation, stimulated an increase in total non-enzymatic antioxidant capacity and reduced glutathione content. Accumulation of hydrogen peroxide and cytochemical analysis of reactive oxygen species (ROS) *in situ* confirmed the role of imbibition in stimulating ROS. Interestingly the effects of imbibition with gibberellic acid were more effective than hormopriming, probably due to the partial degradation of GA<sub>3</sub> during dehydration process.

**Keywords:** hormopriming, germination, gibberellic acid, radicle, reactive oxygen species, *Trigonella foenum-graecum*

## Introduction

Plant production and productivity are strongly determined by seed germination, which is a critical step in the life cycle of higher plants (Cheng and Bradford 1999). Seed germination may be asynchronous and consequently plant performance may be far from satisfactory. There are currently several approaches to overcoming this problem, including priming. Seed priming is a pregermination technique that ensures rapid, uniform and synchronized germination and improves seedling vigour and growth under normal and adverse environmental conditions (Varier et al. 2010). The priming process involves the imbibition of seeds to allow pregermination metabolic activation, followed by dehydration prior to the reversible phase (avoiding radicle breakthrough) to avoid radicle emergence (Finch-Savage and Leubner-Metzger 2006). This technique is economical and environment-friendly. There are different priming methods for improving seed germination performance, such as hydropriming (imbibing seeds in water), os-

mopriming (imbibing in an osmotic solution), halopriming (imbibing in a saline solution), chemopriming (imbibing in chemical solutions), and hormopriming (imbibing with phytohormones). Hormopriming, the technique used in this study, is based on treatment with plant hormones such as auxins, abscisic acid, cytokinins, and gibberellins. This type of treatment results in a higher germination performance and the resulting plants can be more tolerant to abiotic stresses including water stress (Singh and Maheswari 2017). Several authors have shown that the positive effects of priming are associated with various physiological, biochemical, cellular, molecular and genetic changes such as mobilization of reserves, degradation of albumen, stimulation of osmolyte synthesis and activation of the cell cycle and some abiotic stress tolerance genes. The activation of antioxidant enzyme systems has been extensively studied in primed seeds (Varier et al. 2010, Boucelha et al. 2019a). The signalling roles of reactive oxygen species (ROS) in seed ger-

\* Corresponding author e-mail: reda\_djebbar@yahoo.fr

mination and dormancy have been well documented and it is assumed that ROS accumulation is beneficial for seed germination and seedling growth (see the review by Bailly et al. 2008 and Bailly, 2023). In a more precise way, Bailly et al. (2008) proposed the concept of the "oxidative window for germination", which restricts the occurrence of the onset of germination to a critical range of ROS level. Thus, ROS homeostasis regulates the beginning of germination, which is why activity of ROS-scavenging systems plays an important role. Furthermore, several authors also studied the involvement of ROS in the priming phenomenon (Boucelha et al. 2019a, Ren et al. 2023).

The role of ROS in seed dormancy control is due to their interaction with plant hormones that have central functions in seed dormancy and germination such as gibberellic acid (GA) (see review by Bailly et al. 2008). At low levels applied to the seed, GA stimulates germination by breaking dormancy through the activation of hydrolytic enzymes such as amylases, which promote the breakdown of reserves that provide energy for germination, resulting in rapid cell division and radicle elongation (Gubler et al. 1995). ROS have been reported to stimulate GA biosynthesis through a transcriptional regulation (Li et al. 2018) and GA treatment has been shown to induce ROS production (Cembrowska-Lech et al. 2015) by modifying the redox status of aleurone proteins, a process which might be related to ROS accumulation (Maya-Ampudia and Bernal-Lugo 2006).

Few studies have focused on the redox status of root radicles following treatment with GA, particularly applied as hormoprimer (Ellouzi et al. 2023). Thus, the objective of this work was to study the effect of GA applied in two different ways, by seed imbibition in GA followed by rinsing (pretreatment with GA) and by hormoprimer (GA treatment followed by rehydration) on the redox status of *Trigonella foenum-graecum* L. radicles by investigating enzymatic and non-enzymatic antioxidant systems. Fenugreek (*Trigonella foenum-graecum*) is an annual herb that belongs to the family Leguminosae cultivated worldwide, especially in southwest Asia, Middle East and Mediterranean. Due to its strong flavour and aroma fenugreek is one of those plants whose leaves and seeds are widely consumed as a spice in food preparations, and as an ingredient in traditional medicine. Its seeds are used for their carminative, tonic and aphrodisiac effects (Chopra et al. 1986) and it is assumed to have antidiabetic effect, hypocholesterolemic influence, antioxidant potency, digestive stimulant action, and hepatoprotective effect (Srinivasan 2006).

## Materials and methods

### Plant material

Our study was carried out on fenugreek seeds (*Trigonella foenum-graecum* L.), variety Halba from southern Algeria. Homogeneous seeds were selected from the same lot and rinsed with 5% hypochlorite solution for disinfection.

### Treatments, seed germination and measurement of seedlings radicle growth

For pretreatment seeds were treated by imbibition in distilled water or 0.1 mM GA<sub>3</sub> for 8 hours and then rinsed before germination. Hormoprimer and hydropriming were performed by seed soaking in 0.1 mM GA<sub>3</sub> or in distilled water, respectively, for 8 h and then rinsed and dehydrated under ventilation for 48 h before germination. Control seeds were not treated in any way before germination. Treated and control seeds were germinated in Petri dishes (9 cm in diameter) on four layers of absorbent paper soaked in distilled water. Three Petri dishes, with 50 seeds each, were used per treatment. For the first 24 h the germination experiment was performed in an oven at 25 °C, after which the Petri dishes were moved to a 14 h photoperiod of natural daylight and room temperature (24 ± 1 °C) for the next two days. After three days radicles were sampled for ROS detection, biochemical analyses and measurement of activity of antioxidant enzymes.

The length of radicle obtained from treated and untreated seeds was measured by graph paper each day for three days and expressed in cm. The fresh weight of treated and untreated radicles was measured after the 3<sup>rd</sup> day and expressed in grams.

### Cytochemical detection of ROS

Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) *in situ* was detected by the method of Thordal-Christensen et al. (1997), using a cytochemical method with 3,3'-diaminobenzidine (DAB). A brown colour is visible at the site of reaction of DAB with hydrogen peroxide, due to polymerisation of the DAB molecule.

Superoxide anions (O<sub>2</sub><sup>•-</sup>) *in situ* were detected by a cytochemical method using nitro blue tetrazolium (NBT) (Rao and Davis 1999). The superoxide radicals present in tissue reduce the NBT to a stable formazan of blue-indigo colour (Beyer and Fridovich 1987).

### Biochemical analyses

Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) was measured according to the method described by Alexieva et al. (2001) after reaction with 1 M potassium iodide (KI) in 100 mM potassium phosphate buffer (pH 7.0). The reaction was developed for 1 h in darkness (yellowish colour) and absorbance measured at 390 nm. The amount of hydrogen peroxide per fresh weight (FW) was calculated using a standard curve prepared with known concentrations of H<sub>2</sub>O<sub>2</sub> (0 to 10 nM) and expressed as nmol g<sup>-1</sup> FW.

Lipid peroxidation was measured by quantification of the coloured malondialdehyde complex (MDA-TBA) detected at a wavelength of 532 nm, as described by Popham and Novacky (1991). The MDA content was expressed as μmol g<sup>-1</sup> FW. This was calculated using the molar extinction coefficient of MDA (ε = 155 mM<sup>-1</sup> cm<sup>-1</sup>).

Reduced glutathione (GSH) was measured according to Moron et al. (1979) by using 5,5'- dithiobis-2-nitrobenzoic acid (DTNB) (Ellman's reaction) to give a yellow-coloured product that absorbs at 412 nm. The measurement was calculated from the molar extinction coefficient ( $\epsilon = 13.3 \text{ mM}^{-1} \text{ cm}^{-1}$ ) and expressed as  $\mu\text{mol g}^{-1} \text{ FW}$ .

The total non-enzymatic antioxidant capacity (TAC) was estimated by the phosphomolybdenum method (Prieto et al. 1999). TAC is expressed in mg equivalents of ascorbic acid per g of dry weight. A standard curve was constructed using  $0\text{-}300 \mu\text{g mL}^{-1}$  ascorbic acid.

### Activities of antioxidant enzymes

For extraction of catalase (CAT), superoxide dismutase (SOD) and ascorbate peroxidase (APX), 100 mg of radicles were cold ground in extraction buffer (0.1 M Tris-HCl, pH 8.1). The guaiacol peroxidase (GPOX) was extracted from 100 mg of radicles ground in 0.1 M potassium phosphate buffer ( $\text{KH}_2\text{PO}_4 / \text{K}_2\text{HPO}_4$ , pH 6.5) under cold conditions. The enzyme activity was measured in protein extracts by spectrophotometry. Total soluble proteins were determined by the Bradford method (Bradford 1976).

CAT activity was determined following the decomposition of  $\text{H}_2\text{O}_2$  at 240 nm using the method described by Anderson et al. (1995). The activity was expressed as  $\mu\text{mol}$  of  $\text{H}_2\text{O}_2$  degraded per minute per mg of protein. This activity was calculated using the molar extinction coefficient of  $\text{H}_2\text{O}_2$  ( $\epsilon = 36 \text{ mM}^{-1} \text{ cm}^{-1}$ ).

GPOX activity was determined according to the method of MacAdam et al. (1992), slightly modified by Boucelha et al. (2019b). The activity is measured by a colorimetric technique based on the increase in absorbance at 470 nm due to the polymerisation of guaiacol to tetraguaiacol (oxidation), which gives an orange colour in the presence of hydrogen peroxide. The activity was expressed in  $\mu\text{mol}$  of oxidised guaiacol per minute per mg of protein, using the molar extinction coefficient of tetraguaiacol ( $\epsilon = 26.6 \text{ mM}^{-1} \text{ cm}^{-1}$ ).

APX activity was measured according to the method of Nakano and Asada (1981) by following the oxidation of ascorbate by hydrogen peroxide, which absorbs at a wavelength of 290 nm. The enzymatic activity is expressed as  $\mu\text{mol}$  of ascorbate oxidised per minute per mg of protein. This activity was calculated using the extinction coefficient of ascorbate ( $\epsilon = 2.8 \text{ mM}^{-1} \text{ cm}^{-1}$ ).

SOD activity was measured by method reported by Marklund and Marklund (1974) and slightly modified by Boucelha et al. (2019a). It is based on the competition between the oxidation reaction of pyrogallol by superoxide ions and the dismutation by SOD. The increase in absorbance at 420 nm was due to the auto-oxidation of pyrogallol. An enzymatic unit was defined as the quantity of enzyme capable of inhibiting 50% of the autooxidation of pyrogallol under the conditions of the assay. The activity of SOD was expressed in units per minute and per mg of protein.

### Decomposition kinetic of $\text{GA}_3$

To study the stability of  $\text{GA}_3$  in water used for imbibition, we studied the spontaneous decomposition kinetic of 0.1 mM  $\text{GA}_3$  by spectrophotometry at 254 nm, measuring the appearance of gibberellic acid, a degradation product, according to the method of Pérez et al. (1996).

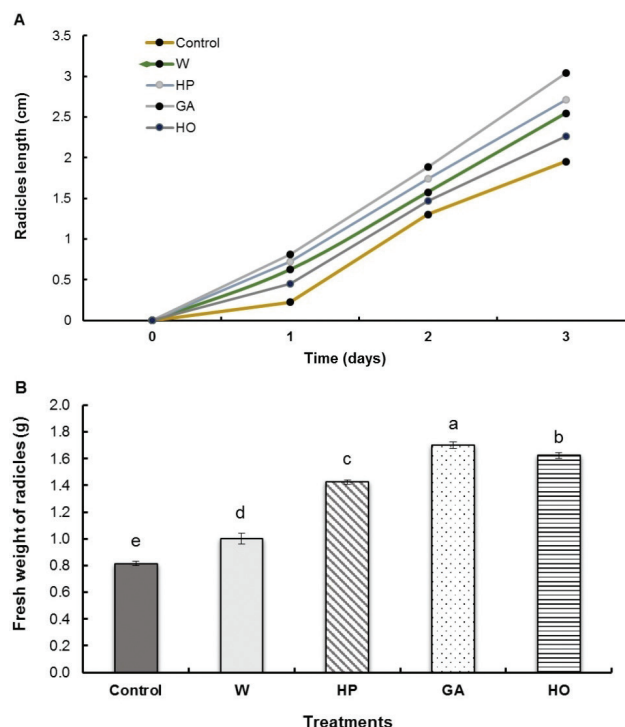
### Statistical analysis

The ANOVA test was used to compare the results. The statistical significance between the results was assessed according to the Tukey post-hoc test and values were considered statistically significant at  $P < 0.05$ . The analysis of data was performed through STATISTICA 6.0 software (Stat Soft, Inc.). All values were expressed as mean and standard deviation.

## Results

### Radicles growth

Fenugreek seed pregermination treatments significantly improved radicle length, following the same, almost linear, kinetics for all treatment types (Fig. 1A). However, this improvement was closely related to the type of treatment applied. In fact, we found that GA imbibition (without re-dehydration) showed the best growth, with an increase of about 66% on day 3 compared to the control batch. This was



**Fig. 1.** Radicle length (A) and fresh weight (B) of fenugreek seedlings after different treatments of seeds. W – imbibition with water, HP – hydropriming, GA – imbibition with 0.1 mM gibberellic acid, HO – hormoprimering with 0.1 mM gibberellic acid. Results are expressed as means ( $N = 3$ ) and error bars represent standard errors. Different alphabetical letters indicate a significant difference ( $P < 0.05$ ) between means.



**Fig. 2.** Detection of reactive oxygen species in fenugreek radicles after different treatments of seeds: hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) after 24 h of germination (A) and superoxide anion (O<sub>2</sub><sup>•-</sup>) after 48 h of germination (B). W – imbibition with water, HP – hydropriming, GA – imbibition with 0.1 mM gibberellic acid, HO – hormopriming with 0.1 mM gibberellic acid.

followed by the hydropriming (HP) with an increase of 49%, then the imbibition with water (W) and hormopriming (HO) with an increase of 36% and 20%, respectively, compared to the control. All treatments induced an increase in the fresh weight of fenugreek radicles (Fig. 1B). The greatest increases are recorded for imbibition with GA (107%) and hormopriming (95%). For the hydropriming, the increase was 75% while for imbibitions, it was only 23%.

**ROS production *in situ***

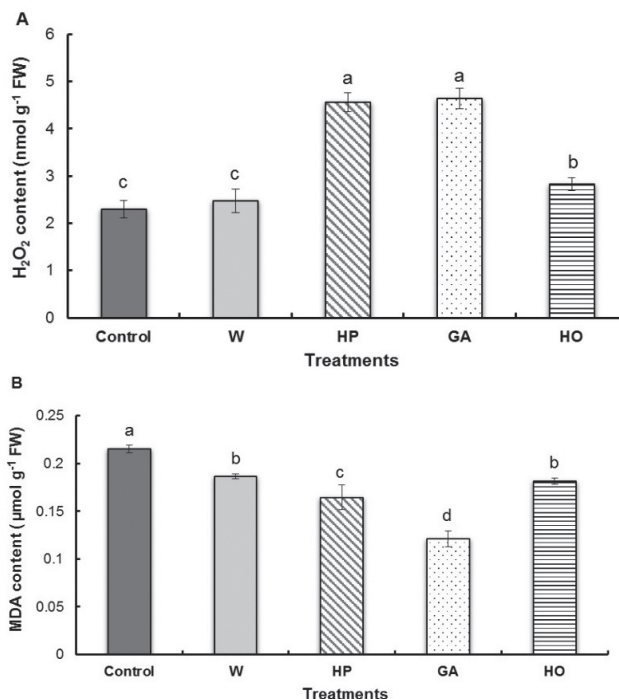
DAB assay revealed that the degree of H<sub>2</sub>O<sub>2</sub> accumulation in tissue varied according to the type of treatment applied. Control seed radicles were characterized by the lowest production of H<sub>2</sub>O<sub>2</sub> (Fig. 2A). A high accumulation of H<sub>2</sub>O<sub>2</sub> was observed in the radicles of GA and hydropriming treated seeds, especially in the region of the root cap and elongation. In contrast, a low H<sub>2</sub>O<sub>2</sub> accumulation was observed in the radicles resulting from hormone priming and water imbibition treatments.

Fenugreek radicles treated with NBT showed an accumulation of superoxide anions in the tissue (Fig. 2B). In the control radicles, the accumulation of superoxide anions was lower than in the radicles from treated seeds. Gibberellic acid imbibition showed the highest production of superoxide anions localized throughout the radicle. Hydropriming treatment also induced an accumulation of superoxide anions. However, in water imbibition and hormopriming treatments the presence of superoxide anions was only slightly visible.

**Hydrogen peroxide and malondialdehyde content**

Results indicate that the pregermination treatments of fenugreek seeds induced a significant increase in H<sub>2</sub>O<sub>2</sub> con-

centration in the radicles except for water imbibition (Fig. 3A). Indeed, hydropriming and GA imbibition resulted in almost the same increase in H<sub>2</sub>O<sub>2</sub> content (101% and 98%,



**Fig. 3.** Contents of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) (A) and malondialdehyde (MDA) (B) in the radicles of fenugreek seedlings after different treatments of seeds. W – imbibition with water, HP – hydropriming, GA – imbibition with 0.1 mM gibberellic acid. Results are expressed as means (N = 3) and error bars represent standard errors. Different alphabetical letters indicate a significant difference (P < 0.05) between means.



respectively), as the control. An increase of 23% was observed in the case of the hormopriming treatment compared to the control.

Pregermination treatments of fenugreek seeds showed a significantly higher MDA content of control radicles than the radicles of treated seeds (Fig. 3B). Seed imbibition with gibberellic acid, favoured a decrease in MDA content for 44% compared to the control. This was followed by the hydropriming treatment (decrease of 24%), then the hormopriming treatment (decrease of 16 %) and finally the water imbibition treatment (decrease of 14%) compared to the control.

### Activities of antioxidative enzymes

The pregermination treatment of fenugreek seeds induced a significant increase in antioxidant enzyme activities in some treatments (Fig. 4).

We observed significantly increased activity of catalase in seedlings from the gibberellic acid and hydropriming treated seeds, with increases of 88% and 33%, respectively, compared to the control. However, a less significant increase was observed in the water-imbibition (15%) and hormopriming treatment (12%) (Fig. 4A).

GPOX activity was significantly higher in fenugreek radicles pretreated with gibberellic acid, with an increase of 46% compared to the control. Hydropriming had a less prominent effect on the activation of this enzyme, with an increase of 16% over the control. In contrast, hormopriming and water-imbibition of the seeds had no effect on GPOX activity in the radicle (Fig. 4B).

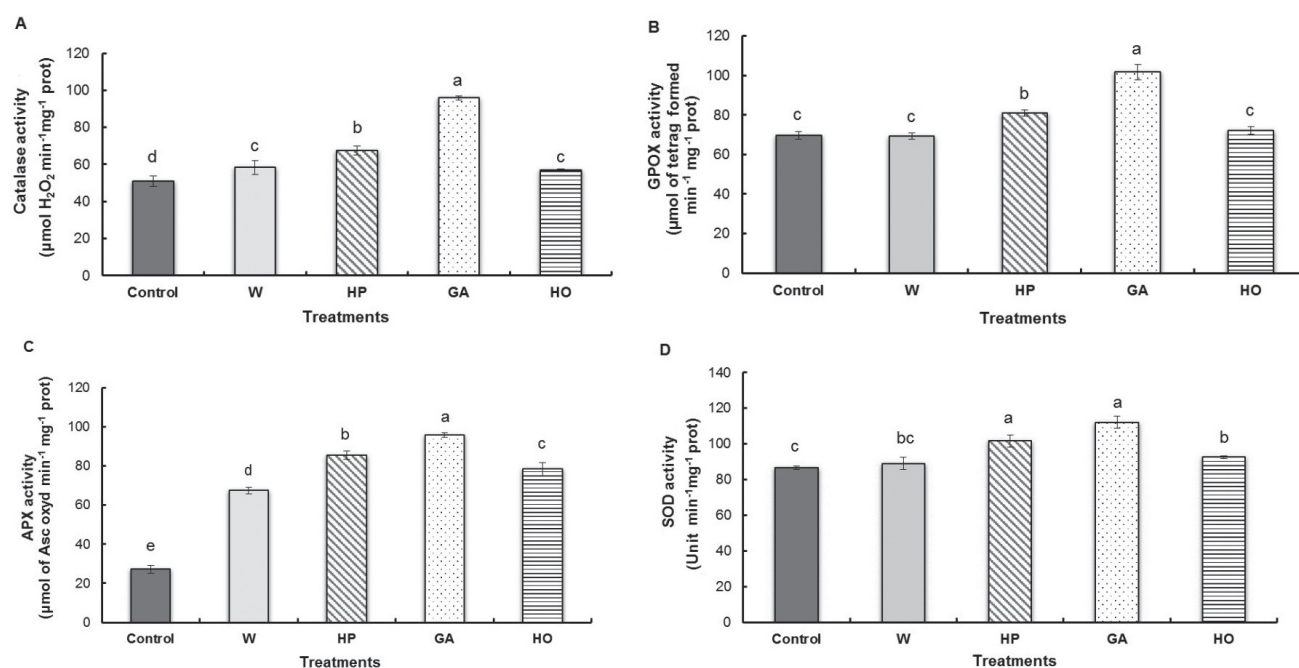
The results of the APX activity showed that the pretreatments caused a very significant activation of this enzyme. Indeed, GA imbibition and hydropriming caused the significantly highest increase compared to the control (252% and 215% respectively), while in hormopriming and water-imbibition, the increase was of 188% and 148% respectively (Fig. 4C).

The antioxidant enzyme activity of SOD in the radicle revealed that GA imbibition and hormopriming induced respectively significant increases of 29% and 17% over the control. A less significant increase was observed in the activity of this enzyme for hormopriming while water-imbibition treatments did not show any significant change from the control (Fig. 4D).

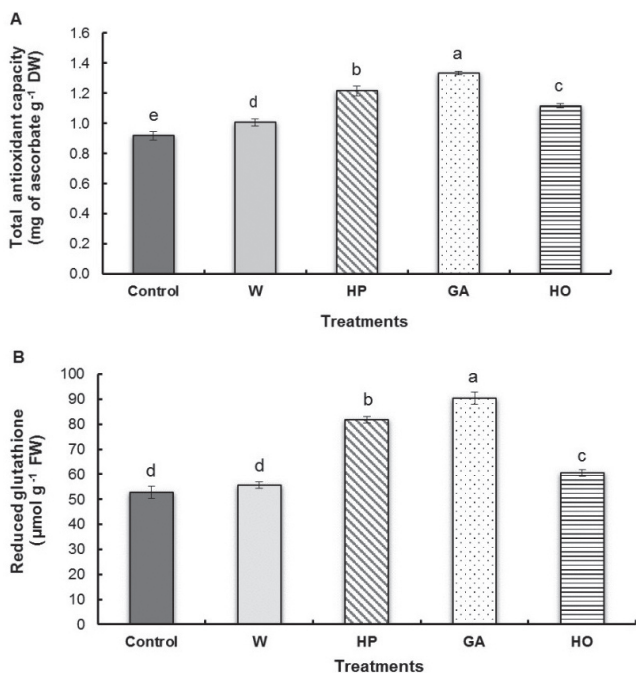
### Total antioxidant capacity and reduced glutathione content

All seed treatments significantly stimulated antioxidant capacity in the radicles of fenugreek seedlings; however, this increase was dependent on the type of treatment applied (Fig. 5A). Indeed, we observed that radicles soaked with gibberellic acid were characterized by the highest value in comparison to control (45%) followed by hydropriming (32%), hormopriming (22 %) and water-imbibition (10%) compared to the control.

Gibberellic acid and hydropriming treatments of fenugreek seeds caused a high increase in reduced glutathione content in the radicles (Fig. 5B) with percentages of 72% and 55% respectively compared to the control. However, we observed that hormopriming induced a weaker increase with 15% while water-imbibition treatment did not present any significant difference with the control.



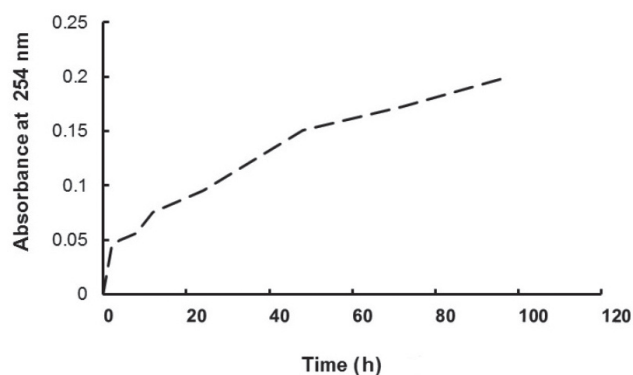
**Fig. 4.** Antioxidant activities of catalase (A), guaiacol peroxidase – GPOX (B), ascorbate peroxidase – APX (C) and superoxide dismutase – SOD (D) in fenugreek radicles after different treatments of seeds. W – imbibition with water, HP – hydropriming, GA – imbibition with 0.1 mM gibberellic acid, HO – hormopriming with 0.1 mM gibberellic acid. Results are expressed as means ( $N = 3$ ) and error bars represent standard errors. Different alphabetical letters indicate a significant difference ( $P < 0.05$ ) between means.



**Fig. 5.** Total antioxidant capacity (A) and reduced glutathione content (B) of fenugreek radicles after different treatments of seeds. W – imbibition with water, HP – hydropriming, GA – imbibition with 0.1 mM gibberellic acid, HO – hormopriming with 0.1 mM gibberellic acid. Results are expressed as means (N = 3) and error bars represent standard errors. Different alphabetical letters indicate a significant difference (P < 0.05) between means.

### Gibberellic degradation

In order to follow the stability of gibberellic acid in an aqueous medium, we followed its spontaneous degradation at 25 °C. From the curve obtained (Fig. 6), we were able to deduce that the gibberellic acid solution decomposes spontaneously into gibberellinic acid (inactive form).



**Fig. 6.** Kinetics of gibberellic acid degradation at 25 °C represented by gibberellinic acid appearance.

### Discussion

In this study, we found that pregermination treatments of fenugreek seeds, particularly imbibition with gibberellic acid, induced biochemical changes in the radicles derived

from these fenugreek seeds and consequently better seed germination and seedling growth.

A significant improvement in germination performance was observed in *Trigonella foenum-graecum* L. seeds that had undergone pregermination treatments, particularly in the gibberellic acid and hydropriming treatments, followed by the hormopriming and water imbibition.

Hydropriming is known to improve seed germination performance and has been reported for rice (Hussain et al. 2015), sunflower (Hussain et al. 2006), wheat (Ahmadi et al. 2007), maize (Janmohammadi et al. 2008), bean (Ghassemi-Golezani et al. 2010), lentil (Saglam et al. 2010) and black-eyed bean (Boucelha et al. 2019a). These authors have shown that priming was an effective method for improving germination performance, resulting in uniform and homogeneous cultures. Several authors have suggested that this improvement is related to changes at the seed level, such as accelerated water uptake (Gelormini 1995), increased respiration intensity Corbineau et al. (2000), nucleic acid synthesis, strong degradation of reserves (Varier et al. 2010), activation of antioxidant enzyme activities (Amooghaie and Vaviland 2011) and strong protein synthesis under genetic control (Varier et al. 2010). All these phenomena could be the consequence of a "memorization by the embryo" of events that occurred during the rehydration imposed during priming. It has been shown that a plant can store information when exposed to stress events and can use this memory to aid responses when these events reoccur (see review by Kinoshita and Seki, 2014). It has been demonstrated that epigenetic mechanisms are essential for stress memory and adaptation in plants (Chen and Arora 2013). Gibberellic acid is known to play a role in stimulating seed germination and is involved in many physiological and biochemical processes in plants (Mirheidari et al. 2022). Moreover, gibberellic acid is widely used in the laboratory and greenhouse to trigger the germination of some seeds that would otherwise remain dormant (Riley 1987). Thus, several works have reported the use of gibberellic acid as an exogenous treatment to enhance germination (Seandhalaksmi et al. 2022). Based on the work of Ogawa et al. (2003), exogenous GA enters the seed and is added to endogenous GA. Indeed, these authors showed that GA-deficient *Arabidopsis thaliana* seeds could not germinate without the addition of exogenous GA. This improvement has been linked to the synthesis and activation of amylases (Vieira et al. 2002) and lipases (Jridi et al. 2004), allowing accelerated germination with faster division cells (Li et al. 2018). Thus, seeds treated with gibberellic acid showed significant cell elongation compared to the other treatments. This cell expansion would also be due to the GA activation of aquaporins, membrane channels involved in water transport, which increases cell turgidity (Ogawa et al. 2003).

The treatments applied to fenugreek seeds stimulated the production of ROS in embryonic tissues, more precisely in the meristematic zone. This ROS formation, as evidenced by cytochemical tests and increased H<sub>2</sub>O<sub>2</sub> content, was more

pronounced in the gibberellic acid and hydropriming treatments. These effects are in line with the concept of the "oxidative window" proposed by Bailly et al. (2008) and confirmed in the case of hydropriming by Boucelha et al. (2019a). According to Bailly et al. (2008), for seed germination it is necessary that ROS content be within a range that allows signalling, while lower or higher amounts of ROS would lead to inability to germinate. Published data have shown that ROS are key players in several physiological processes in seeds such as seed dormancy control due to their interaction with plant hormones like gibberellic acid (Finkelstein et al. 2008) and the perception and transduction of environmental conditions during imbibition (Bailly 2019). In dry seeds, i.e. at low moisture content, ROS accumulation would probably result mainly from non-enzymatic reactions. In this physiological state, glucose and amino groups derived from amino acid and nucleic acids are condensed to form Amadori and Maillard products, which are major sources for ROS production (Sun and Leopold 1995) and lipid peroxidation (McDonald 1999). However, during imbibition, the reactivation of metabolism causes an enhanced accumulation of ROS, generally resulting from electron leakage within the mitochondrial electron transport chain (Kranter et al. 2010).

Bailly et al. (2008) proposed a mechanism for the control of dormancy and germination through a dialogue between ROS and hormone. Indeed, according to these authors, there is an interaction between ROS and the gibberellic acid signalling pathway during germination. Under imbibition conditions, NADPH oxidase and  $\beta$ -oxidation increase ROS levels, which in turn repress the DELLA protein responsible for the negative regulation of GA synthesis. This induces the synthesis of gibberellic acid, which then triggers the activation of GA-inducible transcription factors GAMYB which in turn induces the transcription of  $\alpha$ -amylase in the aleurone layers of many seeds (Gubler et al. 1995, Kaneko et al. 2002).

Membrane lipid peroxidation, which causes damage to cell membranes, is a good indicator of the presence of reactive forms of oxygen and thus allows assessment of cellular oxidative stress intensity. Oracz et al. (2007) supported the hypothesis of an inverse correlation between MDA content and seed dormancy since they observed that the MDA content increased during dormancy alleviation. Our results showed a more pronounced decrease in MDA content in the gibberellic acid and hydropriming treatments compared to the control as well as hormoprimering and water imbibition where the decrease was less prominent. A reduction in lipid peroxide content in hydroprimed seeds has already been reported by El-Araby and Hegazi (2004) for tomato and Sharma et al. (2014) for okra seeds. This decrease was also observed in gibberellic acid-treated seeds, as shown in the work of Li et al. (2013) and Ahmad (2010). The reduction in MDA accumulation in primed seeds could be explained by improved membrane repair during the priming process and induction of antioxidant enzymes (Nawaz et al. 2013). For

all treatments in this study, the decrease of MDA content correlated with the increase of the different antioxidative enzymes activities as well as increased level of glutathione, but did not correlate with the level of ROS. Therefore, the MDA content could be the result of the enhanced antioxidative defence which prevents oxidative damage despite higher level of ROS. Thus, cells maintain ROS homeostasis during germination (Li et al. 2013).

Fenugreek seed treatment stimulated the activation of antioxidant enzymes such as superoxide dismutase, catalase, ascorbate peroxidase and guaiacol peroxidase at the radicle level, with the most significant effect in the gibberellic acid pretreatment. These results are in agreement with those of different authors who showed that treatment of seeds with gibberellic acid leads to an increase in enzymatic antioxidant activity (Li et al. 2013). The increase in antioxidant enzymes activities was also observed in fenugreek radicles after hydropriming treatment although less prominently. Several studies showing that hydroprimed seeds of several crop species are characterised by very high antioxidant activities (Varier et al. 2010, Boucelha et al. 2019a, Melzi Ou Mezzi et al. 2021). Thus, several works have linked germination improvement to increased antioxidant enzymatic activities, which allow the elimination of free radicals, and, then, the restoration of the homeostasis of the redox status (Varier et al. 2010, Boucelha et al. 2019a, Boucelha et al. 2021).

Total antioxidant capacity corresponds to the presence of natural antioxidants capable of preventing oxidative damage (Priando et al. 1999). These non-enzymatic antioxidants include glutathione, ascorbic acid (vitamin C), tocopherols (vitamin E), carotenoids and phenolic compounds including flavonoids (Asada 2006). TAC levels reflect the reduced state of these molecules. Our results showed that radicles from seeds soaked in gibberellic acid had the highest total non-enzymatic antioxidant activity, which is in agreement with lower MDA content. Few studies have measured non-enzymatic antioxidant activities after pregermination treatments. Boucelha et al. (2019a) showed no significant changes in hydroprimed *Vigna unguiculata* seeds, while the results of Melzi Ou Mezzi et al. (2021) suggested that hydropriming stimulates total antioxidant activities in fenugreek radicles.

Imbibition in gibberellic solution (without rehydration) and hydromoprimering induced separately increases in almost all parameters (radicles growth,  $H_2O_2$  content, catalase, GPOX, APX and SOD activities, TAC, GSH content). However, hormoprimering in which seeds were dehydrated after treatment with gibberellic acid resulted in values more like those obtained for seeds imbibed in water. This could mean that some of the exogenous gibberellic acid that entered the seed during imbibition may be degraded to gibberellic acid during drying. Pérez et al. (1996) reported that gibberellic acid loses its biological activity in aqueous alkaline solutions over time by degradation to gibberellic acid via an isomeric form *iso*-GA (degradation intermediate),

both of which are inactive and do not induce amylase activity in barley endosperm. On the basis of these observations and our results, we suggest that drying the seeds for 48 h caused a partial degradation of GA to inactive forms, which was unable to exert a beneficial effect on the seeds.

## Conclusion

Pretreatment of seeds with gibberellic acid and hydropriming resulted in improved germination performance and balanced redox status of fenugreek (*Trigonella foenum-graecum* L.) seeds. These physiological and biochemical changes at the radicle level would be the result of the activation of certain cell signalling pathways leading to a change in gene expression that needs to be further elucidated.

## References

- Ahmad, P., 2010: Growth and antioxidant responses in mustard (*Brassica juncea* L.) plants subjected to combined effect of gibberellic acid and salinity. *Archives of Agronomy and Soil Science* 56(5), 575–588. <https://doi.org/10.1080/03650340903164231>
- Ahmadi, A., Mardeh, A. S., Poustini K., Jahromi M. E., 2007: Influence of osmo and hydropriming on seed germination and seedling growth in wheat (*Triticum aestivum* L.) cultivars under different moisture and temperature conditions. *Pakistan Journal of Biological Sciences* 10(22), 4043–4049. <https://doi.org/10.3923/pjbs.2007.4043.4049>
- Alexieva, V., Sergiev, I., Mapelli, S., Karanov, E., 2001: The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant, Cell and Environment* 24(12), 1337–1344. <https://doi.org/10.1046/j.1365-3040.2001.00778.x>
- Amooghaie R., and Vaviland M., 2011: The combined effect of gibberellic acid and long time osmo-priming on seed germination and subsequent seedling growth of *Klussia odoratissima* Mozaff. *African Journal of Biotechnology* 10(66), 14873–14880. <https://doi.org/10.5897/AJB11.1984>
- Anderson, M. D., Prasad, T. K., Stewart, C. R., 1995: Changes in isozyme profiles of catalase, peroxidase, and glutathione reductase during acclimation to chilling in mesocotyls of maize seedlings. *Plant Physiology* 109(4), 1247–1257. <https://doi.org/10.1104/pp.109.4.1247>
- Asada, K., 2006: Production and scavenging of reactive oxygen species in chloroplasts and their functions. *Plant Physiology* 141(2), 391–396. <https://doi.org/10.1104/pp.106.082040>
- Bailly, C., El-Maarouf-Bouteau, H., Corbineau, F., 2008: From intracellular signaling networks to cell death: the dual role of reactive oxygen species in seed physiology. *Comptes Rendus Biologies* 331(10), 806–814. <https://doi.org/10.1016/j.crv.2008.07.022>
- Bailly, C., 2023: ROS, seed germination. In: Mittler, R., Van Breusegem, F. (eds.), *Oxidative stress response in plants*, 177–204. Academic Press, London. <https://doi.org/10.1016/bs.abr.2022.10.003>
- Beyer, W. F. Jr, Fridovich, I., 1987: Assaying for superoxide dismutase activity: some large consequences of minor changes in conditions. *Analytical Biochemistry* 161(2), 559–566. [https://doi.org/10.1016/0003-2697\(87\)90489-1](https://doi.org/10.1016/0003-2697(87)90489-1)
- Boucelha, L., Abrous-Belbachir, O., Djebbar, R., 2021: Is protein carbonylation a biomarker of seed priming and ageing. *Functional Plant Biology* 48(6), 611–623. <https://doi.org/10.1071/FP21001>
- Boucelha, L., Djebbar, R., Abrous-Belbachir, O., 2019a: *Vigna unguiculata* L. seed priming is related to redox status of plumule, radicle and cotyledons. *Functional Plant Biology* 46(6), 584–594. <https://doi.org/10.1071/FP18202>
- Boucelha, L., Djebbar, R., Abrous-Belbachir O., 2019b: The involvement of guaiacol peroxidase in embryo development of *Vigna unguiculata* (L.) Walp. during germination. *Analele Universității din Oradea, Fascicula Biologie* 26(2), 110–116.
- Bradford, M.M., 1976: A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72, 248–254. <https://doi.org/10.1006/abio.1976.9999>
- Cembrowska-Lech D., Koprowski M., Kępczyński J., 2015: Germination induction of dormant *Avena fatua* caryopses by KAR1 and GA3 involving the control of reactive oxygen species (H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub><sup>•-</sup>) and enzymatic antioxidants (superoxide dismutase and catalase) both in the embryo and the aleurone layers. *Journal of Plant Physiology* 176, 169–179. <https://doi.org/10.1016/j.jplph.2014.11.010>
- Chen, K., Arora, R., 2013: Priming memory invokes seed stress-tolerance. *Environmental and Experimental Botany* 94, 33–45. <https://doi.org/10.1016/j.envexpbot.2012.03.005>
- Cheng, Z., Bradford, K. J., 1999: Hydrothermal time analysis of tomato seed germination responses to priming treatments. *Journal of Experimental Botany* 50(33), 89–99. <https://doi.org/10.1093/jxb/50.330.89>
- Chopra, R. N., Nayar, S. L., Chopra, I. C., 1986: *Glossary of indian medicinal plants (including the Supplement)*. Publisher Council of Scientific and Industrial Research, New Delhi, 51–83.
- Corbineau, F., Özbingöl, N., Vinel, D., Come, D., 2000: Improvement of tomato seed germination by osmo-priming as related to energy metabolism. In: Black, M., Bradford, K. J., Vasquez, Ramos, J. (eds.), *Seed biology*, 467–476. *Advances and applications: proceedings of the sixth international workshop on seeds*. Merida, Mexico CABI, Cambridge. 2000. <https://doi.org/10.1079/9780851994048.0467>
- El-Araby, M. M., Hegazi, A. Z., 2004: Responses of tomato seeds to hydro- and osmo-priming, and possible relations of some antioxidant enzymes and endogenous polyamine fractions. *Egyptian Journal of Biology* 6, 81–93. <https://doi.org/10.1104/pp.122.2.327>
- Ellouzi, H., Zorrig, W., Amraoui, S., Oueslati, S., Abdelly, C., Rabhi, M., Siddique, K. H. M., Hessini, K., 2023: Seed priming with salicylic acid alleviates salt stress toxicity in barley by suppressing ROS accumulation and improving antioxidant defense systems, compared to halo- and gibberellin priming. *Antioxidants* 12, 1779. <https://doi.org/10.3390/antiox12091779>
- Finch-Savage, W. E., Leubner-Metzger, G., 2006: Seed dormancy and the control of germination. *New Phytologist* 171(3), 501–523. <https://doi.org/10.1111/j.14698137.2006.01787.x>
- Finkelstein, R., Reeves, W., Ariizumi, T., Steber, C., 2008: Molecular aspects of seed dormancy. *Annual Review of Plant Biology* 59, 387–415. <https://doi.org/10.1146/annurev.arplant.59.032607.092740>
- Gelormini, G., 1995: *Optimisation des propriétés germinatives des graines de colza par initialisation: aspects méthodologiques et fondamentaux*. Thèse nouveau doctorat, Université de Lille, France 171 p.
- Ghassemi-Golezani, K., Chadordooz-Jeddi, A., Nasrullahzadeh, S., Moghaddam, M., 2010: Influence of hydro-priming duration on field performance of pinto bean (*Phaseolus vulgaris*

- L.) cultivars. *African Journal of Agricultural Research* 5(9), 893–897. <https://doi.org/10.5897/AJAR.9000422>
- Gubler, F., Kalla, R., Roberts, J. K., Jacobsen, J. V., 1995: Gibberellin-regulated expression of a *myb* gene in barley aleurone cells: evidence for Myb transactivation of a high-pI  $\alpha$ -amylase gene promoter. *The Plant Cell* 7(11), 1879–1891. <https://doi.org/10.1105/tpc.7.11.1879>
- Hussain, M., Farooq, M., Basra, S. M., Ahmad, N., 2006: Influence of seed priming techniques on the seedling establishment, yield and quality of hybrid sunflower. *International Journal of Agriculture and Biology* 8(1), 14–18.
- Hussain, S., Zheng, M., Khan, F., Khaliq, A., Fahad, S., Peng, S., Nie, L., 2015: Benefits of rice seed priming are offset permanently by prolonged storage and the storage conditions. *Scientific Reports* 5(1), 1–12. <https://doi.org/10.1038/srep08101>
- Janmohammadi, M., Dezfuli, P. M., Sharifzadeh, F., 2008: Seed invigoration techniques to improve germination and early growth of inbred line of maize under salinity and drought stress. *General and Applied Plant Physiology* 34, 215–226.
- Jridi, T., Belguith, H., Hammami, M., Hamida, J. B., 2004: Effect of gibberellic acid on lipid reserve mobilization during sunflower (*Helianthus annuus* L.) seed germination. *Rivista Italiana delle Sostanze Grasse* 81(4), 239–243.
- Kaneko, M., Itoh, H., Ueguchi-Tanaka, M., Ashikari, M., Matsuo, M., 2002: The  $\alpha$ -amylase induction in endosperm during rice seed germination is caused by gibberellin synthesized in epithelium. *Plant Physiology*, 128, 1264–1270. <https://doi.org/10.1104/pp.010785>
- Kinoshita, T., Seki, M., 2014: Epigenetic memory for stress response and adaptation in plants. *Plant and Cell Physiology* 55(11), 1859–1863. <https://doi.org/10.1093/pcp/pcu125>
- Kranner, I., Roach, T., Beckett, R.P., Whitaker, C., Minibayeva, F., 2010: Extracellular production of reactive oxygen species during seed germination and early seedling growth in *Pisum sativum*. *Journal of Plant Physiology* 167, 805–811. <https://doi.org/10.1016/j.jplph.2010.01.019>
- Li, X., Jiang, H., Liu, F., Cai, J., Dai, T., Cao, W., Jiang, D., 2013: Induction of chilling tolerance in wheat during germination by pre-soaking seed with nitric oxide and gibberellin. *Plant Growth Regulation* 71, 31–40. <https://doi.org/10.1007/s10725-013-9805-8>
- Li, Z., Gao, Y., Zhang, Y., Lin, C., Gong, D., Guan, Y., Hu, J., 2018: Reactive oxygen species and gibberellin acid mutual induction to regulate tobacco seed germination. *Frontiers in Plant Science* 9, 1279. <https://doi.org/10.3389/fpls.2018.01279>
- MacAdam, J. W., Nelson, C. J., Sharpe, R. E., 1992: Peroxidase activity in the leaf elongation zone of Tall Fescue. *Plant Physiology* 99, 872–878. <https://doi.org/10.1104/pp.99.3.872>
- Marklund, S., Marklund, G., 1974: Involvement of the superoxide anion radical in the autoxidation of pyrogallol and a convenient assay for superoxide dismutase. *European Journal of Biochemistry* 47(3), 469–74. <https://doi.org/10.1111/j.1432-1033.1974.tb03714.x>
- Maya-Ampudia, V., Bernal-Lugo, I., 2006: Redox-sensitive target detection in gibberellic acid-induced barley aleurone layer. *Free Radical Biology and Medicine* 40, 1362–1368. <https://doi.org/10.1016/j.freeradbiomed.2005.12.004>
- McDonald, M. B., 1999. Seed deterioration: Physiology, repair and assessment. *Seed Science and Technology* 27, 177–237.
- Melzi Ou Mezzi, C., Boucelha, L., Abrous-Belbachir, O., Djebbar, R., 2021: Effects of hydropriming and chemical pretreatments of *Trigonella foenum-graecum* (L.) seeds on germination, antioxidant activities and growth. *Analele Universității din Oradea, Fascicula Biologie* 28(2), 165–175.
- Mirheidari, F., Hatami, M., Ghorbanpour, M., 2022: Effect of different concentrations of IAA, GA3 and chitosan nano-fiber on physio-morphological characteristics and metabolite contents in roselle (*Hibiscus sabdariffa* L.). *South African Journal of Botany* 145, 323–333. <https://doi.org/10.1016/j.sajb.2021.07.021>
- Moron, M. S., Depierre, J. W., Mannervik, B., 1979: Levels of glutathione, glutathione reductase and glutathione S-transferase activities in rat lung and liver. *Biochimica et Biophysica Acta (BBA)-general subjects* 582(1), 67–78. [https://doi.org/10.1016/0304-4165\(79\)90289-7](https://doi.org/10.1016/0304-4165(79)90289-7)
- Nakano, Y., Asada, K., 1981: Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. *Plant and Cell Physiology* 22, 867–880. <https://doi.org/10.1093/oxfordjournals.pcp.a076232>
- Nawaz, J., Hussain, M., Jabbar, A., Nadeem, G. A., Sajid, M., Subtain, M., Shabbir, I., 2013: Seed priming a technique. *International Journal of Agriculture and Crop Sciences* 6(20), 1373–1381.
- Ogawa, M., Hanada, A., Yamauchi, Y., Kuwahara, A., Kamiya, Y., Yamaguchi, S., 2003: Gibberellin biosynthesis and response during Arabidopsis seed germination. *The Plant Cell* 15(7), 1591–1604. <https://doi.org/10.1105/tpc.011650>
- Oracz, K., El-Maarouf Bouteau, H., Farrant, J. M., Cooper, K., Belghazi, M., Job, C., Job, D., Corbineau, F., Bailly, C., 2007: ROS production and protein oxidation as a novel mechanism for seed dormancy alleviation. *Plant Journal* 50, 452–465. <https://doi.org/10.1111/j.1365-313X.2007.03063.x>
- Pérez, F. J., Vecchiola, A., Pinto, M., Agosin, E., 1996: Gibberellic acid decomposition and its loss of biological activity in aqueous solutions. *Photochemistry* 41(3), 675–679. [https://doi.org/10.1016/0031-9422\(95\)00689-3](https://doi.org/10.1016/0031-9422(95)00689-3)
- Popham, P. L., Novacky, A., 1991: Use of dimethyl sulfoxide to detect hydroxyl radical during bacteria-induced hypersensitive reaction. *Plant Physiology* 96(4), 1157–1160. <https://doi.org/10.1104/pp.96.4.1157>
- Prieto, P., Pineda, M., Aguilar, M., 1999: Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. *Analytical Biochemistry* 269, 337–341. <https://doi.org/10.1006/abio.1999.4019>
- Rao, M.V., Davis, K.R., 1999: Ozone-induced cell death occurs via two distinct mechanisms in Arabidopsis: the role of salicylic acid. *Plant Journal* 17, 603–614. <https://doi.org/10.1046/j.1365-313x.1999.00400.x>
- Ren, M., Tan, B., Xu, J., Yang, Z., Zheng, H., Tang, Q., Zhang, X., Wang, W., 2023: Priming methods affected deterioration speed of primed rice seeds by regulating reactive oxygen species accumulation, seed respiration and starch degradation. *Frontiers in Plant Science* 14, 1267103. <https://doi.org/10.3389/fpls.2023.1267103>
- Riley, M., 1987: Gibberellic acid for fruit set and seed germination. *CRFG Journal* 19, 10–12.
- Saglam, S., Sibel, D. A. Y., Gamze, K. A. Y. A., Gürbüz, A., 2010: Hydropriming increases germination of lentil (*Lens culinaris* Medik.) under water stress. *Notulae Scientia Biologicae* 2(2), 103–106. <https://doi.org/10.15835/nsb224602>
- Sharma, A. D., Rathore, S. V. S., Srinivasan, K., Tyagi, R. K., 2014: Comparison of various seed priming methods for seed germination, seedling vigour and fruit yield in okra (*Abelmoschus esculentus* L. Moench). *Scientia Horticulturae* 165, 75–81. <https://doi.org/10.1016/j.scienta.2013.10.044>
- Srinivasan, K., 2006: Fenugreek (*Trigonella foenum-graecum*): A review of health beneficial physiological effects. *Food Re-*

- views International 2, 203–224. <https://doi.org/10.1080/87559120600586315>
- Sun, W.Q., Leopold, A.C., 1995: The Maillard reaction and oxidative stress during aging of soybean seeds. *Physiologia Plantarum* 94, 94–104. <https://doi.org/10.1111/j.1399-3054.1995.tb00789.x>
- Thordal-Christensen, H., Zhang, Z., Wei, Y., Collinge, D.B., 1997: Subcellular localization of H<sub>2</sub>O<sub>2</sub> in plants, H<sub>2</sub>O<sub>2</sub> accumulation in papillae and hypersensitive response during barley- powdery mildew interaction. *Plant Journal* 11, 1187–1194. <https://doi.org/10.1046/j.1365-313X.1997.11061187.x>
- Variar, A., Vari, A. K., Dadlani, M., 2010: The subcellular basis of seed priming. *Current Science* 99, 450–456
- Vieira, A. R., Gracas, M. D., Vieira, G. C., Fraga, A. C., Oliveira, J. A., Santos C. D. D., 2002: Action of gibberellic acid (GA<sub>3</sub>) on dormancy and activity of  $\alpha$ -amylase in rice seed. *Revista Brasileira de Sementes* 24 (2): 43–48. <https://doi.org/10.1590/s0101-31222002000200008>

# Effects of acetic acid treatment on growth and pigment contents in barley

Aslihan Temel<sup>1\*</sup>, Taylan Kosesakal<sup>2</sup>

<sup>1</sup>Istanbul University, Faculty of Science, Department of Molecular Biology and Genetics, 34134 Vezneciler, Istanbul, Türkiye

<sup>2</sup>Istanbul University, Faculty of Science, Department of Biology, 34134 Vezneciler, Istanbul, Türkiye

**Abstract** – Acetic acid (AA) is an organic acid and has been widely used as food preservative and a dietary spice in vinegar form. In addition to its therapeutic uses in its vinegar form, AA attenuates inhibitory effects of stress in plants. However, in some plant species toxic effects of AA have been found. Therefore, in this study, 0, 2.5 and 5 mM concentrations of AA were applied to 2-day-old barley (*Hordeum vulgare* L. cv. Bornova-92) seedlings for 2 days in order to investigate the possible toxic effects of AA. After treatments, seedlings were grown in AA-free conditions for 2 days to recover. AA inhibited root and shoot growth; decreased water content, fresh weight, chlorophyll, pheophytin, and carotenoid contents. However, anthocyanin and flavonoid contents, as well as the levels of UV-absorbing compounds and UV-B marker increased in the leaves of AA-treated plants. AA increased hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) content in shoots and induced cell death in roots. Soluble carbohydrate content decreased in roots of AA-treated plants while insoluble carbohydrate content increased. Our results demonstrate that AA in young barley seedlings can exhibit its toxic effects through oxidative stress, which induced antioxidative response in the form of molecules with antioxidative activities. These effects persisted for 2 days after the removal of AA.

**Keywords:** acetic acid, anthocyanins, cell death, hydrogen peroxide, UV-absorbing compounds

## Introduction

Soil organic acids are a water-soluble fraction of organic molecules in the rhizosphere. Organic acids (OAs) in soil may be produced by plants and microorganisms or are the result of organic matter decomposition (Adeleke et al. 2017). The concentration of monocarboxylic OAs in soil can reach up to 1 mM. Acetic acid (AA), a monocarboxylic OA, is the smallest OA and is less effective in mobilizing minerals than high molecular weight OAs. Half-life of AA can be a few days (Adeleke et al. 2017). AA, also known as ethanoic acid or vinegar, absorbs moisture; is corrosive to metals, and is widely used in industry and household cleaning. Once AA enters the cell, it dissociates into an acetate anion and a proton. Acetate is converted by acetyl-CoA synthetase (ACSS) to produce acetyl-coenzyme A (acetyl-CoA), an essential molecule produced in carbohydrate metabolism and histone acetylation (Pietrocola et al. 2015). Lynch (1977) reported that 0.1-1 mM AA treatment stimulates root growth in barley seedlings. Recent papers have suggested that AA can protect plants against biotic (Chen et al. 2019) and abiotic stress

(Utsumi et al. 2019) through jasmonic acid (JA) and abscisic acid (ABA) synthesis. AA treatment induces *de novo* JA synthesis and enhanced survival during drought stress is linked to histone acetylation (Kim et al. 2017). AA-treated plants show increased leaf water content and pigment levels, as well as upregulation of stress-response and stress-tolerance genes (Chen et al. 2019, Utsumi et al. 2019). AA supplementation ameliorates the toxic effects of seawater in mung bean by increasing uptake of Ca<sup>2+</sup> and Mg<sup>2+</sup> and decreasing uptake of Na<sup>+</sup>, enhancing antioxidant activity, water use efficiency and the contents of several metabolites *e.g.* soluble sugars, phenolics and flavonoids (Rahman et al. 2019). AA also enhances drought tolerance in maize and *Arabidopsis* through proline metabolism (Mahmud et al. 2023) and in soybean through improved antioxidant defence and photosynthesis and accumulation of soluble sugars and free amino acids (Rahman et al. 2021). However, besides these positive effects there are some reports about negative effects of AA on plants. In cassava plants, application of

\* Corresponding author e-mail: atemel@istanbul.edu.tr

20–50 mM AA induced wilting (Utsumi et al. 2019) while in maize seedlings root and shoot growth was inhibited by AA at doses as low as 10 mM (Allen and Allen 2020). In reed (*Phragmites australis*) root growth was reduced at 0.3 mM, and entirely inhibited at 1.7 mM acetic acid (Armstrong et al. 1996). In *Chlamydomonas reinhardtii* AA can even cause cell death (Zuo et al. 2012).

In this study, the effects of AA treatment on early seedling growth were investigated in barley. For this purpose, root and shoot length, fresh (FW) and dry (DW) weight and water content (WC) of seedlings, as well as pigment and H<sub>2</sub>O<sub>2</sub> content in leaves, and soluble and insoluble carbohydrate contents in roots were measured. Possible cytotoxic effects of AA treatment were investigated by measuring cell viability in roots by Evans Blue staining, which can penetrate into dead cells and therefore discriminates between viable and non-viable cells. Results of our study show that AA treatment inhibited growth and caused decreases in the levels of photosynthetic pigments e.g. chlorophyll, carotenoid, and pheophytin, while increasing the contents of anthocyanin, flavonoids, UV-absorbing compounds and UV-B marker. AA treatment caused the accumulation of H<sub>2</sub>O<sub>2</sub> in leaves and decreased cell viability in roots. Insoluble carbohydrate content was enhanced by AA treatment, while soluble carbohydrate content decreased.

## Materials and methods

### Plant material, growth conditions and treatment

Barley (*H. vulgare* cv. Bornova-92) mature seeds were obtained from the Aegean Agricultural Research Institute (AARI, Izmir, Türkiye).

Ten seeds were placed between two filter papers in a 9-cm-diameter Petri dish containing 6 mL water for overnight imbibition at 6 °C and then germinated in the dark (16/8 h, 25/18 °C, 70 ± 5% humidity, darkness) for 3 days. Uniformly-germinated seedlings were subsequently placed on a filter paper containing 6 mL of the test solution containing AA for 2 days under a light intensity of 1400 μmol m<sup>-2</sup> s<sup>-1</sup> (16/8 h, 25/18 °C, 70 ± 5% humidity). At the end of the treatment, seedlings were rinsed, transferred to dishes containing 6 mL of water, and then further incubated for 48 h for recovery at the same growth conditions. Test solutions contained 0, 2.5 mM (0.0143% v/v) or 5 mM (0.0286% v/v) AA (glacial, Merck). Plant samples were harvested immediately after AA treatment (day 0) or 2 days after AA was removed (day 2). Day 2 group represents recovery plants.

### Measurement of seedling growth and water content

Seedlings were briefly soaked on filter paper and weighed to determine FW. To determine DW, seedlings were dried at 65 °C until the weight became constant. FW and DW were expressed as milligrams (mg). Water content was calculated according to the formula  $WC = ((FW - DW) / FW) \times 100$  and expressed as a percentage. Root and shoot lengths were expressed as centimetres (cm).

### Determination of pigment content

To estimate the content of chlorophyll, carotenoid and pheophytin, leaves were homogenized in 80% acetone, incubated at –20 °C and, centrifuged at 6000 × g for 5 minutes at 4 °C. Supernatants were used to determine chlorophyll *a* and *b*, carotenoid and pheophytin levels. The absorbances of the extracts at 470, 655, 663 and 666 nm were measured in a glass cuvette (104.002-OS, Hellma) using Nanodrop (2000C, Thermo Fisher) due to the large volume of the samples and expressed as mg g<sup>-1</sup> FW (Lichtenthaler 1987, Costa et al. 2006).

To estimate the content of UV-B-absorbing compounds, leaves were homogenized in a methanol : HCl solution (99:1) and centrifuged at 10000 × g for 15 min at 4 °C. To determine the levels of UV-B-absorbing compounds and flavonoids, the absorbances (A) of the supernatants were measured at 300 and 350 nm in a quartz cuvette (104.002-QS, Hellma) using Nanodrop. To determine the levels of anthocyanins, the absorbances of the supernatants were measured at 535 and 657 nm. Anthocyanin contents were calculated according to the formula  $((A_{535} - (0.25 \times A_{657}))$  using a molar extinction coefficient of 38000 L mol<sup>-1</sup> cm<sup>-1</sup>. Flavonoid contents were calculated using a molar extinction coefficient of 20000 L mol<sup>-1</sup> cm<sup>-1</sup>. Anthocyanin and flavonoid contents were expressed as μmol g<sup>-1</sup> FW and μmol mg<sup>-1</sup> FW, respectively. Content of other UV-B absorbing compounds measured at A<sub>300</sub> was expressed as A<sub>300</sub> g<sup>-1</sup> FW (Cicek et al. 2012).

### Determination of hydrogen peroxide and UV-B marker content

Leaves were homogenized in 0.1% trichloroacetic acid and centrifuged at 10000 × g for 15 min and the supernatants were collected. The absorbances of the supernatants at 440 nm were measured to determine UV-B marker content which was expressed as A g<sup>-1</sup> FW (Cicek et al. 2012).

For the determination of H<sub>2</sub>O<sub>2</sub>, supernatants were mixed with 0.1 M Tris-HCl and 1 M potassium iodide and incubated at room temperature (RT) for 90 min. The absorbances were read at 390 nm and the H<sub>2</sub>O<sub>2</sub> amounts of the unknown samples were estimated according to the standard curve (0–330 nmol) of H<sub>2</sub>O<sub>2</sub> (Merck) and expressed as nmol g<sup>-1</sup> FW (Cicek et al. 2012).

### Evans Blue staining

Cell viability in roots was measured according to Baker and Mock (1994). Root samples were immersed in 0.25% Evans Blue stain for 20 min at RT. Then, roots were rinsed with water for 30 min, and 10 root tips (1 cm long) were incubated in 1% SDS : 50% methanol at 50 °C for 1 h. The absorbances of the methanol : SDS solution containing stain released from cells were measured at 595 nm.

### Determination of carbohydrate content

Carbohydrate extractions were performed according to Sonjaroon et al. (2018). Briefly, roots were homogenized in 80% ethanol, incubated at 75 °C for 15 min and centrifuged



at  $6000 \times g$  for 5 min. After the collection of the supernatant, the extraction was repeated twice. The supernatants were combined and used to measure soluble sugar content. The pellet phase containing ethanol-insoluble material was used for starch analysis.

Starch extraction was performed according to McCready et al. (1950) and Sonjaroon et al. (2018). Briefly, the pellet was dried at RT to evaporate EtOH completely and then dissolved in 52% perchloric acid at  $6^\circ\text{C}$  for 30 min. After centrifugation at  $2000 \times g$  for 5 minutes and recovery of the supernatant, extraction was repeated twice. The combined supernatants were used to estimate starch content. Soluble sugar and starch (insoluble carbohydrate) contents were measured according to an optimized phenol-sulphuric acid method (Masuko et al. 2005). Fifty  $\mu\text{L}$  of the sample was mixed with 150  $\mu\text{L}$  of  $\text{H}_2\text{SO}_4$  and 30  $\mu\text{L}$  of 5% phenol solution (a generous gift from Chembio, Türkiye) incubated at  $90^\circ\text{C}$  for 5 min and then cooled to RT. The absorbances were read at 490 nm. The glucose amount of the unknown samples was estimated according to the standard curve (0–100  $\mu\text{g}$ ) of glucose. For the estimation of starch, the concentration value was multiplied by 0.9. Soluble sugar and starch contents were expressed as  $\mu\text{g mg}^{-1}$  FW.

### Statistical analysis

All experiments were conducted as independent triplicates. Each datapoint is the arithmetic mean of biological triplicates ( $N = 3$ ) and the technical triplicates were also included in each experiment. Data were analysed by 2-Way ANOVA, and Tukey's multiple comparison test using Graphpad Prism (version 8.0.1.244). The ANOVA included 2 independent variables as time (day 0 and day 2) and AA concentration (0, 2.5 and 5 mM).

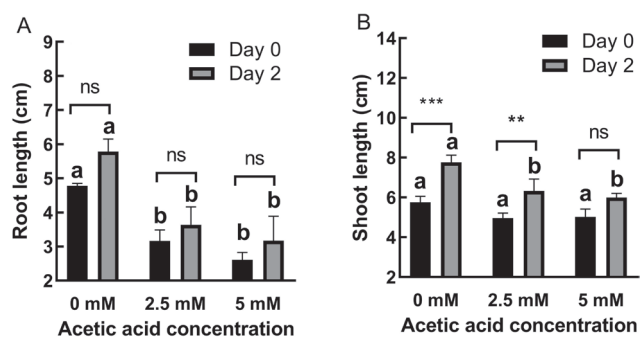
## Results

### Seedling growth and water content

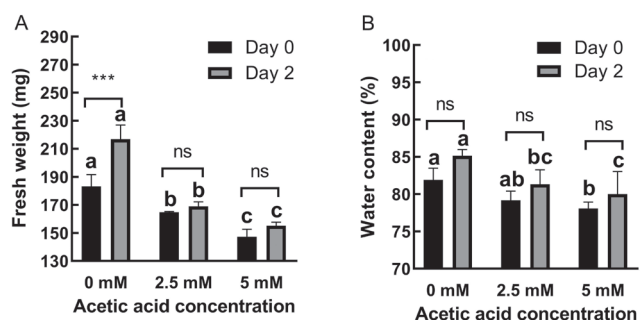
AA treatment significantly ( $P < 0.01$ ) decreased root (Fig. 1A) and shoot (Fig. 1B) lengths. At day 0, immediately after AA treatment with 2.5 and 5 mM concentration for 2 days 33.79% and 45.42% decreases in root lengths, respectively, were observed. At day 2, in seedlings recovering for 2 days under non-AA conditions, decreases were 37.07% and 45.15%, at 2.5 and 5 mM concentrations, respectively. For shoot lengths, AA treatment caused 13.73% and 12.63% decreases at day 0 and 18.47% and 22.70% decreases at day 2 for 2.5 and 5 mM concentrations, respectively.

AA-treated plants could not restore shoot and root growth after AA was removed. The effects of AA on root and shoot length were time-independent ( $P > 0.05$ ).

AA treatment significantly decreased ( $P < 0.01$ ) FW (Fig. 2A) and WC (Fig. 2B) without affecting ( $P > 0.05$ ) DW (Online Suppl. Fig. 1A). AA-treated plants could not restore FW after AA was removed. At day 0, AA treatment caused 10.02% and 19.58% decreases in FW at 2.5 and 5 mM concentrations, respectively. At day 2, these decreases were



**Fig. 1.** Changes in root (A) and shoot (B) lengths in 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors ( $N = 3$ ). Columns indicated by different letters are statistically different at the same timepoint ( $P < 0.05$ ). Data representing the same concentration at different timepoints are indicated as not significant (ns,  $P > 0.05$ ), \*\* for  $P < 0.01$  and \*\*\* for  $P < 0.001$ .



**Fig. 2.** Changes in fresh weight (A) and water content (B) in 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors ( $N = 3$ ). Columns indicated by different letters are statistically different at the same timepoint ( $P < 0.05$ ). Data representing the same concentration at different timepoints are indicated as not significant (ns,  $P > 0.05$ ) and \*\*\* for  $P < 0.001$ .

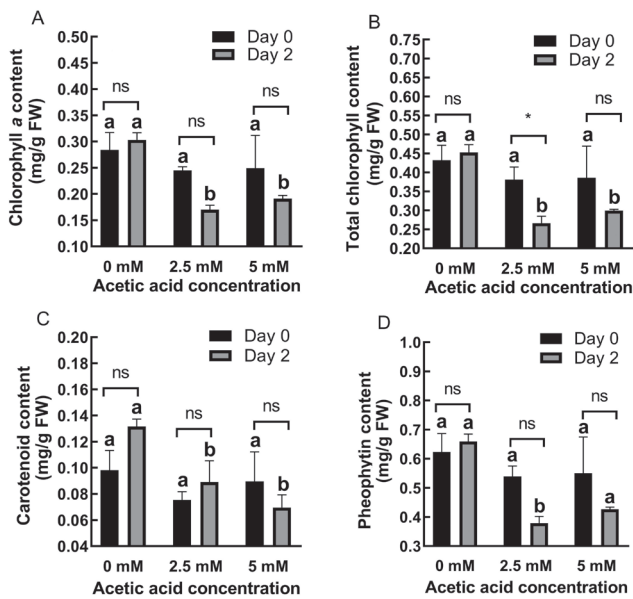
22.08% and 28.43%, respectively. AA treatment also caused 3.32% and 4.68% decreases in WC at day 0 and 4.48% and 6.04% decreases at day 2 for 2.5 and 5 mM concentrations, respectively.

The effects of AA on FW were time-dependent ( $P < 0.01$ ), while its effects on WC were not time dependent ( $P > 0.05$ ).

### Chlorophyll, carotenoid and pheophytin content

AA treatment decreased chlorophyll *a* (Fig. 3A), total chlorophyll (Fig. 3B), carotenoid (Fig. 3C), and pheophytin (Fig. 3D) levels but significantly ( $P < 0.01$ ) at day 2, in the recovery group of plants.

At day 0, AA treatment caused 13.77% and 12.29% decreases in chlorophyll *a* content at 2.5 and 5 mM concentrations, respectively. At day 2, these decreases were 43.82% and 36.89%, respectively. For total chlorophyll content, AA treatment caused 11.74% and 10.68% decreases at day 0 and 41.22% and 33.84% decreases at day 2 for 2.5 and 5 mM concentrations, respectively. At day 0, AA treatment caused 23.33% and 8.92% decreases in carotenoid content at 2.5 and



**Fig. 3.** Changes in chlorophyll *a* (A), total chlorophyll (B), carotenoid (C) and pheophytin (D) content in shoots of 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors (N = 3). Columns indicated by different letters are statistically different at the same timepoint (P < 0.05). Data representing the same concentration at different timepoints are indicated as not significant (ns, P > 0.05) and \* for P < 0.05.

5 mM concentrations, respectively. At day 2, these decreases were 32.29% and 47.18%, respectively. For pheophytin content, AA treatment caused 13.54% and 11.70% decreases at day 0 and 42.51% and 35.21% decreases at day 2 for 2.5 and 5 mM concentrations, respectively. Chlorophyll *b* content (On-line Suppl. Fig. 1B) was not significantly affected (P > 0.05).

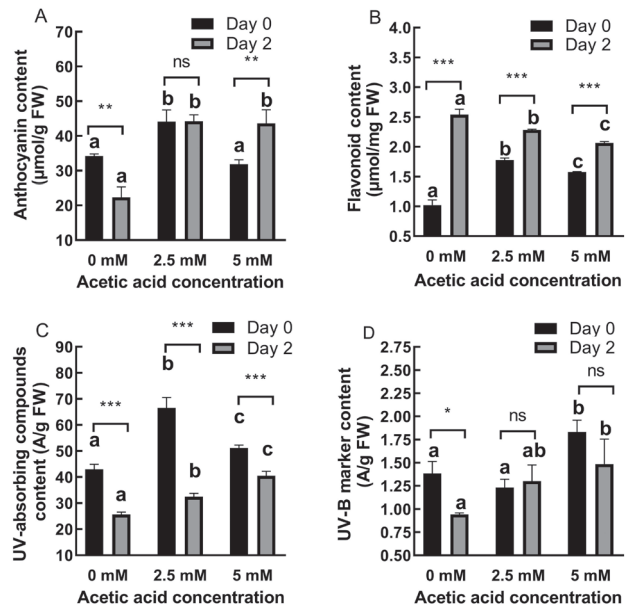
The effects of AA on photosynthetic pigments were found to be time-dependent (P < 0.05).

**Content of anthocyanins, flavonoids, UV-absorbing compounds and UV-B marker**

AA treatment mostly increased (P < 0.01) content of anthocyanins (Fig. 4A), flavonoids (Fig. 4B), UV-absorbing compounds (Fig. 4C) and UV-B marker (Fig. 4D) in shoots. At day 0, AA treatment caused a 28.98% increase and a 6.89% decrease in anthocyanin content at 2.5 and 5 mM concentrations, respectively. At day 2, both concentrations caused increases (97.94% and 95.37%, respectively). For flavonoid content, AA treatment caused 73.89% and 54.61% increases at day 0 while at day 2, 10.02% and 18.65% decreases for 2.5 and 5 mM concentrations, respectively, were noticed.

For UV-absorbing compounds, AA treatment caused 54.80% and 19% increases at day 0 and 26.93% and 58.04% increases at day 2 for 2.5 and 5 mM concentrations, respectively.

At day 0, AA treatment caused an 11.06% decrease and a 32.28% increase in UV-B marker content at 2.5 and 5 mM



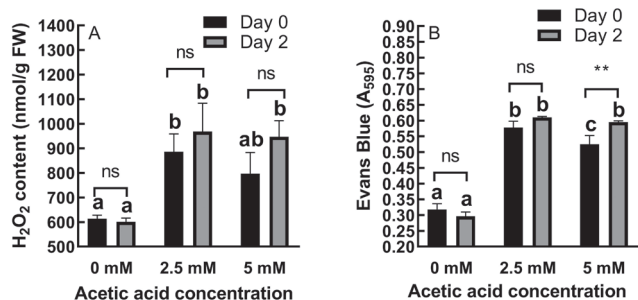
**Fig. 4.** Changes in anthocyanin (A), flavonoid (B), UV-absorbing compounds (C) and UV-B marker (D) content in shoots of 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors (N = 3). Columns indicated by different letters are statistically different at the same timepoint (P < 0.05). Data representing the same concentration at different timepoints are indicated as not significant (ns, P > 0.05), \* for P < 0.05, \*\* for P < 0.01 and \*\*\* for P < 0.001.

concentrations, respectively. At day 2, both concentrations caused increases (38.14% and 57.66%, respectively).

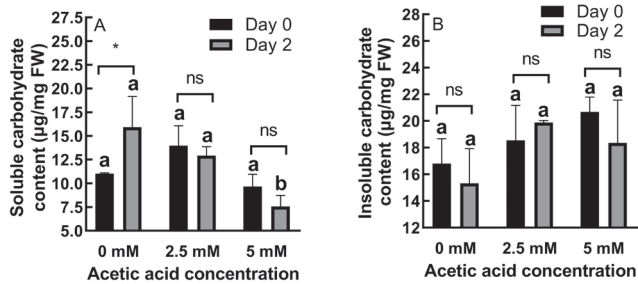
AA-treated plants accumulated all pigments except flavonoids even after AA was removed. The effects of AA on UV-protective pigments were time-dependent (P < 0.05).

**H<sub>2</sub>O<sub>2</sub> content and cell viability**

AA treatment dramatically increased (P < 0.01) H<sub>2</sub>O<sub>2</sub> content in shoots (Fig. 5A) while in roots, increase in A<sub>600</sub> (Fig. 5B), indicated reduced cell viability, and these effects persisted even after AA was removed. With respect to H<sub>2</sub>O<sub>2</sub> content, AA treatment caused 44.22% and 29.71% increases



**Fig. 5.** Changes in H<sub>2</sub>O<sub>2</sub> content in barley shoots (A) and cell viability measured by Evans Blue stain (B) in roots of 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors (N = 3). Columns indicated by different letters are statistically different (P < 0.05). Data representing the same concentration at different timepoints are indicated as not significant (ns, P > 0.05) and \*\* for P < 0.01.



**Fig. 6.** Changes in soluble (A) and insoluble carbohydrate (B) contents in roots of 2-day old barley seedlings under acetic acid (AA) treatment. Results are expressed as means  $\pm$  standard errors ( $N = 3$ ). Columns indicated by different letters are statistically different ( $P < 0.05$ ). Data representing the same concentration at different timepoints are indicated as not significant (ns,  $P > 0.05$ ) and \* for  $P < 0.05$ .

at day 0 and 61.08% and 57.61% increases at day 2 for 2.5 and 5 mM concentrations, respectively.

AA treatment caused 81.76% and 65.09% increases in  $A_{600}$  values at day 0 and 105.96% and 100.89% increases at day 2 for 2.5 and 5 mM concentrations, respectively.

The effects of AA on cell viability were time-dependent ( $P < 0.01$ ), while its effects on  $H_2O_2$  content were not time-dependent ( $P > 0.05$ ).

### Carbohydrate content

AA treatment had significant ( $P < 0.05$ ) effects on carbohydrate content in roots. At day 0, AA treatment caused a 26.66% increase and a 12.43% decrease in soluble carbohydrate content at 2.5 and 5 mM concentrations, respectively. At day 2, both concentrations caused decreases (18.83% and 52.54%, respectively) (Fig. 6A).

With respect to insoluble carbohydrate content, AA treatment induced 10.42% and 23.15% increases at day 0 and 29.78% and 19.76% increases at day 2 for 2.5 and 5 mM concentrations, respectively (Fig. 6B).

The effects of AA on soluble carbohydrate content were time-dependent ( $P < 0.05$ ) while its effects on insoluble carbohydrate content were not time dependent ( $P > 0.05$ ).

### Discussion

Stress conditions inhibit plant growth in terms of root and shoot length, DW and leaf area (Rahman et al. 2019). Under stressful conditions plants synthesize various phyto-protectant metabolites that help them defend against stress (Parvin et al. 2022). These metabolites can also be applied exogenously e.g. salicylic acid (SA) and phenolic compounds and alleviate stress-induced decreases in yield and productivity by affecting several processes including plant growth, photosynthesis, membrane permeability, antioxidant systems and synthesis of various molecules (Parvin et al. 2022). OAs were also reported to accumulate in plants under stress as these molecules, besides being intermediates in carbon

metabolism, are important metabolically active solutes for osmotic adjustment, the balance of cation excess or in coping with nutrient deficiencies and metal tolerance (López-Bucio et al. 2000). AA, an OA, was shown to confer tolerance in various plant species, including tomato, cassava, Arabidopsis, rapeseed, maize, rice and wheat against several stressors (Kim et al. 2017, Chen et al. 2019, Rahman et al. 2019, Utsumi et al. 2019, Rahman et al. 2021, Mahmud et al. 2023). AA-sprayed mung bean plants exhibited enhanced shoot and primary root length and shoot DW under salt conditions (Rahman et al. 2019). The same group later showed that foliar application of AA resulted in improvement of root biomass and leaf area in soybean exposed to drought (Rahman et al. 2021). Utsumi et al. (2019) reported the maintenance of leaf relative WC in drought-treated cassava plants. AA resulted in higher WC and elevated water use efficiency due to decreased stomatal conductance in drought stressed *Cunninghamia lanceolata* plants (Li et al. 2022). A recent paper by Mahmud et al. (2023) similarly stated that AA restored shoot and root growth in maize under drought stress. Interestingly, in our study AA treatment of barley seedlings inhibited plant growth, especially root growth and decreased FW and WC but did not affect DW. These detrimental effects mostly could not be repaired after the 2 day-recovery period. Thus, it can be inferred that, when applied directly to the young seedlings through roots, AA might be regarded as a stressor and the roots, being the first exposed to the effects of AA, are more affected than the shoots. Our results are different than those of most papers, which reveal the positive effects of AA (Kim et al. 2017, Li et al. 2022, Mahmud et al. 2023). However, these studies were performed on different species, with higher concentrations of AA and for longer periods and demonstrated its effects in plants growing in a culture or soil. In the present study, a shorter treatment duration (2 days) with lower concentrations (2.5 and 5 mM) of AA was tested on very young barley seedlings growing on moist filter papers. Lynch (1977) reported that the growth promoting effects of AA depend on pH and plant species. In an interesting study, Allen and Allen (2020) suggested that adjusting pH of the medium can compensate the harmful effects of AA in maize roots. They demonstrated that under unbuffered conditions, such as during germination on filter paper, acetic acid exists in the membrane permeable undissociated form which caused maize seedling root inhibition. Similarly, in our study the pH of the water with AA was not adjusted, which could explain the toxic effects of AA on barley seedlings, even at low concentrations.

Severe stress conditions cause degradation of chlorophyll, while low stress stimulates chlorophyll content, to allow plants to cope with stress (Agathokleous et al. 2020). Under stressful conditions, degradation of free chlorophyll is necessary to prevent cell damage (Takamiya et al. 2000) and chlorophyll content can reflect the damage caused by stress (Agathokleous et al. 2020). Exogenously applied AA reduced the impact of drought stress in cassava plants by increasing chlorophyll and carotenoid amounts (Utsumi et

al. 2019). AA also improved chlorophyll fluorescence in *C. lanceolata* under drought stress by maintaining higher chlorophyll contents due to delay of degradation of pigments or induction of their synthesis (Li et al. 2022). Hawrylak-Nowak et al. (2015) stated that AA had slight effects on carotenoid content in the first leaf of sunflower plant. In the present study, AA decreased chlorophyll and carotenoid contents as well as pheophytin content. A decrease in chlorophyll and carotenoid could be correlated with decreased growth and WC, but we expected accumulation of pheophytin in leaves of AA-treated seedlings because of the acidic environment in which the seedlings had grown. Pheophytin, a breakdown product of chlorophyll is observed mainly during leaf senescence (Lin and Charng 2021) or under oxidative stress (Szafrńska et al. 2017). Acidity replaces magnesium ions in the chlorophylls with hydrogen atoms and chlorophylls are converted to pheophytins (Kusmita et al. 2015). This unexpected result can be explained by the severity of stress, which caused reduced pheophytin content, as it did in duckweed plants exposed to UV-B radiation for 7 days (Farooq et al. 2005).

The growth-restricting effects of AA observed in our study suggests that AA may affect cell viability particularly in roots, which were more susceptible to AA than shoots. We also investigated  $H_2O_2$  content in shoots, in which AA-induced reductions were less prominent than in roots.  $H_2O_2$  is a type of ROS characterized by low reactivity. It serves as a signalling molecule due to its long life span and small size (Khan et al. 2018). ROS are produced as byproducts of various metabolic pathways and scavenged by antioxidant defence systems. Under certain (stress or non-stress) conditions, ROS production is elevated in plants (Apel and Hirt 2004). Higher concentrations of  $H_2O_2$  deplete the ascorbic acid and glutathione pool, cause damage to proteins, nucleic acids and lipids and compromise cell integrity and eventually result in cell death (Khan et al. 2018). Stress conditions increase  $H_2O_2$  content and decrease cell viability in plants, while compounds that confer stress tolerance usually act inversely (Cikili et al. 2019). Hawrylak-Nowak et al. (2015) reported that AA and particularly malic acid promoted cell viability in roots of Cd-treated plants due to increased high dehydrogenase activity. They also observed that AA decreased the content of  $H_2O_2$  in leaves and roots of sunflower plants subjected to Cd stress. However, in our study, we observed the accumulation of  $H_2O_2$  in shoots and decreased cell viability in roots after AA treatment, even after recovery step. Consistently with our findings, Armstrong et al. (1996) reported necrosis of the roots of reed subjected to 1.67 mM AA. These results suggest that AA can induce the oxidative stress in shoots and reduce the root length due to cell death and imply that AA, even at low concentrations, acts as a stressor in barley. In the study by Hawrylak-Nowak et al. (2015) where opposite results were found, 7-day-old sunflower seedlings were treated with 0.25 or 0.5 mM AA in 1.5-times strength Hoagland's II nutrient solution for 14 days. Different plant species and different exper-

imental conditions, especially concerning the pH of the medium may explain the contradictory results.

Plants accumulate phenolic compounds (PCs) when exposed to stress. PCs can be found in several intracellular locations; and stimulate stress tolerance in plants by performing diverse functions such as scavenging of ROS, enhancement of cell division and improvement of photosynthesis (Parvin et al. 2022). The most abundant group of PCs is that of flavonoids, containing anthocyanins as a subgroup (Parvin et al. 2022). Flavonoids, anthocyanins, and other UV-absorbing compounds accumulate under stress conditions, although UV-B exposure was shown to be the most efficient in increasing the levels of UV-absorbing compounds and UV-B marker (Cicek et al. 2012). Depending on the plant species and its genotype, abiotic stress such as drought affects contents of flavonoids, which act as signalling molecules, antioxidant molecules and UV protectant and regulate hormones (Shin et al. 2021, Parvin et al. 2022, Kumar et al. 2023). Molecules that can enhance stress tolerance of plants can stimulate synthesis of secondary metabolites e.g. flavonoids (Kumar et al. 2023). In the present study, AA treatment increased not only anthocyanin and flavonoid contents but also UV-absorbing compounds ( $A_{300}$ ) and the UV-B marker ( $A_{440}$ ). Except for the flavonoids, the increasing trend persisted after recovery step. AA treatment immediately and directly impacted UV-absorbing pigments probably as a result of increased  $H_2O_2$  which induced synthesis of various phenolic compounds to act as ROS scavengers.

Because AA is a precursor of acetyl-CoA, an essential molecule of carbohydrate metabolism, we decided to investigate how AA treatment affects sugar contents in roots. There are contradictory reports on the effects of acetate on starch synthesis. Starch degradation is usually activated under abiotic stress to redirect carbon for stress responses, but starch can also accumulate when growth is inhibited more than photosynthesis (Ribeiro et al. 2022). Fan et al. (2012) reported acetate-induced starch accumulation in *C. reinhardtii*. However, Rengel et al. (2018) observed that overexpression of chloroplastic ACSS under nutrient replete conditions enhanced starch content in *C. reinhardtii*, while acetate treatment did not. Under nitrogen starvation, excess acetyl-CoA was stored as triacylglycerol. Interestingly, Arabidopsis plants with reduced activity of ATP-citrate lyase, which is responsible for conversion of mitochondria-derived citrate to acetyl-coA, have higher amounts of anthocyanin and starch (Fatland et al. 2005). Huang et al. (2017) found that microalgae *Chlorella sorokiniana* GXNN01 favours AA as a carbon source over glucose under normal and high light conditions. Use of AA as carbon source resulted in higher biomass with high lipid and low starch contents. In this study, soluble carbohydrate contents decreased in AA-treated roots, while insoluble carbohydrate contents increased, which is consistent with the findings of Fan et al. (2012). Moreover, changes in the content of soluble carbohydrates were more prominent than those of insoluble carbohydrates. Soluble sugars in seeds serve as fast-use

reserves for energy production but they can also be efficient in protecting membrane integrity during stress conditions (Ferreira et al. 2009).

In conclusion, even at low concentrations, AA treatment can induce dramatic and persistent changes in non-stressed plants that endure even after AA is removed. Previous studies focused on the amelioration of stress, particularly drought, by AA treatment that was usually applied to plants growing in a culture medium or soil. In this study, AA was applied to young (2-day-old) seedlings growing on a filter paper and exhibited its toxic effects *i.e.* restriction of growth, decrease in WC, photosynthetic pigments and viability in root cells and the accumulation of UV-absorbing pigments and H<sub>2</sub>O<sub>2</sub> in shoots and insoluble carbohydrates in roots.

## Acknowledgments

This work was supported by the Research Fund of Istanbul University (28371 and 30235).

## References

- Adeleke, R., Nwangburuka, C., Oboirien, B., 2017: Origins, roles and fate of organic acids in soils: A review. *South African Journal of Botany* 108, 393–406. <https://doi.org/10.1016/j.sajb.2016.09.002>
- Agathokleous, E., Feng, Z., Peñuelas, J., 2020: Chlorophyll hormesis: Are chlorophylls major components of stress biology in higher plants? *Science of the Total Environment* 726: 138637. <https://doi.org/10.1016/j.scitotenv.2020.138637>
- Allen, M. M., Allen, D. J., 2020: Biostimulant potential of acetic acid under drought stress is confounded by pH-dependent root growth inhibition. *Frontiers in Plant Science* 11, 647. <https://doi.org/10.3389/fpls.2020.00647>
- Apel, K., Hirt, H., 2004: Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annual Review of Plant Biology* 55, 373–399. <https://doi.org/10.1146/annurev.arplant.55.031903.141701>
- Armstrong, J., Afreen-Zobayed, F., Armstrong, W., 1996: Phragmites die-back: sulphide- and acetic acid-induced bud and root death, lignifications, and blockages within aeration and vascular systems. *New Phytologist* 134(4), 601–614. <https://doi.org/10.1111/j.1469-8137.1996.tb04925.x>
- Baker, C. J., Mock, N. M., 1994: An improved method for monitoring cell death in cell suspension and leaf disc assays using Evans blue. *Plant Cell Tissue and Organ Culture* 39, 7–12. <https://doi.org/10.1007/BF00037585>
- Chen, D., Shao, M., Sun, S., Liu, T., Zhang, H., Qin, N., Zeng, R., Song Y., 2019: Enhancement of jasmonate-mediated antiherbivore defense responses in tomato by acetic acid, a potent inducer for plant protection. *Frontiers in Plant Science* 10, 764. <https://doi.org/10.3389/fpls.2019.00764>
- Cicek, N., Fedina, I., Cakirlar, H., Velitchkova, M., Georgieva, K., 2012: The role of short-term high temperature pretreatment on the UV-B tolerance of barley cultivars. *Turkish Journal of Agriculture and Forestry* 36(2), 153–165. <https://doi.org/10.3906/tar-1102-15>
- Cikili, Y., Kulac, S., Samet, H., Filiz, E., 2019: Effects of exogenous nitric oxide on cadmium toxicity in black poplar (*Populus nigra*): physiological approaches. *Acta Botanica Croatica* 78(2), 116–124. <https://doi.org/10.2478/botcro-2019-0018>
- Costa, L., Vicente, A. R., Civello, P. M., Chaves, A. R., Martinez, G. A., 2006: UV-C treatment delays postharvest senescence in broccoli florets. *Postharvest Biology and Technology* 39(2), 204–210. <https://doi.org/10.1016/j.postharvbio.2005.10.012>
- Fan, J., Yan, C., Andre, C., Shanklin, J., Schwender, J., Xu, C., 2012: Oil accumulation is controlled by carbon precursor supply for fatty acid synthesis in *Chlamydomonas reinhardtii*. *Plant and Cell Physiology* 53(8), 1380–1390. <https://doi.org/10.1093/pcp/pcs082>
- Farooq, M., Shankar, U., Ray, R. S., Misra, R. B., Agrawal, N., Verma, K., Hans, R. K., 2005: Morphological and metabolic alterations in duckweed (*Spirodela polyrrhiza*) on long-term low-level chronic UV-B exposure. *Ecotoxicology and Environmental Safety* 62(3), 408–414. <https://doi.org/10.1016/j.ecoenv.2005.01.011>
- Fatland, B. L., Nikolau, B. J., Wurtele, E. S., 2005: Reverse genetic characterization of cytosolic acetyl-CoA generation by ATP-citrate lyase in Arabidopsis. *Plant Cell* 17(1), 182–203. <https://doi.org/10.1105/tpc.104.026211>
- Ferreira, C. S., Piedade, M. T., Tiné, M. A., Rossatto, D. R., Parolin, P., Buckeridge, M. S., 2009: The role of carbohydrates in seed germination and seedling establishment of *Himatanthus sucuuba*, an Amazonian tree with populations adapted to flooded and non-flooded conditions. *Annals of Botany* 104(6), 1111–1119. <https://doi.org/10.1093/aob/mcp212>
- Hawrylak-Nowak, B., Dresler, S., Matraszek, R., 2015: Exogenous malic and acetic acids reduce cadmium phytotoxicity and enhance cadmium accumulation in roots of sunflower plants. *Plant Physiology and Biochemistry* 94, 225–234. <https://doi.org/10.1016/j.plaphy.2015.06.012>
- Huang, A., Sun, L., Wu, S., Liu, C., Zhao, P., Xie, X., Wang, G., 2017: Utilization of glucose and acetate by *Chlorella* and the effect of multiple factors on cell composition. *Journal of Applied Phycology* 29, 23–33. <https://doi.org/10.1007/s10811-016-0920-6>
- Khan, T. A., Yusuf, M., Fariduddin, Q., 2018: Hydrogen peroxide in regulation of plant metabolism: Signalling and its effect under abiotic stress. *Photosynthetica* 56(4), 1237–1248. <https://doi.org/10.1007/s11099-018-0830-8>
- Kim, J. M., To, T. K., Matsui, A., Tanoi, K., Kobayashi, N. I., Matsuda, F., Habu, Y., Ogawa, D., Sakamoto, T., Matsunaga, S., et al., 2017: Acetate-mediated novel survival strategy against drought in plants. *Nature Plants* 3, 17097. <https://doi.org/10.1038/nplants.2017.97>
- Kumar, K., Debnath, P., Singh, S., Kumar, N., 2023: An overview of plant phenolics and their involvement in abiotic stress tolerance. *Stresses* 3(3), 570–585. <https://doi.org/10.3390/stresses3030040>
- Kusmita, L., Puspitaningrum, I., Limantara, L., 2015: Identification, isolation and antioxidant activity of pheophytin from green tea (*Camellia sinensis*, (L.) Kuntze). *Procedia Chemistry* 14, 232–238. <https://doi.org/10.1016/j.proche.2015.03.033>
- Li, M., Gao, S., Luo, J., Cai, Z., Zhang, S., 2022: Mitigation effects of exogenous acetic acid on drought stress in *Cunninghamia lanceolata*. *Plant and Soil*. <https://doi.org/10.1007/s11104-022-05628-6>
- Lichtenthaler, H. K., 1987: Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Methods in Enzymology* 148, 350–382. [https://doi.org/10.1016/0076-6879\(87\)48036-1](https://doi.org/10.1016/0076-6879(87)48036-1)
- Lin Y. P., Charng Y. Y., 2021: Chlorophyll dephytylation in chlorophyll metabolism: a simple reaction catalyzed by various enzymes. *Plant Science* 302: 110682. <https://doi.org/10.1016/j.plantsci.2020.110682>
- López-Bucio, J., Nieto-Jacobo, M. F., Ramírez-Rodríguez, V., Herrera-Estrella, L., 2000: Organic acid metabolism in plants:

- from adaptive physiology to transgenic varieties for cultivation in extreme soils. *Plant Science* 160(1), 1–13. [https://doi.org/10.1016/s0168-9452\(00\)00347-2](https://doi.org/10.1016/s0168-9452(00)00347-2)
- Lynch, J. M., 1977: Phytotoxicity of acetic acid produced in the anaerobic decomposition of wheat straw. *Journal of Applied Bacteriology* 42(1), 81–87. <https://doi.org/10.1111/j.1365-2672.1977.tb00672.x>
- Mahmud, S., Kamruzzaman, M., Bhattacharyya, S., Alharbi, K., Abd El Moneim, D., Mostofa, M. G., 2023: Acetic acid positively modulates proline metabolism for mitigating PEG-mediated drought stress in maize and *Arabidopsis*. *Frontiers in Plant Science* 14, 1167238. <https://doi.org/10.3389/fpls.2023.1167238>
- Masuko, T., Minami, A., Iwasaki, N., Majima, T., Nishimura, S. I., Lee, Y. C. 2005: Carbohydrate analysis by a phenol-sulfuric acid method in microplate format. *Analytical Biochemistry* 339(1), 69–72. <https://doi.org/10.1016/j.ab.2004.12.001>
- McCready, R. M., Guggolz, J., Silviera, V., Owens, H. S. 1950: Determination of starch and amylose in vegetables. *Analytical Chemistry* 22(9), 1156–1158. <https://doi.org/10.1021/ac60045a016>
- Parvin, K., Nahar, K., Mohsin, S. M., Al Mahmud, J., Fujita, M., Hasanuzzaman, M., 2022: Plant Phenolic Compounds for Abiotic Stress Tolerance. In: Hasanuzzaman, M., Ahammed, G. J., Nahar, K. (eds.) *Managing Plant Production Under Changing Environment*. 193–237. Springer, Singapore. [https://doi.org/10.1007/978-981-16-5059-8\\_8](https://doi.org/10.1007/978-981-16-5059-8_8)
- Pietrocola, F., Galluzzi, L., Bravo-San Pedro, J. M., Madeo, F., Kroemer, G., 2015: Acetyl coenzyme A: A central metabolite and second messenger. *Cell Metabolism* 21(6), 805–821. <https://doi.org/10.1016/j.cmet.2015.05.014>
- Rahman, M., Mostofa, M. G., Keya, S. S., Rahman, A., Das, A. K., Islam, R., Abdelrahman, M., Bhuiyan, S. U., Naznin, T., Ansary, M. U., Tran, L. P. 2021: Acetic acid improves drought acclimation in soybean: an integrative response of photosynthesis, osmoregulation, mineral uptake and antioxidant defense. *Physiologia Plantarum* 172(2), 334–350. <https://doi.org/10.1111/ppl.13124>
- Rahman, M. M., Mostofa, M. G., Rahman, M. A., Islam M. R., Keya, S. S., Das, A. K., Miah, M. G., Kawser, A. Q. M. R. S., Ahsan, M., Hashem, A., Tabassum, B., AbdAllah, E. F., Tran, L. S. P., 2019: Acetic acid: a cost-effective agent for mitigation of seawater-induced salt toxicity in mung bean. *Scientific Reports* 9, 15186. <https://doi.org/10.1038/s41598-019-51178-w>
- Rengel, R., Smith, R. T., Haslam, R., Sayanova, O., Vila, M., León, R., 2018: Overexpression of acetyl-CoA synthetase (ACS) enhances the biosynthesis of neutral lipids and starch in the green microalga *Chlamydomonas reinhardtii*. *Algal Research* 31, 183–193. <https://doi.org/10.1016/j.algal.2018.02.009>
- Ribeiro, C., Stitt, M., Hotta, C. T., 2022: How stress affects your budget-stress impacts on starch metabolism. *Frontiers in Plant Science* 13, 774060. <https://doi.org/10.3389/fpls.2022.774060>
- Shin, Y. K., Bhandari, S. R., Jo, J. S., Song, J. W., Lee, J. G., 2021: Effect of drought stress on chlorophyll fluorescence parameters, phytochemical contents, and antioxidant activities in lettuce seedlings. *Horticulturae* 2021, 7(8), 238. <https://doi.org/10.3390/horticulturae7080238>
- Sonjaroon, W., Jutamane, K., Khamsuk, O., Thussagunpanit, J., Kaveeta, L., Suksamrarn, A., 2018: Impact of brassinosteroid mimic on photosynthesis, carbohydrate content and rice seed set at reproductive stage under heat stress. *Agriculture and Natural Resources* 52(3), 234–240. <https://doi.org/10.1016/j.anres.2018.09.001>
- Szafrańska, K., Reiter, R. J., Posmyk, M. M., 2017: Melatonin improves the photosynthetic apparatus in pea leaves stressed by paraquat via chlorophyll breakdown regulation and its accelerated *de novo* synthesis. *Frontiers in Plant Science* 8, 878. <https://doi.org/10.3389/fpls.2017.00878>
- Takamiya, K. I., Tsuchiya, T., Ohta, H., 2000: Degradation pathway(s) of chlorophyll: what has gene cloning revealed? *Trends in Plant Science* 5(10), 426–431. [https://doi.org/10.1016/S1360-1385\(00\)01735-0](https://doi.org/10.1016/S1360-1385(00)01735-0)
- Utsumi, Y., Utsumi, C., Tanaka, M., Takahashi, S., Matsui, A., Matsunaga, T. M., Matsunaga, S., Okamoto, Y., Moriya, E., Seki, M., 2019: Acetic acid treatment enhances drought avoidance in cassava (*Manihot esculenta* Crantz). *Frontiers in Plant Science* 10, 521. <https://doi.org/10.3389/fpls.2019.00521>
- Zuo, Z., Zhu, Y., Bai, Y., Wang, Y., 2012: Acetic acid-induced programmed cell death and release of volatile organic compounds in *Chlamydomonas reinhardtii*. *Plant Physiology and Biochemistry* 51, 175–184. <https://doi.org/10.1016/j.plaphy.2011.11.003>

# Trehalose-induced metabolic responses in basil (*Ocimum basilicum*) seedlings under salt treatment

Ramazan Karamzahi, Alireza Einali\*

Department of Biology, Faculty of Science, University of Sistan and Baluchestan, Zahedan 98167, Iran

**Abstract** – Trehalose (Tre) is an osmoprotectant known to be an important player in regulating response to salinity. In this research, the effect of Tre foliar application on the growth rate, the content of photosynthetic pigments, accumulation of metabolites, and activity of antioxidant enzymes of basil seedlings under salt stress has been investigated. Basil seedlings were factorially treated twice a week with levels of 0, 25, 50, 100, and 150 mM NaCl and weekly with concentrations of 0 and 5 mM Tre for 4 weeks. Growth characteristics, the content of photosynthetic pigments including chlorophyll and carotenoid, and starch content decreased in response to salt stress, while the activity of antioxidant enzymes and the accumulation of metabolites including soluble sugars, proteins, amino acids, and proline increased. Tre treatment caused severe inhibition of plant growth, further reduction of photosynthetic pigments, and amount of soluble proteins during salinity stress. Also, starch, total protein, amino acids, and proline were hyperaccumulated in response to Tre. These results indicate that Tre not only does not reduce the detrimental effects of salinity in basil seedlings but also inhibits plant growth possibly by diverting carbon to other metabolic pathways.

**Keywords:** proline, salt stress, soluble sugars, starch, trehalose

## Introduction

Salinity is one of the most important factors limiting the yield of crops, especially in arid, semi-arid, and coastal areas (Mostofa et al. 2015). This stress negatively affects plant physiology by imposing several major limitations leading to reduced crop growth and productivity (Ali et al. 2021, Zhang et al. 2021, Khan et al. 2023). Salinity causes osmotic and ionic stress, and production of reactive oxygen species (ROS) in plant cells (Ahammed et al. 2018, Khan et al. 2023, Colin et al. 2023). High production of ROS induces oxidative stress, which can destroy membrane components and biomolecules and, at higher levels, cause cell, tissue, and plant death (Kaur et al. 2022, Khan et al. 2023, Peng et al. 2023). Salinity tolerance is a complex process that involves various molecular, physiological, and biochemical mechanisms (Zhang and Shi 2013, Islam et al. 2023).

The use of osmoprotectants can augment stress tolerance mechanisms in plants (Chen and Jiang 2010, Islam et al. 2023). These compounds, which are known as compatible solutes, are highly soluble compounds with low molecular weight that directly or indirectly protect plants against stresses through various mechanisms such as regulating cellular osmosis, preventing membrane damage, stabilizing proteins and enzymes, and eliminating ROS (Singh et al. 2015, He et al. 2018). Application of exogenous osmoprotectants

can be considered an alternative approach to improve plant productivity under saline conditions (Nakayama et al. 2005). The high concentration of compatible solutes can balance the salts entering the cell and at the same time deal with the high concentration of salts inside the cell (Türkan and Demiral 2009).

Sugars are one of the most important compatible solutes that, in addition to the role of signal molecules, can act as a source of metabolic energy and play a role in regulating metabolism in plants (Islam and Mohammad 2021). Trehalose (Tre) is a non-reducing disaccharide consisting of two glucose molecules with a chemically non-reactive nature that makes it compatible with cellular metabolism even at high concentrations, and plays an important role as an osmoprotectant in stress tolerance of some plants (Ali and Ashraf 2011, Lunn et al. 2014). It is naturally distributed in many organisms, from bacteria and fungi to plants and invertebrates (Nounjan et al. 2012, Kosar et al. 2019), where it is used for carbon storage and transport as well as being an osmoprotectant (Lunn et al. 2014). Among plants, Tre synthesis has been reported in some resurrection plants and other plants such as *Arabidopsis* (Muller et al. 1999, Lunn et al. 2014). It has been shown that the expression of plant genes involved in Tre metabolism undergoes remarkable changes

\* Corresponding author e-mail: aeinali@science.usb.ac.ir

in response to various abiotic stresses (Nakashima et al. 2009). Tre and trehalose-6-phosphate, a phosphorylated intermediate in Tre biosynthesis, act as signal molecules in carbohydrate metabolism (Wingler et al. 2000, Lunn et al. 2014). Tre induces sucrose synthase activity in soybean (Muller et al. 1998) and trehalose-6-phosphate acts as a specific signal of sucrose status in *Arabidopsis*, and may be seen as part of a homeostatic mechanism to control the level of sucrose, as well as part of regulatory networks in sucrose export in source organs and in growing sink organs (Lunn et al. 2014). Tre has also a key role in the control of metabolism during plant growth and development, and reduces the effects of stress through mechanisms such as membrane protection, stabilization of proteins and enzymes, detoxification and removal of ROS, molecular signaling, and increasing the accumulation of osmolytes (Luo et al. 2010, Abdallah et al. 2016, Islam and Mohammad 2021). Most studies on the role of osmoprotectants in enhancing stress tolerance have focused on proline and glycine betaine, but there is relatively little knowledge about the effects of Tre on higher plants under stress conditions. Some studies have shown that the exogenous application of Tre on plants has induced tolerance to salinity stress (Nounjan et al. 2012, Abdallah et al. 2016, Rohman et al. 2019). In contrast, it has been found that Tre can have an inhibitory effect on the growth of some plants (Schluepmann et al. 2004, O'Hara et al. 2013) and algae under salinity (Panjekobi and Einali 2021). However, in most plants, endogenous Tre cannot adequately ameliorate the harmful effects of stress (Mostofa et al. 2015).

Basil (*Ocimum basilicum* L.) is a medicinal and aromatic plant that is used in the pharmaceutical and food industries due to its abundant essential oils, high content of phyto-medicines and natural antioxidant molecules, and is usually produced for economic purposes (Kwee and Niemeyer 2011, Caliskan et al. 2017). It is commonly used in traditional medicine and herbal therapy (Bahcesular et al. 2020, Farouk and Omar 2020). To date, no study has been conducted on the role of Tre in the physiological and biochemical responses of basil, an important agricultural and medicinal plant, to salinity stress. Accordingly, basil was taken as a model system to determine the effect of exogenous Tre on growth, accumulation of osmolytes, and the activity of some enzymes of the antioxidant system under salt stress.

## Materials and methods

### Plant material and experimental design

Basil seeds (*Ocimum basilicum* L.) obtained from the Botanic Garden at the University of Sistan and Baluchestan were sown in a greenhouse with a temperature of  $29 \pm 1$  °C in trays containing moist cocopeat. After germination, seedlings with two or three leaves that had grown uniformly were selected and each seedling was transferred to a 14 x12 cm plastic pot containing 1 kg of cocopeat and placed at the same temperature. Seedlings were irrigated with 1/2 Hoagland's standard nutrient solution at three-day intervals until reaching the 6-8 leaf stage. Three days before salt treatment, the seedlings were divided into two parts. By foliar spraying

on the seedlings, one part was treated with a concentration of 5 mM Tre and the other part received only distilled water. Salinity treatment was applied by adding sodium chloride (NaCl) in concentrations of 0, 25, 50, 100, and 150 mM to the nutrient solutions of each plant group. Tre-untreated plants, which received only 1/2 Hoagland's solution without NaCl, were considered as the control for salinity experiments. Seedlings were treated twice a week with salinity and every week with Tre for four weeks. After this period, seedlings were harvested and studied to investigate the morphological and biochemical responses to salt stress and Tre treatment. Each morphological or biochemical experiment was performed individually with three biological replicates.

### Morphological traits

The length of root and shoot along with the length, width, and number of leaves of basil seedlings were measured. To determine the amount of biomass, first the fresh weight (FW) of each shoot and root was measured, and after exposure to 70 °C for 72 h, the dry weight (DW) was determined. Shoot water content (SWC) of basil seedlings was obtained using the following formula:

$$\text{SWC} = ((\text{shoot FW} - \text{shoot DW}) / \text{shoot FW}) \times 100$$

### Determination of photosynthetic pigments content

Photosynthetic pigments including chlorophyll (Chl) and total carotenoids (Car) were extracted from 1 g of fresh leaf tissue with 80% acetone. The resulting mixture was filtered through filter paper (Whatman No. 1) and the residue was saved to measure soluble sugars and total protein. The absorbance of the filtrate was recorded at 663, 645, and 652 nm, and the amounts of Chl<sub>a</sub>, Chl<sub>b</sub>, and total were measured using the following equations (Arnon 1949):

$$\text{Chl } a \text{ (mg mL}^{-1}\text{)} = 0.0127 \times A_{663} - 0.00269 \times A_{645}$$

$$\text{Chl } b \text{ (mg mL}^{-1}\text{)} = 0.0229 \times A_{645} - 0.00468 \times A_{663}$$

$$\text{Total Chl (mg mL}^{-1}\text{)} = A_{652} / 34.5$$

Total Car content was measured at 470 nm using the following equation (Lichtenthaler and Buschmann, 2001):

$$\text{Total Car (}\mu\text{g mL}^{-1}\text{)} = (1000 \times A_{470} - 1.82 \times \text{Chl } a - 85.02 \times \text{Chl } b) / 198$$

The content of pigments was expressed as mg per g FW.

### Determination of soluble sugars and starch content

Soluble sugars, including reducing and non-reducing sugars, were extracted from 40 mg of acetone powder (resulting from the extraction of photosynthetic pigments) using 80% ethanol (Einali and Valizadeh 2017). Ethanol extracts were used to determine the content of soluble sugars. Reducing sugars (RS) were determined by the method of Miller (1959) and non-reducing sugars (NRS) were measured by Handel's (1968) method. The residues obtained from the extraction of soluble sugars were used to extract and measure the amount of starch (McCready et al. 1950). The content of soluble sugars and starch was expressed as mg per g FW.



### Determination of total proteins, total amino acid, and proline content

Total proteins refer to proteins extracted with a sample buffer containing 60 mM Tris-HCl buffer (pH 6.8), 10% (v/v) glycerol, and 2% (w/v) sodium dodecyl sulfate (Stone and Gifford (1997). Extraction of total proteins from 20 mg of acetone powder was done with 0.5 mL of sample buffer at 90 °C for 1 h, followed by centrifugation at 10000 g for 15 minutes (Alisofi et al. 2020). The amount of total proteins was measured at a wavelength of 750 nm by the method of Markwell et al. (1981) and expressed as mg per g FW.

Free amino acids and proline were extracted from 0.2 g of fresh leaf tissue using 80% ethanol at 70 °C for 10 min, followed by centrifugation at 2000 g for 10 min (Einali and Valizadeh 2017). The extraction process was repeated four more times. The concentrated ethanol extract was decolorized by chloroform (1:5, v/v). The ninhydrin (1% (w/v) ninhydrin and 0.06% (w/v) KCN in acetone) method using a glycine calibration curve at 570 nm was used to determine total free amino acid (Yemm and Cocking 1955). Proline content was measured at 520 nm by another ninhydrin (1% (w/v) ninhydrin in 60% (v/v) acetic acid) method using proline as a standard (Bates et al. 1973). The content of free amino acids and proline was expressed as  $\mu\text{mol}$  per g FW.

### Soluble protein extraction and enzyme assays

Crude enzyme extract was prepared from 0.2 g of fresh leaf tissue with 3 mL of enzyme extraction buffer containing 100 mM cold potassium phosphate buffer (pH 7.0), 10% glycerol, 1 mM EDTA, 10 mM KCl, 1 mM  $\text{MgSO}_4$ , 1 mM phenylmethylsulphonyl fluoride (PMSF), 50 mM 2-mercaptoethanol, 0.1% (v/v) Triton X-100, and 1% (w/v) polyvinylpyrrolidone (PVPP) as described elsewhere (Alisofi et al. 2020). Extraction was done in a cold mortar with pestle. The extraction buffer for ascorbate peroxidase (APX) determination contained 5 mM ascorbic acid as well. The homogenate was filtered through four layers of cheesecloth and incubated at 4 °C for 1 h. The amount of 10 mg of charcoal was added to the filter to remove the extracted pigments, and then it was centrifuged at 12000 g for 10 min at 4 °C. The supernatant,

containing soluble protein fraction, was used for enzyme assays. Soluble proteins refer to proteins extracted in the absence of sodium dodecyl sulfate. Soluble protein content was measured by Bradford's (1976) method using the albumin standard curve and expressed as mg per g FW.

APX reaction mixture (1 mL) consisted of 50 mM potassium phosphate buffer (pH 7.0), 1 mM  $\text{H}_2\text{O}_2$ , 0.5 mM ascorbic acid, and 50  $\mu\text{L}$  enzyme extract. The oxidation of ascorbate to dehydroascorbate was monitored at 290 nm and the activity of the enzyme was calculated using the extinction coefficient  $2.8 \text{ mM}^{-1} \text{ cm}^{-1}$  and expressed as  $\mu\text{mol}$  of oxidized ascorbate per min per g FW (Chen and Asada 1992).

The activity of pyrogallol peroxidase (PPX) was measured in a reaction mixture (1 mL) containing 50 mM potassium phosphate buffer (pH 7.0), 40 mM pyrogallol, 1 mM  $\text{H}_2\text{O}_2$ , and 50  $\mu\text{L}$  enzyme extract. The conversion rate of pyrogallol to purpurogallin was monitored at 430 nm and the enzyme activities were calculated using the extinction coefficient  $2.47 \text{ mM}^{-1} \text{ cm}^{-1}$  and expressed as  $\mu\text{mol}$  of purpurogallin produced per min per g FW (Nakano and Asada 1981). Polyphenol oxidase (PPO) activity was measured in the same way as PPX at 430 nm using the extinction coefficient  $2.47 \text{ mM}^{-1} \text{ cm}^{-1}$ , except that the reaction mixture was without  $\text{H}_2\text{O}_2$  (Nakano and Asada 1981).

### Statistical analysis

All results obtained from plant growth and biochemical studies were expressed as mean and standard deviation (SD) of three independent measurements. The statistically significant difference between treatments was determined in the form of factorial design using two-way ANOVA and Duncan's test at the level of 5% ( $P < 0.05$ ).

## Results

### Effect of salinity and Tre on plant growth

Salt treatment at low concentration (25 mM) caused a clear 15% increase in shoot length compared to the control, but higher salt concentrations gradually decreased these values (Tab. 1).

**Tab. 1.** Longitudinal growth and leaf characteristics of trehalose-treated (+Tre) or untreated (–Tre) basil seedlings in response to different salinity concentrations. Values are the mean  $\pm$  standard deviation of three separate measurements. Different letters in each column indicate significant differences between the various treatments at  $P < 0.05$  according to the Duncan test. LL – leaf length; LW – leaf weight.

Salt treatment (mM)		Plant length (cm)		LL (cm)	LW (cm)	LL/LW	LL $\times$ LW ( $\text{cm}^2$ )	Number of leaves
		Shoot	Root					
0	–Tre	26.00 $\pm$ 1.00 <sup>b</sup>	19.67 $\pm$ 2.08 <sup>a</sup>	3.77 $\pm$ 0.46 <sup>a</sup>	1.93 $\pm$ 0.12 <sup>a</sup>	1.95 $\pm$ 0.20 <sup>a</sup>	7.30 $\pm$ 1.18 <sup>a</sup>	14.00 $\pm$ 1.00 <sup>ab</sup>
	+Tre	23.33 $\pm$ 0.58 <sup>c</sup>	22.67 $\pm$ 2.08 <sup>a</sup>	2.67 $\pm$ 0.29 <sup>bc</sup>	1.63 $\pm$ 0.06 <sup>b</sup>	1.64 $\pm$ 0.21 <sup>a</sup>	4.35 $\pm$ 0.41 <sup>c</sup>	14.00 $\pm$ 0.00 <sup>a</sup>
25	–Tre	30.00 $\pm$ 1.00 <sup>a</sup>	20.00 $\pm$ 2.00 <sup>a</sup>	4.33 $\pm$ 0.76 <sup>a</sup>	2.10 $\pm$ 0.17 <sup>a</sup>	2.07 $\pm$ 0.39 <sup>a</sup>	9.12 $\pm$ 1.84 <sup>a</sup>	16.00 $\pm$ 0.00 <sup>a</sup>
	+Tre	24.00 $\pm$ 2.65 <sup>c</sup>	21.00 $\pm$ 1.00 <sup>a</sup>	2.93 $\pm$ 0.31 <sup>bc</sup>	1.70 $\pm$ 0.10 <sup>b</sup>	1.73 $\pm$ 0.18 <sup>a</sup>	4.99 $\pm$ 0.69 <sup>bc</sup>	14.67 $\pm$ 1.15 <sup>ab</sup>
50	–Tre	26.33 $\pm$ 2.08 <sup>b</sup>	21.00 $\pm$ 1.00 <sup>a</sup>	4.03 $\pm$ 0.45 <sup>a</sup>	2.23 $\pm$ 0.21 <sup>a</sup>	1.80 $\pm$ 0.07 <sup>a</sup>	9.07 $\pm$ 1.80 <sup>a</sup>	15.33 $\pm$ 1.15 <sup>a</sup>
	+Tre	21.33 $\pm$ 1.53 <sup>d</sup>	21.33 $\pm$ 1.53 <sup>a</sup>	3.13 $\pm$ 0.15 <sup>b</sup>	1.67 $\pm$ 0.15 <sup>b</sup>	1.89 $\pm$ 0.15 <sup>a</sup>	5.23 $\pm$ 0.63 <sup>bc</sup>	14.67 $\pm$ 1.15 <sup>ab</sup>
100	–Tre	24.33 $\pm$ 0.58 <sup>c</sup>	23.00 $\pm$ 2.65 <sup>a</sup>	3.87 $\pm$ 0.32 <sup>a</sup>	2.13 $\pm$ 0.21 <sup>a</sup>	1.81 $\pm$ 0.03 <sup>a</sup>	8.29 $\pm$ 1.46 <sup>a</sup>	15.33 $\pm$ 1.15 <sup>a</sup>
	+Tre	20.33 $\pm$ 2.08 <sup>d</sup>	19.33 $\pm$ 1.15 <sup>a</sup>	3.20 $\pm$ 0.10 <sup>b</sup>	1.77 $\pm$ 0.06 <sup>b</sup>	1.81 $\pm$ 0.08 <sup>a</sup>	5.65 $\pm$ 0.26 <sup>b</sup>	14.67 $\pm$ 1.15 <sup>ab</sup>
150	–Tre	21.33 $\pm$ 1.53 <sup>d</sup>	21.00 $\pm$ 0.00 <sup>a</sup>	3.50 $\pm$ 0.30 <sup>ab</sup>	1.93 $\pm$ 0.12 <sup>a</sup>	1.81 $\pm$ 0.19 <sup>a</sup>	6.77 $\pm$ 0.72 <sup>a</sup>	13.33 $\pm$ 1.15 <sup>b</sup>
	+Tre	14.67 $\pm$ 1.53 <sup>e</sup>	19.67 $\pm$ 1.53 <sup>a</sup>	2.70 $\pm$ 0.17 <sup>c</sup>	1.37 $\pm$ 0.15 <sup>c</sup>	1.98 $\pm$ 0.11 <sup>a</sup>	3.71 $\pm$ 0.63 <sup>d</sup>	13.33 $\pm$ 1.15 <sup>b</sup>

**Tab. 2.** Shoot and root biomass accumulation and water content of trehalose-treated (+Tre) or untreated (-Tre) basil seedlings in response to different salinity concentrations. Values are the mean ± standard deviation of three separate measurements. Different letters in each column indicate significant differences between the various treatments at P < 0.05 according to the Duncan test. FW – fresh weight; DW – dry weight; SWC – shoot water content.

Salt treatment (mM)	FW (g)		DW (g)		FW/DW		SWC (%)	
	Shoot	Root	Shoot	Root	Shoot	Root		
0	-Tre	2.19 ± 0.08 <sup>a</sup>	0.43 ± 0.04 <sup>a</sup>	0.60 ± 0.01 <sup>a</sup>	0.05 ± 0.01 <sup>c</sup>	3.64 ± 0.08 <sup>c</sup>	9.50 ± 2.20 <sup>a</sup>	72.49 ± 0.58 <sup>d</sup>
	+Tre	1.06 ± 0.16 <sup>c</sup>	0.18 ± 0.02 <sup>d</sup>	0.23 ± 0.07 <sup>de</sup>	0.08 ± 0.01 <sup>b</sup>	4.76 ± 1.04 <sup>b</sup>	2.26 ± 0.43 <sup>e</sup>	78.35 ± 4.49 <sup>cd</sup>
25	-Tre	1.99 ± 0.31 <sup>ab</sup>	0.31 ± 0.03 <sup>b</sup>	0.52 ± 0.01 <sup>b</sup>	0.10 ± 0.01 <sup>b</sup>	3.83 ± 0.62 <sup>bc</sup>	3.21 ± 0.21 <sup>d</sup>	73.41 ± 4.20 <sup>d</sup>
	+Tre	1.12 ± 0.25 <sup>c</sup>	0.26 ± 0.04 <sup>bc</sup>	0.25 ± 0.05 <sup>de</sup>	0.06 ± 0.01 <sup>c</sup>	4.61 ± 0.91 <sup>b</sup>	4.66 ± 0.82 <sup>b</sup>	77.67 ± 4.90 <sup>cd</sup>
50	-Tre	2.29 ± 0.31 <sup>a</sup>	0.32 ± 0.02 <sup>b</sup>	0.43 ± 0.06 <sup>c</sup>	0.12 ± 0.01 <sup>a</sup>	5.29 ± 0.09 <sup>b</sup>	2.74 ± 0.30 <sup>e</sup>	81.09 ± 0.31 <sup>c</sup>
	+Tre	1.64 ± 0.14 <sup>b</sup>	0.16 ± 0.03 <sup>de</sup>	0.27 ± 0.03 <sup>d</sup>	0.06 ± 0.02 <sup>c</sup>	6.19 ± 0.79 <sup>b</sup>	2.80 ± 0.55 <sup>e</sup>	83.65 ± 2.26 <sup>c</sup>
100	-Tre	1.71 ± 0.21 <sup>b</sup>	0.24 ± 0.04 <sup>c</sup>	0.41 ± 0.08 <sup>c</sup>	0.09 ± 0.01 <sup>b</sup>	4.28 ± 0.91 <sup>c</sup>	2.52 ± 0.28 <sup>e</sup>	76.00 ± 4.75 <sup>cd</sup>
	+Tre	1.38 ± 0.15 <sup>b</sup>	0.29 ± 0.05 <sup>bc</sup>	0.23 ± 0.02 <sup>e</sup>	0.06 ± 0.00 <sup>c</sup>	6.03 ± 0.60 <sup>b</sup>	4.79 ± 0.94 <sup>b</sup>	83.31 ± 1.57 <sup>bc</sup>
150	-Tre	1.10 ± 0.04 <sup>c</sup>	0.13 ± 0.02 <sup>e</sup>	0.30 ± 0.01 <sup>d</sup>	0.07 ± 0.01 <sup>c</sup>	3.73 ± 0.20 <sup>c</sup>	1.84 ± 0.02 <sup>f</sup>	73.13 ± 1.41 <sup>d</sup>
	+Tre	1.01 ± 0.04 <sup>c</sup>	0.08 ± 0.01 <sup>f</sup>	0.11 ± 0.02 <sup>f</sup>	0.02 ± 0.00 <sup>d</sup>	9.56 ± 0.96 <sup>a</sup>	3.86 ± 0.39 <sup>c</sup>	89.47 ± 1.11 <sup>a</sup>

However, salinity did not affect root length and leaf characteristics including length (LL), width (LW), LL/LW ratio, LL × LW production, and number of leaves. Tre treatment alone or under salt stress had a significant negative effect on the length of the shoot and some leaf characteristics compared to untreated controls, but it did not change the root length and the number of leaves (Tab. 1).

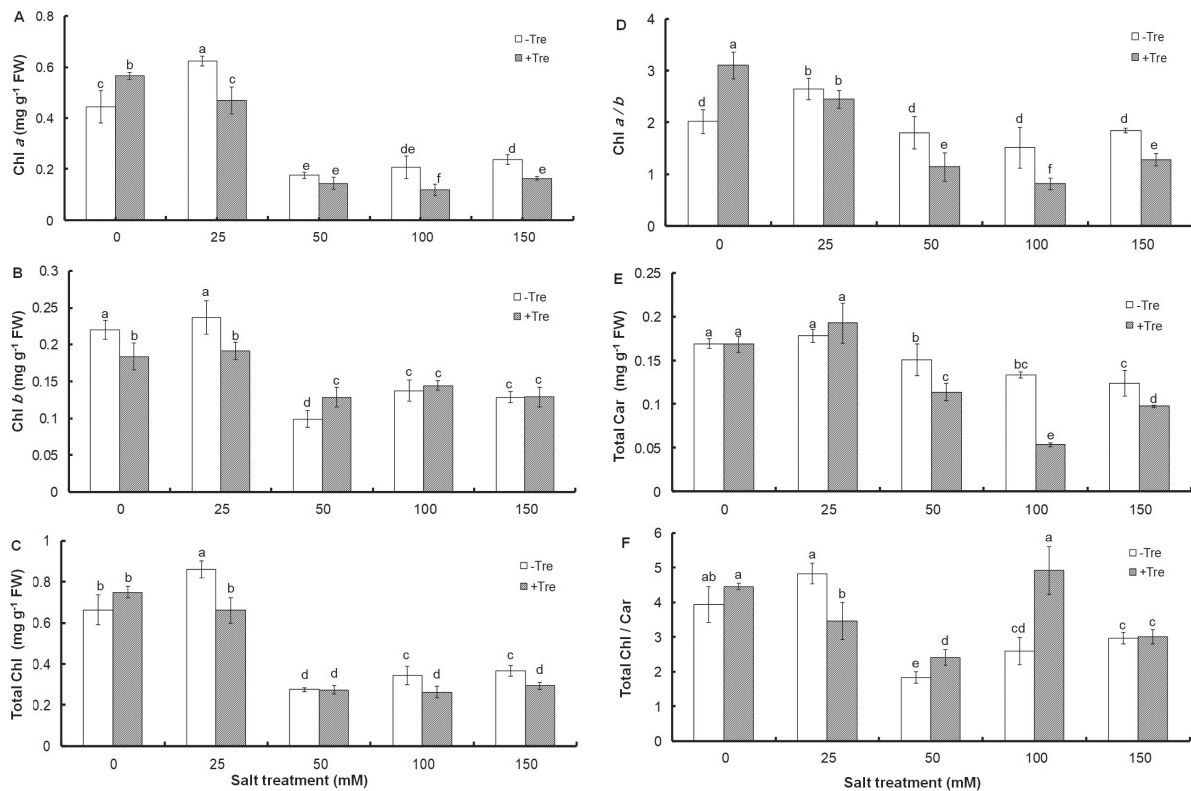
Although SWC remained unchanged, the effect of salinity on fresh and dry weight of shoots and fresh weight of roots was negative and caused a decrease proportional to salt concentration, while the dry weight of roots increased

significantly in response to different salt treatments compared to the control (Tab. 2).

Tre application in most salt treatments caused a decrease in fresh and dry weight of shoot and root, so that the decrease in dry weight was more intense. In contrast, Tre treatment caused a 23% increase in SWC in seedlings exposed to 150 mM NaCl compared to the untreated control (Tab. 2).

**Effect of salinity and Tre on photosynthetic pigments**

The amount of Chl and Car in response to salinity of 50 mM and above decreased significantly compared to the control (Fig. 1).



**Fig. 1.** Effect of different salinity concentrations on content of Chl a (A), Chl b (B), and total Chl (C), as well as on Chl a / b ratio (D), total Car (E), and Chl / Car ratio (F) in trehalose-treated (+Tre) or untreated (-Tre) basil seedlings. Results are the mean ± standard deviation of three separate measurements. Different letters indicate significant differences between the various treatments at P < 0.05 according to the Duncan test.

Although the 25 mM concentration of NaCl had no effect on the amount of Chl *b* and total Car compared to the control (Fig. 1B, 1D), it caused a significant increase in the content of Chl *a* and total Chl by 41 and 30%, respectively (Fig. 1A, 1C). A significant or non-significant increase in response to 25 mM salinity was observed in Chl *a/b* and Chl/Car ratios, respectively, which decreased or remained unchanged at higher salinities compared to the control (Fig. 1D, 1E). Tre treatment resulted in a significant decrease in pigments amount and Chl *a/b* ratio at most salt levels compared to untreated controls (Fig. 1A-1E). However, the Chl/Car ratio increased in salt treatments with a NaCl concentration above 25 mM (Fig. 1F).

### Effect of salinity and Tre on soluble sugars and starch contents

All salt treatments caused a sharp increase in the content of soluble sugars, including reducing, non-reducing and total sugars, compared to the control (Fig. 2A-2C), so that total soluble sugars reached 145.81 mg g<sup>-1</sup> FW under 150 mM NaCl (Fig. 2C).

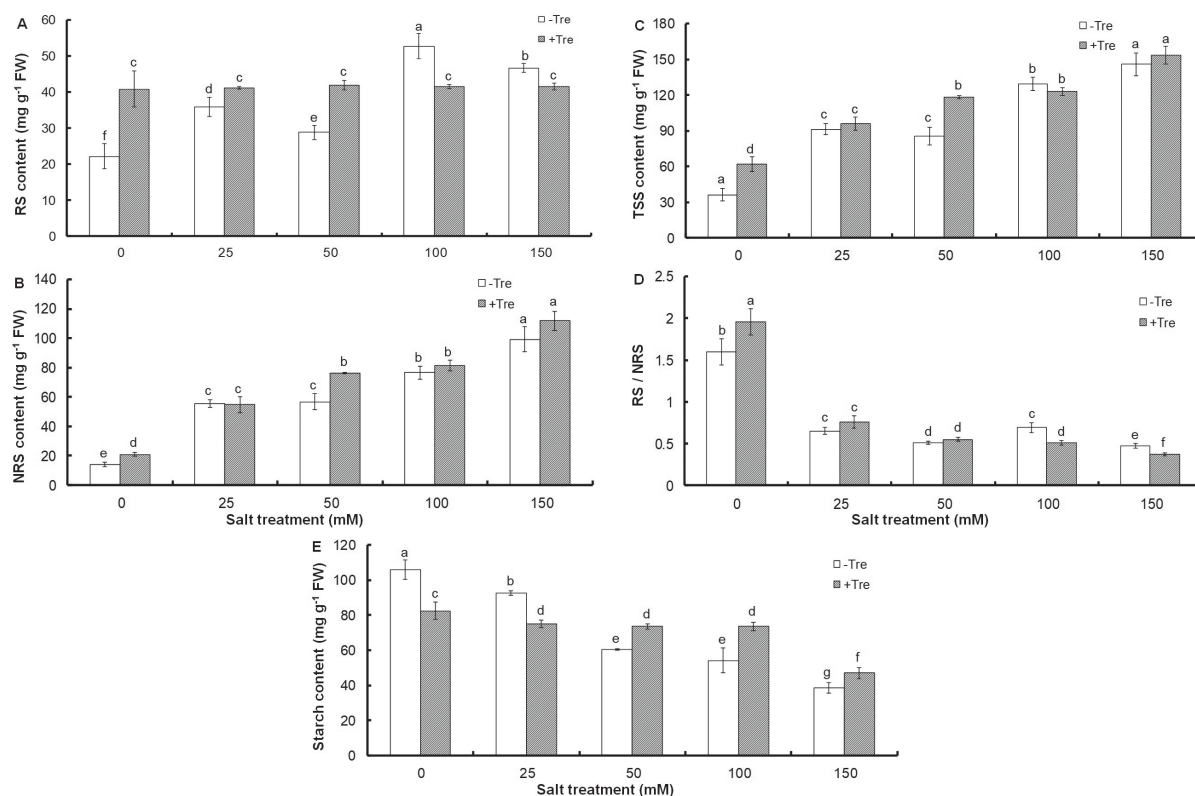
However, a significant decrease in RS/NRS ratio and starch content was found under all salinities compared to control (Fig. 2D, 2E). Tre treatment increased soluble sugars in non-stress or salinity conditions up to 50 mM NaCl concentration compared to untreated plants, but at higher salt concentrations, it caused a slight increase in NRS and

a significant decrease in RS (Fig. 2A-2C). Tre alone increased the ratio of RS/NRS by 22% compared to the untreated control, but did not change this ratio at mild salinity (25 and 50 mM NaCl) or even decreased it to the same extent at 100 and 150 mM NaCl (Fig. 2D). These changes were associated with the accumulation of starch in Tre-treated seedlings at salt concentrations of 50 mM and higher (Fig. 2E).

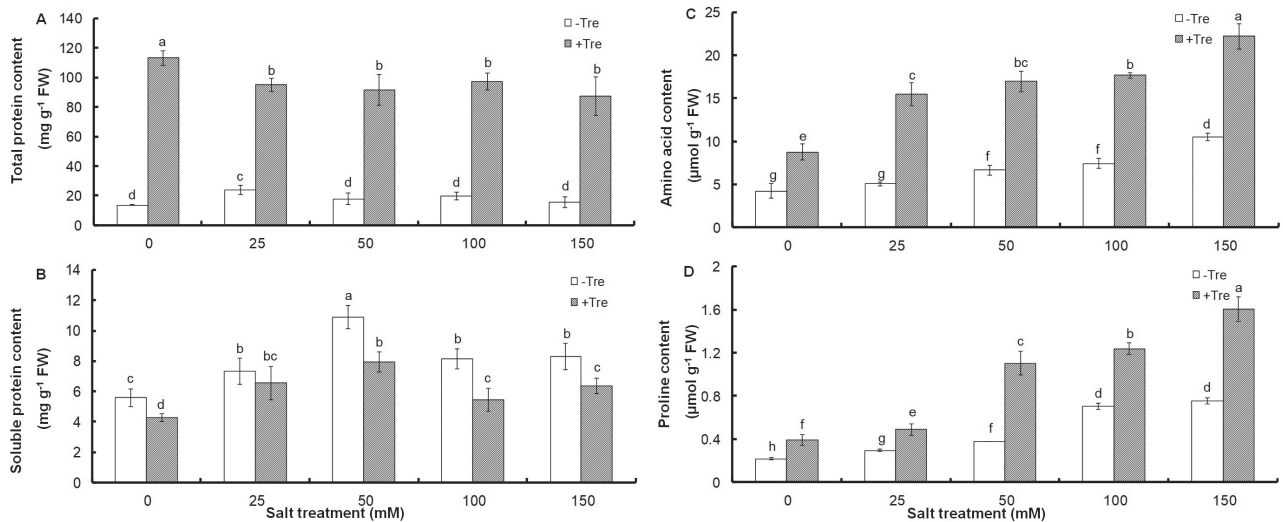
### Effect of salinity and Tre on total proteins, total amino acid, and proline contents

The positive effect of salinity on total protein content was observed only at 25 mM salt concentration with a 75% increase and no significant change was found in other salt treatments, while all salinity levels significantly increased soluble proteins, especially up to 10.88 mg g<sup>-1</sup> FW under 50 mM NaCl (Fig. 3A, 3B).

Salinity also caused the accumulation of amino acids and proline so that this increase was proportional to the salt concentration and reached 151 and 247% of control for amino acids and proline in 150 mM NaCl concentration, respectively (Fig. 3C, 3D). Tre treatment at all salinity concentrations caused a strong increase in total proteins, amino acids, and proline contents, which was associated with a decrease in the concentration of soluble proteins (Fig. 3).



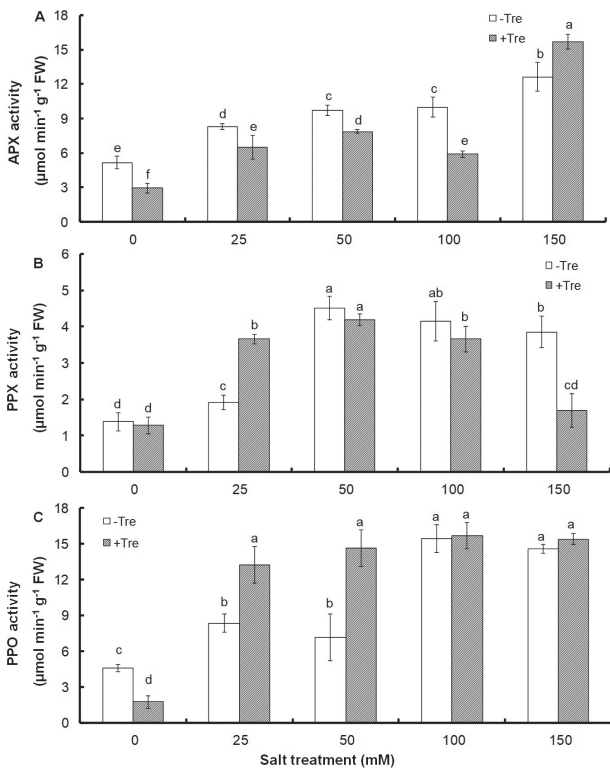
**Fig. 2.** Effect of different salinity concentrations on content of reducing sugars, RS (A), non-reducing sugars, NRS (B), and total soluble sugars, TSS (C), as well as on reducing / non-reducing sugars, RS/NRS ratio (D), and starch content (E) in trehalose-treated (+Tre) or untreated (-Tre) basil seedlings. Results are the mean  $\pm$  standard deviation of three separate measurements. Different letters indicate significant differences between the various treatments at  $P < 0.05$  according to the Duncan test.



**Fig. 3.** Effect of different salinity concentrations on contents of total proteins (A), total soluble proteins (B), total amino acids (C), and proline (D) in trehalose-treated (+Tre) or untreated (-Tre) basil seedlings. Results are the mean ± standard deviation of three separate measurements. Different letters indicate significant differences between the various treatments at  $P < 0.05$  according to the Duncan test.

**Effect of salinity and Tre on enzyme activities**

The activity of antioxidant enzymes including APX, PPX, and PPO increased strongly with salt concentration and reached 144, 179, and 218% at 150 mM NaCl compared to the control (Fig. 4).



**Fig. 4.** Effect of different salinity concentrations on the activity of ascorbate peroxidase, APX (A), pyrogallol peroxidase, PPX (B), and polyphenol oxidase, PPO (C) in trehalose-treated (+Tre) or untreated (-Tre) basil seedlings. Results are the mean ± standard deviation of three separate measurements. Different letters indicate significant differences between the various treatments at  $P < 0.05$  according to the Duncan test.

Tre treatment alone or with concentrations of up to 100 mM salt strongly decreased APX activity but increased its activity at 150 mM salt concentration by 24% compared to untreated control (Fig. 4A). PPX enzyme activity did not change in plants treated with Tre alone, but it was strongly increased at 25 mM salinity by 92%, slightly decreased at higher salt concentrations, and significantly decreased at 150 mM salinity by 56% compared to untreated control (Fig. 4B). The activity of PPO enzyme showed a 62% decrease in Tre-treated seedlings compared to untreated ones under non-stress conditions, but it increased significantly by 58 and 104% at concentrations of 25 and 50 mM salt, respectively, and remained unchanged at higher salinities (Fig. 4C).

**Discussion**

Much research has documented the negative effect of salinity on plant growth and biomass (Ahmad and Jhon 2005, Yoon et al. 2009, Qiu et al. 2014, Ahmad et al. 2018, Scagel et al. 2019, Sheyhakinia et al. 2020). Despite the reduction of some growth indices in basil seedlings under saline conditions, root length, leaf characteristics, and shoot water content did not change and root dry weight increased. Considering the lack of change in the length of the root and also the decrease in the ratio of fresh weight to dry weight of the root under salinity, which is due to a decrease in fresh weight and increase of dry weight, the increase of root biomass can be caused by the accumulation of nutrients in the organ during stress. In fact, salinity causes a change in the pattern of carbon allocation between roots and shoots. This change in the pattern of carbon distribution during salinity is associated with the reduction of shoot biomass. An increase in root biomass and a decrease in shoot biomass under salt stress have been reported (Bernstein and Kafkafi 2002, Imada et al. 2015). Since this process is similar to the

responses of roots and shoots to soil water availability (Zhang et al. 2005, McCarthy and Enquist 2007, Imada et al. 2008), one of the reasons is probably the reduction of soil moisture absorption (McCarthy and Enquist 2007). In fact, the inhibition of growth induced by salinity may be related to the reduction of water absorption due to the reduction of soil osmotic potential. Such a phenomenon leads to cell dehydration and loss of turgor pressure, which leads to growth arrest (Zhao et al. 2021). In addition, salinity leads to the accumulation of Na<sup>+</sup> and Cl<sup>-</sup> ions, which reduces the absorption and transport of nutrients due to competitive interactions with their transporters (Zhao et al. 2021, Gao et al. 2022). However, in our results, salinity-induced growth reduction was observed without a change in shoot water content, indicating that this reduction is associated with continuous water uptake and maintenance of shoot turgor pressure, as previously documented (De Costa et al. 2007, Alisofi et al. 2020, Sheyhakinia et al. 2020). Accumulation of proline in basil seedlings under salinity can explain the uninterrupted water uptake and thus the maintenance of shoot water content despite growth limitation. Therefore, changes in carbon partitioning and accumulation of osmolytes to continue water absorption by plants can be the main reason for growth inhibition under saline conditions. The results of studies on *Hibiscus sabdariffa* (Sheyhakinia et al. 2020) and *Momordica charantia* (Alisofi et al. 2020) under salt stress are in good agreement with our findings.

Contrary to previous studies on the effect of Tre on plant growth under salinity stress (Mostofa et al. 2015, Abdallah et al. 2020), Tre treatment in most salinities decreased shoot length and fresh and dry weight of shoots and roots in basil seedlings, so that the dry weight reduction was more severe. Early experiments on the effect of Tre on higher plants through inhibition of trehalase (a Tre-degrading enzyme) in species with very low trehalase activity showed that Tre accumulation is toxic or at least Tre acts as a plant growth inhibitor possibly through inhibition of cell wall biosynthesis (Veluthambi et al. 1981, Veluthambi et al. 1982a). This effect is associated with a disturbance in carbohydrate metabolism as indicated by a decrease in sucrose content (Veluthambi et al. 1982b). Therefore, the reduction of growth in basil seedlings due to Tre treatment can be related to the low activity of this enzyme. Meanwhile, in plant species with high trehalase enzyme activity, such as *Raphanus sativus*, *Quamoclit phoenicea*, and *Zea mays*, toxicity and inhibition of growth by Tre were not observed (Veluthambi et al. 1981). In transgenic *Arabidopsis* seedlings in which trehalase enzyme activity was overexpressed, there was no effect of growth inhibition in the presence of Tre, unlike the control (Schluepmann et al. 2004). Therefore, the decrease of growth induced by Tre can be due to the plant's inability to metabolize it, which leads to the accumulation of trehalose-6-phosphate and subsequently to the reduction of the glucose-6-phosphate pool (Schluepmann et al. 2004). In addition, it has been found that trehalose-6-phosphate induces plant defense responses, which are associated with growth reduction (Reignault et al. 2001, Brodmann et al.

2002, Renard-Merlier et al. 2007). However, Tre did not affect SWC in basil seedlings up to 100 mM NaCl, but increased it in seedlings exposed to 150 mM NaCl. This event is associated with proline hyperaccumulation, so it can be concluded that changes in carbon partitioning and directing it to proline biosynthesis are the reason for Tre-induced growth inhibition under saline conditions. Considering the role of Tre and its metabolism in the regulation of growth and development (Nunes et al. 2013, O'Hara et al. 2013), it is likely that this metabolite is involved in the growth regulation processes of basil seedlings. In this regard, it has recently been found that methyl jasmonate treatment not only does not reduce the harmful effects of salinity stress in radish (*Raphanus sativus*) but can even inhibit plant growth (Henschel et al. 2023). This shows that signal molecules and osmolytes such as Tre, which usually increase tolerance to environmental stresses, do not work in the same way in all plants and their performance against different stresses can depend on the type of plant species.

It has been found that the concentration of photosynthetic pigments is sensitive to environmental stresses and in most plants, it decreases significantly in response to salinity (Ahmad and Jhon 2005, Gunes et al. 2007, Yoon et al. 2009, Qiu et al. 2014). However, a study on tomato has shown that 0.3 M salt concentration induces Chl production per unit of leaf area (Agong et al. 2004). This shows that very low salinity induces Chl biosynthesis in basil, as observed in the concentration of Chl *a* and total Chl at 25 mM NaCl. In addition, the observed decrease in Chl *a/b* and Chl / Car ratios in response to salinity shows the greater sensitivity of Chl *a* compare to Chl *b* and Chl to Car. Contrary to current studies on the role of Tre in increasing the photosynthetic pigments of different plants under salinity (Theerakulpisut and Phongngarm 2013, Abdallah et al. 2016, 2020), treatment of basil seedlings with Tre in most salinities showed a significant decrease in pigment content compared to untreated plants. The increase in the ratio of Chl *a/b* in Tre-treated seedlings under non-stress conditions or not changing it at the concentration of 25 mM indicates the greater sensitivity of Chl *b* to Tre. It has been found that the biosynthesis of Chl *b* is carried out through the oxidation of a methyl group to a formyl group on the B ring of the Chl *a* molecule (Porra et al. 1994). Therefore, it can be assumed that the enzyme(s) involved in Chl *a* biosynthesis are more active than Chl *b* due to Tre treatment alone or in combination with low salinity. However, the sharp decrease of this ratio in salt concentrations of 50 mM and higher shows the greater sensitivity of Chl *a* to Tre. The change pattern of the Chl to Car ratio also shows the lower sensitivity of Car to Tre in non-stressed conditions or in combination with low salinity and its greater sensitivity in higher salinity concentrations. Considering the negative effect of Tre on the growth of basil seedlings under salt stress, the reduction of photosynthetic pigments can be attributed to the inability of plants to properly metabolize Tre or direct carbon to other metabolic pathways.

Accumulation of sugars or osmotic regulators in plant cells under salinity is a strategy to control plant water content and inhibit water loss (Chaves et al. 2009). The role of the accumulation of soluble sugars in salinity stress tolerance has been widely studied (Mishra et al. 2008, Yin et al. 2010). This indicates the type of basil strategy in salinity tolerance that is done through the accumulation of soluble sugars. Considering the sharp decrease in the ratio of RS/NRS under salt stress compared to the control, it can be concluded that the accumulation of NRS occurs at a higher rate than RS during salt treatment. In addition, the numerical comparison of the amount of these sugars in response to different salt concentrations shows that the accumulation of NRS is almost twice as much as RS. This indicates the important role of non-reducing sugars in basil seedlings for salinity tolerance. On the other hand, the increase of soluble sugars during salinity stress is associated with the decrease in starch content. This suggests that a change in photosynthetic carbon partitioning occurs in basil seedlings during salinity stress, leading to more sucrose synthesis and less starch accumulation. The higher accumulation of NRS than RS confirms this. However, starch breakdown can also increase soluble sugars during salt stress. The conversion of starch to sugars, especially NRS, and a change in their metabolism, which has been introduced as a common defense strategy against water stress, is possible through increasing the activity of starch hydrolyzing enzymes such as amylases and simultaneous decrease of sucrose hydrolyzing activities (Kumari and Asthir 2016). The unremarkable increase of NRS content in Tre-treated seedlings under salinity compared to the untreated controls indicates that this metabolite does not accumulate in basil seedlings. This finding can confirm the hypothesis that Tre accumulates in some Tre-treated plants in another form such as trehalose-6-phosphate (Schluepmann et al. 2004). This proves that the effects of Tre on growth are due to changes in carbohydrate metabolism. Evidence to support this hypothesis comes from the observation that the simultaneous addition of sucrose in the presence of Tre and starch accumulation restores growth in response to Tre feeding (Wingler et al. 2000). Tre feeding leads to an increase in ADP-glucose pyrophosphorylase gene expression as well as an increase in its enzyme activity, which is crucial in starch biosynthesis (Wingler et al. 2000). In addition, trehalose-6-phosphate also plays a role in the activation of this enzyme (Kolbe et al. 2005). Various studies have shown that growth was impaired after an increase of the gene expression of this enzyme in potato plants, but with the addition of sucrose, growth returned to the normal state (Stark et al. 1992). Accordingly, one of the reasons for Tre-induced growth inhibition during salt treatment can be due to the reduction of carbon for export to growth areas because of starch accumulation.

Environmental stresses, including salinity, generally cause protein degradation or reduced synthesis due to the acceleration of the aging process (Mishra et al. 2008, Misra and Saxena 2009). The results of our study on basil seedlings

showed the opposite of this finding, which could indicate an increase in the synthesis of stress-specific proteins or enzyme proteins during the salinity period. The observed decrease in soluble protein levels, which was accompanied by a significant increase in total protein content in Tre-treated seedlings, could be attributed to a change in their solubility due to Tre treatment. This suggestion is supported by the fact that the level of soluble protein is positively correlated with protein solubility (Afify et al. 2012, Ma et al. 2019, Ebert et al. 2020). Therefore, the increase in total protein while the soluble protein decreased can be due to the role of Tre in reducing the solubility of proteins. The response of basil seedlings to Tre treatment can confirm that the pathway of carbon partitioning and its metabolism is changed due to Tre treatment, which in turn reduces growth in Tre-treated seedlings under salinity conditions. Similar results regarding the effects of Tre on salinity tolerance and its role in carbon partitioning have also been reported in *Dunaliella bardawil* (Panjekobi and Einali 2021).

Accumulation of amino acids and proline is considered a common response during environmental stress, and is often associated with the improvement of plant tolerance to stress conditions (Claussen 2005, Khadri et al. 2006, Yoon et al. 2009). However, some studies have shown that the level of accumulated proline indicates the severity of stress symptoms when plants are exposed to different types of abiotic stresses (Metwally et al. 2003, Mostofa et al. 2014, 2015). A study on *Catharanthus roseus* under salt stress showed that proline accumulation under stress conditions is negatively correlated with relative water content, biomass, and potassium accumulation (Chang et al. 2014). However, the lack of change in SWC in saline conditions or its increase due to Tre treatment and high salt dose, which is associated with proline accumulation, can indicate the increase of this metabolite in order to maintain the water content of the plant. Therefore, in our study, the excessive accumulation of amino acids and proline that occurred especially due to Tre treatment is related to the maintenance of water content, which is caused by Tre-induced metabolic changes during salt stress. In this way, the increase of these metabolites in seedlings treated with Tre alone also indicates a kind of stressful condition. On the contrary, studies on plants treated with Tre under salt (Nounjan et al. 2012, Mostofa et al. 2014, Sadak, 2019) and drought stress (Ali and Ashraf 2011) show a lower level of proline or amino acid in these plants, which could indicate a lower demand for proline or a compensatory mechanism for Tre, because both can act as an osmoprotectant.

The increased activity of antioxidant enzymes in response to salinity can be considered as an indication of increased ROS production and a common protective mechanism to reduce oxidative damage caused by salinity. These results are consistent with other studies on the role of ROS-scavenging enzymes under salt stress (Ahmad et al. 2018, Alisofi et al. 2020, Sheyhakinia et al. 2020). Different activities of antioxidant enzymes in response to Tre under salt

stress have been reported in various plants. Tre treatment decreased the activity of peroxidase, catalase, and superoxide dismutase enzymes in rice under salt stress (Rohman et al. 2019). In another study, the treatment of *Chenopodium quinoa* with Tre under salt stress increased the activity of APX, catalase and superoxide dismutase enzymes (Abdallah et al. 2020). In addition, an increase in the activity of antioxidant enzymes has been observed with Tre treatment under different stress conditions (Zhao et al. 2019, Liu et al. 2020). However, in all these studies, Tre-treated plants showed tolerance to salt stress, which can indicate the existence of different strategies in plants to cope with salinity in the presence of Tre. In contrast, our results showed that the changes in enzyme activity patterns induced by Tre treatment do not indicate increased tolerance to salt stress, because these patterns are not correlated with other indicators showing stress reduction.

## Conclusion

The results of this study show that salt stress without change in SWC decreases growth characteristics, photosynthetic pigments, and starch content, but increases the activity of ROS-scavenging enzymes and the accumulation of metabolites including soluble sugars, proteins, free amino acids, and proline. Tre treatment not only does not reduce the adverse effects of salinity but even causes more severe inhibition of plant growth, further reduction of photosynthetic pigments and soluble proteins along with the excessive accumulation of free amino acids and proline. Therefore, Tre treatment is not effective in salinity tolerance of basil seedlings and reduces their growth possibly through diverting carbon to other metabolic pathways rather than growth processes. However, due to the effect of Tre on the change in carbon partitioning, the role of this molecule in various metabolic and physiological pathways is obvious, the detailed understanding of which requires further research on the physiological effects of Tre under stress and non-stress conditions.

## Acknowledgments

We thank the Deputy of Research at the University of Sistan and Baluchestan for financial support in the form of grants for the MSc research project.

This work was funded by the Deputy of Research at the University of Sistan and Baluchestan in the form of a grant for M.Sc. research project.

## Author contribution statement

R. Karamzahi carried out all the laboratory research. A. Einali designed the experiment, provided all the technical support during the laboratory work, analyzed data and wrote the manuscript. All authors have read and approved the submitted manuscript.

## References

- Abdallah, M. M. S., El Sebai, T. N., Ramadan, A. A. E. M., El-Bassiouny, H. M. S., 2020: Physiological and biochemical role of proline, trehalose, and compost on enhancing salinity tolerance of quinoa plant. *Bulletin of the National Research Centre* 44, 96. <https://doi.org/10.1186/s42269-020-00354-4>
- Abdallah, M. S., Abdelgawad, Z. A., El-Bassiouny, H. M. S., 2016: Alleviation of the adverse effects of salinity stress using trehalose in two rice varieties. *South African Journal of Botany* 103, 275–282. <https://doi.org/10.1016/j.sajb.2015.09.019>
- Afify, A. E. M. M., El-Beltagi, H. S., Abd El-Salam, S. M., Omran, A. A., 2012: Protein solubility, digestibility and fractionation after germination of sorghum varieties. *Plos one* 7, e31154. <https://doi.org/10.1371/journal.pone.0031154>
- Agong, S. G., Yoshida, Y., Yazawa, S., Masuda, M., 2004: Tomato response to salt stress. In: XXVI International Horticultural Congress: Advances in Vegetable Breeding, International Society for Horticultural Science. *Acta Horticulturae* 637, 93–97. <https://doi.org/10.17660/ActaHortic.2004.637.10>
- Ahamed, G. J., Li, Y., Li, X., Han, W. Y., Chen, S., 2018: Epigallocatechin-3-gallate alleviates salinity-retarded seed germination and oxidative stress in tomato. *Journal of Plant Growth Regulation* 37, 1349–1356. <https://doi.org/10.1007/s00344-018-9849-0>
- Ahmad, P., Alyemeni, M. N., Abass Ahanger, M., Wijaya, L., Alam, P., Kumar, A., Ashraf, M., 2018: Upregulation of antioxidant and glyoxalase systems mitigates NaCl stress in *Brassica juncea* by supplementation of zinc and calcium. *Journal of Plant Interactions* 13(1), 151–162. <https://doi.org/10.1080/17429145.2018.1441452>
- Ahmad, P., Jhon, R., 2005: Effect of salt stress on growth and biochemical parameters of *Pisum sativum* L. *Archives of Agronomy and Soil Science* 51(6), 665–672. <https://doi.org/10.1080/03650340500274151>
- Ali, M., Kamran, M., Abbasi, G. H., Saleem, M. H., Ahmad, S., Parveen, A., Fahad, S., 2021: Melatonin-induced salinity tolerance by ameliorating osmotic and oxidative stress in the seedlings of two tomato (*Solanum lycopersicum* L.) cultivars. *Journal of Plant Growth Regulation* 40, 2236–2248. <https://doi.org/10.1007/s00344-020-10273-3>
- Ali, Q., Ashraf, M., 2011: Induction of drought tolerance in maize (*Zea mays* L.) due to exogenous application of trehalose: growth, photosynthesis, water relations and oxidative defence mechanism. *Journal of Agronomy and Crop Science* 197(4), 258–271. <https://doi.org/10.1111/j.1439-037X.2010.00463.x>
- Alisofi, S., Einali, A., Sangtarash, M. H., 2020: Jasmonic acid-induced metabolic responses in bitter melon (*Momordica charantia*) seedlings under salt stress. *The Journal of Horticultural Science and Biotechnology* 95(2), 247–259. <https://doi.org/10.1080/14620316.2019.1663135>
- Arnon, D., 1949: Copper enzymes in isolated chloroplasts: polyphenoloxidase in *Beta vulgaris*. *Plant Physiology* 24(1), 1–15. <https://doi.org/10.1104/pp.24.1.1>
- Bahcesular, B., Yildirim, E.D., Karaçocuk, M., Kulak, M., Karaman, S., 2020: Seed priming with melatonin effects on growth, essential oil compounds and antioxidant activity of basil (*Ocimum basilicum* L.) under salinity stress. *Industrial Crops and Products* 146, 112165. <https://doi.org/10.1016/j.indcrop.2020.112165>
- Bates, L. S., Waldren, R. P., Teare, I. D., 1973: Rapid determination of free Proline for water stress studies. *Plant and Soil* 39, 205–207. <https://doi.org/10.1007/BF00018060>
- Bernstein, N., Kafkafi, U., 2002: Root growth under salinity stress. In: *Plant Roots* (Eds. Waisel, Y., Eshel, A., Beekman, T., Kafkafi, U.), 1222–1250. CRC Press.

- Bradford, M. M., 1976: A rapid and sensitive method for the quantitation of microgram quantities utilizing the principle of protein dye binding. *Analytical Biochemistry* 72(1–2), 248–254. [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- Brodmann, D., Schuller, A., Ludwig-Müller, J., Aeschbacher, R.A., Wiemken, A., Boller, T., Winkler, A., 2002: Induction of trehalase in *Arabidopsis* plants infected with the trehalose producing pathogen *Plasmodiophora brassicae*. *Molecular Plant-Microbe Interactions* 15(7), 693–700. <https://doi.org/10.1094/MPMI.2002.15.7.693>
- Caliskan, O., Kurt, D., Temizel, K. E., Odabas, M. S., 2017: Effect of salt stress and irrigation water on growth and development of sweet basil (*Ocimum basilicum* L.). *Open Agriculture* 2(1), 589–594. <https://doi.org/10.1515/opag-2017-0062>
- Chang, B., Yang, L., Cong, W., Zu, Y., Tang, Z., 2014: The improved resistance to high salinity induced by trehalose is associated with ionic regulation and osmotic adjustment in *Catharanthus roseus*. *Plant Physiology and Biochemistry* 77, 140–148. <https://doi.org/10.1016/j.plaphy.2014.02.001>
- Chaves, M. M., Flexas, J., Pinheiro, C., 2009: Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of botany* 103(4), 551–560. <https://doi.org/10.1093/aob/mcn125>
- Chen, G. X., Asada, K., 1992: Inactivation of ascorbate peroxidase by thiols requires hydrogen peroxide. *Plant and cell physiology* 33(2), 117–123. <https://doi.org/10.1093/oxfordjournals.pcp.a078229>
- Chen, H., Jiang, J. G., 2010: Osmotic adjustment and plant adaptation to environmental changes related to drought and salinity. *Environmental Reviews* 18, 309–319. <https://doi.org/10.1139/A10-014>
- Claussen, W., 2005: Proline as a measure of stress in tomato plants. *Plant Science* 168, 241–248. <https://doi.org/10.1016/j.plantsci.2004.07.039>
- Colin, L., Ruhnnow, F., Zhu, J. K., Zhao, C., Zhao, Y., Persson, S., 2023: The cell biology of primary cell walls during salt stress. *Plant Cell* 35(1), 201–217. <https://doi.org/10.1093/plcell/koac292>
- De Costa, W., Zörb, C., Hartung, W., Schubert, S., 2007: Salt resistance is determined by osmotic adjustment and abscisic acid in newly developed maize hybrids in the first phase of salt stress. *Physiologia Plantarum* 131(2), 311–321. <https://doi.org/10.1111/j.1399-3054.2007.00962.x>
- Ebert, S., Gibis, M., Terjung, N., Weiss, J., 2020: Survey of aqueous solubility, appearance, and pH of plant protein powders from carbohydrate and vegetable oil production. *LWT- Food Science and Technology* 133, 110078. <https://doi.org/10.1016/j.lwt.2020.110078>
- Einali, A., Valizadeh, J., 2017: Storage reserve mobilization, gluconeogenesis, and oxidative pattern in dormant pistachio (*Pistacia vera* L.) seeds during cold stratification. *Trees* 31, 659–671. <https://doi.org/10.1007/s00468-016-1498-y>
- Farouk, S., Omar, M.M., 2020: Sweet basil growth, physiological and ultrastructural modification, and oxidative defense system under water deficit and silicon forms treatment. *Journal of Plant Growth Regulation* 39, 1307–1331. <https://doi.org/10.1007/s00344-020-10071-x>
- Gao, Z., Zhang, J., Zhang, J., Zhang, W., Zheng, L., Borjigin, T., Wang, Y., 2022: Nitric oxide alleviates salt-induced stress damage by regulating the ascorbate–glutathione cycle and Na<sup>+</sup>/K<sup>+</sup> homeostasis in *Nitraria tangutorum* Bobr. *Plant Physiology and Biochemistry* 173, 46–58. <https://doi.org/10.1016/j.plaphy.2022.01.017>
- Gunes, A., Anal, A., Alpaslan, M., Eraslan, F., Bagci, E. G., Cick, N., 2007: Salicylic acid induced changes on some physiological parameters symptomatic for oxidative stress and mineral nutrition in maize (*Zea mays* L.) grown under salinity. *Journal of Plant Physiology* 164(6), 728–736. <https://doi.org/10.1016/j.jplph.2005.12.009>
- Handel, E.V., 1968: Direct microdetermination of sucrose. *Analytical Biochemistry* 22(2), 280–283. [https://doi.org/10.1016/0003-2697\(68\)90317-5](https://doi.org/10.1016/0003-2697(68)90317-5)
- He, M., He, C.Q., Ding, N.Z., 2018: Abiotic stresses: general defenses of land plants and chances for engineering multistress tolerance. *Frontiers in Plant Science* 9, 1771. <https://doi.org/10.3389/fpls.2018.01771>
- Henschel, J. M., de Moura, V. S., Silva, A. M. O., Gomes, D. D. S., dos Santos, S. K., Batista, D. S., Dias, T. J., 2023: Can exogenous methyl jasmonate mitigate salt stress in radish plants? *Theoretical and Experimental Plant Physiology* 35, 51–63. <https://doi.org/10.1007/s40626-023-00270-8>
- Imada, S., Matsuo, N., Acharya, K., Yamanaka, N., 2015: Effects of salinity on fine root distribution and whole plant biomass of *Tamarix ramosissima* cuttings. *Journal of Arid Environments* 114, 84–90. <https://doi.org/10.1016/j.jaridenv.2014.11.011>
- Imada, S., Yamanaka, N., Tamai, S., 2008: Water table depth affects *Populus alba* fine root growth and whole plant biomass. *Functional Ecology* 22(6), 1018–1026. <https://doi.org/10.1111/j.1365-2435.2008.01454.x>
- Islam, S., Mohammad, F., 2021: Modulation of growth, photosynthetic efficiency, leaf biochemistry, cell viability and yield of Indian mustard by the application of trehalose. *Scientia Horticulturae* 290, 110527. <https://doi.org/10.1016/j.scienta.2021.110527>
- Islam, S., Mohammad, F., Siddiqui, M. H., Kalaji, H. M., 2023: Salicylic acid and trehalose attenuate salt toxicity in *Brassica juncea* L. by activating the stress defense mechanism. *Environmental Pollution* 326, 121467. <https://doi.org/10.1016/j.envpol.2023.121467>
- Kaur, H., Hussain, S.J., Kaur, G., Poor, P., Alamri, S., Siddiqui, M.H., Khan, M.I.R., 2022: Salicylic Acid Improves Nitrogen Fixation, Growth, Yield and Antioxidant Defence Mechanisms in Chickpea Genotypes Under Salt Stress. *Journal of Plant Growth Regulation* 41, 2034–2047. <https://doi.org/10.1007/s00344-022-10592-7>
- Khadri, M., Tejera, N.A., Lluch, C., 2006: Alleviation of salt stress in common bean by exogenous abscisic acid supply. *Journal of Plant Growth Regulation* 25, 110–119. <https://doi.org/10.1007/s00344-005-0004-3>
- Khan, T.A., Saleem, M., Fariduddin, Q., 2023: Melatonin influences stomatal behavior, root morphology, cell viability, photosynthetic responses, fruit yield, and fruit quality of tomato plants exposed to salt stress. *Journal of Plant Growth Regulation* 42, 2408–2432. <https://doi.org/10.1007/s00344-022-10713-2>
- Kolbe, A., Tiessen, A., Schluepmann, H., Paul, M., Ulrich, S., Geigenberger, P., 2005: Trehalose 6-phosphate regulates starch synthesis via posttranslational redox activation of ADP-glucose pyrophosphorylase. *Proceedings of the National Academy of Sciences* 102(31), 11118–11123. <https://doi.org/10.1073/pnas.0503410102>
- Kosar, F., Akram, N.A., Sadiq, M., Al-Qurainy, F., Ashraf, M., 2019: Trehalose: a key organic osmolyte effectively involved in plant abiotic stress tolerance. *Journal of Plant Growth Regulation* 38, 606–618. <https://doi.org/10.1007/s00344-018-9876-x>
- Kumari, M., Asthir, B., 2016: Transformation of sucrose to starch and protein in rice leaves and grains under two establishment methods. *Rice Science* 23(5), 255–265. <https://doi.org/10.1016/j.rsci.2016.08.003>



- Kwee, E.M., Niemeyer, E.D., 2011: Variations in phenolic composition and antioxidant properties among 15 basil (*Ocimum basilicum* L.) cultivars. *Food Chemistry* 128(4), 1044–1050. <https://doi.org/10.1016/j.foodchem.2011.04.011>
- Lichtenthaler, H.K., Buschmann, C., 2001: Chlorophylls and carotenoids: measurement and characterization by UV-VIS spectroscopy. *Current protocols in food analytical chemistry* F4.3.1–F4.3.8. <https://doi.org/10.1002/0471142913.faf0403s01>
- Liu, T., Ye, X., Li, M., Li, J., Qi, H., Hu, X., 2020: H<sub>2</sub>O<sub>2</sub> and NO are involved in trehalose-regulated oxidative stress tolerance in cold-stressed tomato plants. *Environmental and Experimental Botany* 171, 103961. <https://doi.org/10.1016/j.envexpbot.2019.103961>
- Lunn, J.E., Delorge, I., Figueroa, C.M., Van Dijck, P., Stitt, M., 2014: Trehalose metabolism in plants. *The Plant Journal* 79(4), 544–567. <https://doi.org/10.1111/tj.12509>
- Luo, Y., Li, F., Wang, G.P., Yang, X.H., Wang, W., 2010: Exogenous supplied trehalose protects thylakoid membranes of winter wheat from heat-induced damage. *Biologia Plantarum* 54, 495–501. <https://doi.org/10.1007/s10535-010-0087-y>
- Ma, J., Zhu, X., Shi, L., Ni, C., Hou, J., Cheng, J., 2019: Enhancement of soluble protein, polypeptide production and functional properties of heat-denatured soybean meal by fermentation of *Monascus purpureus* 04093. *CyTA-Journal of Food* 17(1), 1014–1022. <https://doi.org/10.1080/19476337.2019.1695677>
- Markwell, M.A.K., Hass, S.M., Tolbert N.E., Bieber, L.L., 1981: Protein determination in membrane and lipoprotein samples: manual and automated procedures. *Methods in Enzymology* 72, 296–303. [https://doi.org/10.1016/s0076-6879\(81\)72018-4](https://doi.org/10.1016/s0076-6879(81)72018-4)
- McCarthy, M. C., Enquist, B. J., 2007: Consistency between an allometric approach and optimal partitioning theory in global patterns of plant biomass allocation. *Functional Ecology* 21(4), 713–720. <https://doi.org/10.1111/j.1365-2435.2007.01276.x>
- McCready, R.M., Guggolz, J., Silviera, V., Owens, H.S., 1950: Determination of starch and amylose in vegetables. *Analytical Chemistry* 22, 1156–1158. <https://doi.org/10.1021/ac60045a016>
- Metwally, A., Finkemeier, I., Georgi, M., Dietz, K. J., 2003: Salicylic acid alleviates the cadmium toxicity in barley seedlings. *Plant physiology* 132(1), 272–281. <https://doi.org/10.1104/pp.102.018457>
- Miller, G.L., 1959: Use of dinitrosalicylic acid reagent for determination of reducing sugars. *Analytical Chemistry* 31, 426–428. <https://doi.org/10.1021/ac60147a030>
- Mishra, A., Mandoli, A., Jha, B., 2008: Physiological characterization and stress-induced metabolic responses of *Dunaliella salina* isolated from salt pan. *Journal of Industrial Microbiology and Biotechnology* 35(10), 1093–1101. <https://doi.org/10.1007/s10295-008-0387-9>
- Misra, N., Saxena, P., 2009: Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. *Plant Science* 177(3), 181–189. <https://doi.org/10.1016/j.plantsci.2009.05.007>
- Mostofa, M.G., Hossain, M.A., Fujita, M., 2015: Trehalose pretreatment induces salt tolerance in rice (*Oryza sativa* L.) seedlings: oxidative damage and co-induction of antioxidant defense and glyoxalase systems. *Protoplasma* 252, 461–475. <https://doi.org/10.1007/s00709-014-0691-3>
- Mostofa, M.G., Seraj, Z.I., Fujita, M., 2014: Exogenous sodium nitroprusside and glutathione alleviate copper toxicity by reducing copper uptake and oxidative damage in rice (*Oryza sativa* L.) seedlings. *Protoplasma* 251, 1373–1386. <https://doi.org/10.1007/s00709-014-0639-7>
- Muller, J., Boller, T., Wiemken, A., 1998: Trehalose affects sucrose synthase and invertase activities in soybean (*Glycine max* [L.] Merr.) roots. *Journal of Plant Physiology* 153(1–2), 255–257. [https://doi.org/10.1016/S0176-1617\(98\)80078-3](https://doi.org/10.1016/S0176-1617(98)80078-3)
- Muller, J., Wiemken, A., Aeschbacher, R., 1999: Trehalose metabolism in sugar sensing and plant development. *Plant Science* 147(1), 37–47. [https://doi.org/10.1016/S0168-9452\(99\)00094-1](https://doi.org/10.1016/S0168-9452(99)00094-1)
- Nakano, Y., Asada, K., 1981: Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. *Plant and cell physiology* 22(5), 867–880. <https://doi.org/10.1093/oxfordjournals.pcp.a076232>
- Nakashima, K., Ito, Y., Yamaguchi-Shinozaki, K., 2009: Transcriptional regulatory networks in response to abiotic stresses in *Arabidopsis* and grasses. *Plant Physiology* 149(1), 88–95. <https://doi.org/10.1104/pp.108.129791>
- Nakayama, H., Horie, T., Yonamine, I., Shinmyo, A., Yoshida, K., 2005: Improving salt tolerance in plant cells. *Plant Biotechnology* 22(5), 477–487. <https://doi.org/10.5511/plantbiotechnology.22.477>
- Nounjan, N., Nghia, P.T., Theerakulpisut, P., 2012: Exogenous proline and trehalose promote recovery of rice seedlings from salt-stress and differentially modulate antioxidant enzymes and expression of related genes. *Journal of Plant Physiology* 169(6), 596–604. <https://doi.org/10.1016/j.jplph.2012.01.004>
- Nunes, C., Schluepmann, H., Delatte, T.L., Wingler, A., Silva, A.B., Fevereiro, P.S., Jansen, M., Fiorani, F., Wiese-Klinkenberg, A., Paul, M.J., 2013: Regulation of growth by the trehalose pathway: relationship to temperature and sucrose. *Plant signaling & behavior* 8(12), e26626. <https://doi.org/10.4161/psb.26626>
- O'Hara, L. E., Paul, M. J., Wingler, A., 2013: How do sugars regulate plant growth and development? New insight into the role of trehalose-6-phosphate. *Molecular Plant* 6(2), 261–274. <https://doi.org/10.1093/mp/sss120>
- Panjekobi, M., Einali, A., 2021: Trehalose treatment alters carbon partitioning and reduces the accumulation of individual metabolites but does not affect salt tolerance in the green microalga *Dunaliella bardawil*. *Physiology and Molecular Biology of Plants* 27, 2333–2344. <https://doi.org/10.1007/s12298-021-01078-z>
- Peng, X., Wang, N., Sun, S., Geng, L., Guo, N., Liu, A., Chen, S., Ahammed, G.J., 2023: Reactive oxygen species signaling is involved in melatonin-induced reduction of chlorothalonil residue in tomato leaves. *Journal of Hazardous Materials* 443(A), 130212. <https://doi.org/10.1016/j.jhazmat.2022.130212>
- Porra, R.J., Schafer, W., Cmiel, E., Katheder, I., Scheer, H., 1994: The derivation of the formyl-group oxygen of chlorophyll b in higher plants from molecular oxygen. Achievement of high enrichment of the 7-formyl-group oxygen from 18O<sub>2</sub> in greening maize leaves. *European Journal of Biochemistry* 219(1–2), 671–679. <https://doi.org/10.1111/j.1432-1033.1994.tb19983.x>
- Qiu, Z., Guo, J., Zhu, A., Zhang, L., Zhang, M., 2014: Exogenous jasmonic acid can enhance tolerance of wheat seedlings to salt stress. *Ecotoxicology and Environmental Safety* 104, 202–208. <https://doi.org/10.1016/j.ecoenv.2014.03.014>
- Reignault, P., Cogan, A., Muchembled, J., Sahraoui, A.L.H., Durand, R., Sancholle, M., 2001: Trehalose induces resistance to powdery mildew in wheat. *New Phytologist* 149(3), 519–529. <https://doi.org/10.1046/j.1469-8137.2001.00035.x>
- Renard-Merlier, D., Randoux, B., Nowak, E., Farcy, F., Durnad, R., Reignault, P., 2007: Iodine 40, salicylic acid, heptanoyl salicylic acid and trehalose exhibit different efficacies and defense targets during wheat/powdery mildew infection. *Phytochemistry* 68(8), 1156–1164. <https://doi.org/10.1016/j.phytochem.2007.02.011>

- Rohman, M. M., Islam, M. R., Monsur, M. B., Amiruzzaman, M., Fujita, M., Hasanuzzaman, M., 2019: Trehalose protects maize plants from salt stress and phosphorus deficiency. *Plants* 8(12), 568. <https://doi.org/10.3390/plants8120568>
- Sadak, M. S., 2019: Physiological role of trehalose on enhancing salinity tolerance of wheat plant. *Bulletin of the National Research Centre* 43, 53. <https://doi.org/10.1186/s42269-019-0098-6>
- Scagel, C.F., Lee, J., Mitchell, J.N., 2019: Salinity from NaCl changes the nutrient and polyphenolic composition of basil leaves. *Industrial Crops and Products* 127, 119–128. <https://doi.org/10.1016/j.indcrop.2018.10.048>
- Schluepmann, H., van Dijken, A., Aghdasi, M., Wobbes, B., Paul, M., Smeekens, S., 2004: Trehalose mediated growth inhibition of *Arabidopsis* seedlings is due to trehalose-6-phosphate accumulation. *Plant Physiology* 135(2), 879–890. <https://doi.org/10.1104/pp.104.039503>
- Sheyhakinia, S., Bamary, Z., Einali, A., Valizadeh, J., 2020: The induction of salt stress tolerance by jasmonic acid treatment in roselle (*Hibiscus sabdariffa* L.) seedlings through enhancing antioxidant enzymes activity and metabolic changes. *Biologia* 75, 681–692. <https://doi.org/10.2478/s11756-020-00444-8>
- Singh, M., Kumar, J., Singh, S., Singh, V.P., Prasad, S.M., 2015: Roles of osmoprotectants in improving salinity and drought tolerance in plants: a review. *Reviews in Environmental Science and Bio/Technology* 14, 407–426. <https://doi.org/10.1007/s11157-015-9372-8>
- Stark, D. M., Timmerman, K. P., Barry, G. F., Preiss, J., Kishore, G. M., 1992: Regulation of the amount of starch in plant tissues by ADP glucose pyrophosphorylase. *Science* 258(5080), 287–292. <https://doi.org/10.1126/science.258.5080.287>
- Stone, S.L., Gifford, D.J., 1997: Structural and biochemical changes in loblolly pine (*Pinus taeda* L.) seeds during germination and early seedling growth: I. Storage protein reserves. *International Journal of Plant Science* 158(6), 727–737. <https://doi.org/10.1086/297484>
- Theerakulpisut, P., Phongngarm, S., 2013: Alleviation of adverse effects of salt stress on rice seedlings by exogenous trehalose. *Asian Journal of Crop Science* 5(4), 405–415. <https://doi.org/10.3923/ajcs.2013.405.415>
- Türkan, I., Demiral, T., 2009: Recent developments in understanding salinity tolerance. *Environmental and Experimental Botany* 67(1), 2–9. <https://doi.org/10.1016/j.envexpbot.2009.05.008>
- Veluthambi, K., Mahadevan, S., Maheshwari, R., 1981: Trehalose toxicity in *Cuscuta reflexa*: correlation with low trehalase activity. *Plant Physiology* 68(6), 1369–1374. <https://doi.org/10.1104/pp.68.6.1369>
- Veluthambi, K., Mahadevan, S., Maheshwari, R., 1982a: Trehalose toxicity in *Cuscuta reflexa*: cell wall synthesis is inhibited upon trehalose feeding. *Plant Physiology* 70(3), 686–688. <https://doi.org/10.1104/pp.70.3.686>
- Veluthambi, K., Mahadevan, S., Maheshwari, R., 1982b: Trehalose toxicity in *Cuscuta reflexa*: sucrose content decreases in shoot tips upon trehalose feeding. *Plant Physiology* 69(6), 1247–1251. <https://doi.org/10.1104/pp.69.6.1247>
- Wingler, A., Fritzius, T., Wiemken, A., Boller, T., Aeschbacher, R. A., 2000: Trehalose induces the ADP-glucose pyrophosphorylase gene, ApL3, and starch synthesis in *Arabidopsis*. *Plant physiology* 124(1), 105–114. <https://doi.org/10.1104/pp.124.1.105>
- Yemm, E.W., Cocking, E.C., 1955: The determination of amino acids with ninhydrin. *Analyst* 80, 209–214. <https://doi.org/10.1039/AN9558000209>
- Yin, Y.-G., Kobayashi, Y., Sanuki, A., Kondo, S., Fukuda, N., Ezura, H., Sugaya, S., Matsukura, C., 2010: Salinity induces carbohydrate accumulation and sugar-regulated starch biosynthetic genes in tomato (*Solanum lycopersicum* L. cv. 'Micro-Tom') fruits in an ABA- and osmotic stress-independent manner. *Journal of Experimental Botany* 61(2), 563–574. <https://doi.org/10.1093/jxb/erp333>
- Yoon, J.Y., Hamayun, M., Lee, S.K., Lee, I.J., 2009: Methyl jasmonate alleviated salinity stress in soybean. *Journal of Crop Science and Biotechnology* 12, 63–68. <https://doi.org/10.1007/s12892-009-0060-5>
- Zhang, J.L., Shi, H., 2013: Physiological and molecular mechanisms of plant salt tolerance. *Photosynthesis Research* 115, 1–22. <https://doi.org/10.1007/s11120-013-9813-6>
- Zhang, X., Wu, N., Li, C., 2005: Physiological and growth responses of *Populus davidiana* ecotypes to different soil water contents. *Journal of Arid environments* 60(4), 567–579. <https://doi.org/10.1016/j.jaridenv.2004.07.008>
- Zhang, Y., Wang, Y., Wen, W., Shi, Z., Gu, Q., Ahammed, G.J., Guo, S., 2021: Hydrogen peroxide mediates spermidine-induced autophagy to alleviate salt stress in cucumber. *Autophagy* 17(10), 2876–2890. <https://doi.org/10.1080/15548627.2020.1847797>
- Zhao, D.Q., Li, T.T., Hao, Z.J., Cheng, M.L., Tao, J., 2019: Exogenous trehalose confers high temperature stress tolerance to herbaceous peony by enhancing antioxidant systems, activating photosynthesis, and protecting cell structure. *Cell Stress and Chaperones* 24(1), 247–257. <https://doi.org/10.1007/s12192-018-00961-1>
- Zhao, S., Zhang, Q., Liu, M., Zhou, H., Ma, C., Wang, P., 2021: Regulation of plant responses to salt stress. *International Journal of Molecular Sciences* 22(9), 4609. <https://doi.org/10.3390/ijms22094609>

# Mitigation of cadmium toxicity stress by magnetopriming during germination of soybean

Anjali Vyas, Sunita Kataria\*, Rajkumar Prajapati, Meeta Jain\*

School of Biochemistry, Devi Ahilya Vishwavidyalya, Takshila Campus, Khandwa Road, Indore-452001 (M.P.), India

**Abstract** – Cadmium (Cd) is a highly toxic heavy metal that poses a significant threat to food safety and agricultural production worldwide. Its solubility enables easy entry into plants, leading to reductions in seed germination, growth and crop yield. Thus, it is crucial to discover methods to alleviate the harmful impact of Cd on plant growth. Magnetopriming (MP) is a non-invasive and cost-effective technique that has been proposed to improve plant growth under abiotic stress conditions. The current study investigated the potential of MP to improve the seed germination, early seedling growth and biochemical responses of soybean under Cd toxicity. Soybean seeds were magnetoprimed with static magnetic field (SMF) strength of 200 mT for 1 hour before germination. The unprimed and magnetoprimed seeds were grown under different concentrations of cadmium chloride (0, 25, 50, 75, 100, 200 and 300  $\mu\text{M}$   $\text{CdCl}_2$ ) in Petri plates for five days in the dark at  $25 \pm 1$  °C. The results revealed that Cd toxicity significantly reduced the germination percentage and inhibited the seedling growth parameters and increased oxidative stress, as determined by malondialdehyde (MDA) content in soybean seedlings from unprimed seeds. The inhibitory effect was increased with increasing concentration of Cd. However, MP remarkably increased the germination percentage, seedling growth parameters, activities of total amylase and protease, and hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) content and decreased MDA and proline content in germinating soybean seedlings at all concentrations of Cd. These findings suggest that MP can alleviate the adverse effects of Cd stress in soybean seedlings and increase the tolerance index towards Cd toxicity by enhancing the activity of amylolytic and proteolytic enzymes and reducing oxidative stress.

**Keywords:** cadmium toxicity, germination, lipid peroxidation, magnetopriming, tolerance index

## Introduction

Abiotic stressors, which include temperature extremes, water availability, salinity, heavy metals, and UV radiation, can have detrimental effects on plants and result in significant crop yield losses (Saharan et al. 2022). Heavy metals such as arsenic, cadmium, lead, and mercury are toxic to plants and animals even at low concentrations and can accumulate in the environment from human activities such as mining, industrial discharge, and use of pesticides and fertilizers (Alengebawy et al. 2021, Singhal et al. 2023). Cadmium (Cd) is a nonessential, heavy metal element and a widespread environmental contaminant that is persistent, non-biodegradable, bio-accumulative and highly toxic even at low concentrations (Hussain et al. 2019, Mahawar and Shekhawat 2023). Cd toxicity, can affect many physiological and morphological processes in plants, including plant growth, respiration, water and nutrient uptake, and root growth (Chen et al. 2011, Liu et al. 2012, Sheirdil et al. 2012).

Cd causes adverse effects on germination, seedling vigour index, and plant growth (Sheirdil et al. 2012, He et al. 2014). Plants exposed to Cd have damaged photosynthetic apparatus and inhibited chlorophyll biosynthesis, which negatively impacts growth and biomass, leaf chlorophyll fluorescence, and photosynthetic parameters. (Ci et al. 2010, Xue et al. 2014, Yang et al. 2015, Mahawar et al. 2021). Cd contamination in agricultural soils limits crop production, which is a major concern given the increasing demand for food worldwide.

Soybean (*Glycine max* (L.) Merr.) is the most significant legume crop in the world, as 20% of the seed is oil and 40% is high-quality protein (Ferguson and Gresshoff 2009). It plays a significant role in the global oilseed agriculture landscape because of its high productivity, profitability, and essential role in preserving soil fertility. The increase in Cd concentration significantly inhibited seed germination, root

\* Corresponding author e-mails: sunita\_kataria@yahoo.com and mjjainmeeta@gmail.com

and shoot growth in soybean plants (Sheirdil et al. 2012). Therefore, it is crucial to explore the approaches to alleviating heavy metal toxicity in soybean. Seed priming has proven to be an effective strategy for increasing seed vigour and germination, as well as seedling growth and field establishment in unfavourable conditions of heavy metal toxicity (Prajapati et al. 2020). It appears that seed priming is a helpful method for reducing cadmium toxicity in plants (Singhal et al. 2023). In the field of seed technology, physical techniques have proven to be much superior to conventional protocols of osmo, chemical, hydro and hormonal priming (Bilalis et al. 2012). One of the most researched physical pre-sowing seed treatments in agriculture is based on the use of magnetic field (MF) for the alleviation of heavy metal toxicity (cadmium, arsenic and mercury) in plants (Chen et al. 2011, Fatima et al. 2020, Prajapati et al. 2023).

Magnetopriming (MP) has a significant effect on a number of processes, including morpho-structural elements (seed germination, plant growth, and yield) and modifications in the expression of genes linked to nitrogen metabolism and seed germination in non-stress and abiotic stress conditions (Kataria et al. 2020, Sarraf et al. 2020, Raipuria et al. 2021). According to some theories, MF could attract iron particles, growth hormones, and carbohydrates (Xue et al. 2014). It may also have an impact on transcription-related factors and gene expression processes (Stutte et al. 2006, Kataria et al. 2020). Additionally, it might immediately trigger the production of calcium ions ( $\text{Ca}^{2+}$ ) in seed embryos (Kataria et al. 2023). The germination of seeds and the physiological growth of tomato plants are significantly impacted by neodymium earth magnets by the stimulation of seed germination and speed of germination by 50% and production of larger leaf areas and more root hairs in the growing plantlets (Abhary and Akhkhah 2023). It was earlier reported that MP mitigates the adverse effect of cadmium stress in mung beans by reducing the level of malondialdehyde, nitric oxide, hydrogen peroxide and superoxide radical content and increased the growth and photosynthetic performance in mung bean plants (Chen et al. 2011). However, the mitigating effect of MP during seed germination has not been reported for heavy metal toxicity especially for cadmium in soybean seedlings. Hence the objective of the present study is to examine the impact of MP on seed germination, seedling growth and biochemical parameters in soybean under cadmium toxicity and to reveal if MP could enhance tolerance of soybean towards Cd toxicity during early seedling stage.

## Materials and methods

### Plant material

Breeder seeds of soybean (*Glycine max* [L.] var. JS-9560 (commonly grown variety of soybean in the Malwa region, Indore, Madhya Pradesh, India), were obtained from the ICAR-Indian Institute of Soybean Research, Khandwa Road, Indore, M.P., India.

### Magnetic field treatment

To pre-treat the seeds with a static magnetic field (SMF), we used an electromagnet (AETec), fabricated by the Academy of Embedded Technology in Delhi, India. The pole components measured 16 cm in length and 9 cm in diameter while the coil consisted of 3000 turns and a resistance of 16 Ohms. Nearly 100 dry seeds of soybean at a time were placed in a cylindrical sample holder made of thin transparent plastic between the two poles of the electromagnet for magnetopriming, while maintaining the temperature at  $25 \pm 1^\circ\text{C}$ . The distance between the two poles was 5 cm. The SMF was developed using a direct current (DC) power supply (80 V/10 A) with a continuously changing output. Based on previous studies, soybean seeds were treated with 200 mT of SMF for an hour (Kataria et al. 2020). The SMF strength was obtained by adjusting the current and voltage by power supply and the obtained SMF strength was measured using a digital gauss meter (AETech model DGM-102) with a probe consisting of an indium arsenide crystal encapsulated by a  $5 \times 4 \times 1$  mm non-magnetic sheet. The local geomagnetic field was less than 10 mT in the north to south direction. With the exception of the SMF treatment (200 mT for 1 hour) for primed seeds, all other conditions were the same for unprimed and magnetoprimed seeds. Every experiment was carried out concurrently with unprimed seeds (which were used as controls) stored at room temperature ( $25 \pm 1^\circ\text{C}$ ) and kept away from the magnetic field (less than 5 mT).

### Treatment to induce cadmium toxicity

To conduct the germination experiment, both magnetoprimed and unprimed soybean seeds were first surface-sterilized with 0.01%  $\text{HgCl}_2$  for 2 minutes, followed by washing three times with distilled water. The fifteen seeds were then allowed to germinate on wet Whatman filter paper in each Petri plate (15 cm diameter) in three replications with 10 mL of different concentrations of  $\text{CdCl}_2$  (25, 50, 100, 200, and 300  $\mu\text{M}$ ) while 10 mL of distilled water was used as a control (0  $\mu\text{M}$ ). The seeds were kept for germination in darkness at  $25 \pm 1^\circ\text{C}$  for 5 days in the incubator. Germination percentage was calculated based on the number of normal seedlings, as the ratio of the number of germinated seeds to the total number of seeds on the third day of imbibition in different concentrations of  $\text{CdCl}_2$ .

### Early growth characteristics of seedlings

To estimate the effect of magnetopriming on soybean seedling growth, ten seedlings were randomly selected from each treatment (magnetoprimed and unprimed seeds germinated under different concentrations of  $\text{CdCl}_2$ ) in biological triplicates ( $N = 3$ ). The length of shoot and root, along with the length of the whole seedling, were measured for each selected seedling. Fresh weight was also measured. For dry weight determination, the seedlings were dried in an oven at  $80^\circ\text{C}$  for 72 h and weighed. The vigour index was calculated using the formulae proposed by Abdul-Baki and Anderson (1973):

Vigour index I = germination % × seedling length (root + shoot) cm

Vigour index II = germination % × seedling dry weight (root + shoot) g

Vigour index I is the multiple of the percentage of germination and seedling length, and Vigour index II is the multiple of percentage of germination and seedling dry weight.

### Enzyme activities during germination

Total amylase and protease activities were measured in the soybean seedlings derived from unprimed and magnetoprimed seeds after 5 days (120 h) of imbibition under different CdCl<sub>2</sub> concentrations. After 5 days of imbibition, the fresh seedlings with root, shoot and cotyledons were taken for analysis of enzymes and biochemical parameters. Each experiment was repeated in three biological replicates with three technical replicates.

The assay of total amylase activity was performed using the method of Sawhney et al. (1970). The seedling homogenate (100 mg) was mixed with 5 mL of 80% chilled acetone and centrifuged at 15000 rpm for 10 min at 4 °C. The resulting pellet was dissolved in 10 mL of 0.2 M phosphate buffer (pH 6.4) and centrifuged again at 15000 rpm for 20 min at 4 °C. The amylase activity in the obtained supernatant was then determined by the addition of 2 mL of phosphate buffer (pH 6.4), 1 mL of starch (1%), and incubation for 30 min at room temperature. Subsequently, 1 mL of 0.1 M HCl and 0.1 mL of 0.1 M potassium iodide were added to the reaction mixture and the absorbance was measured at 660 nm using a spectrophotometer. The total amylase activity was expressed as mg starch hydrolysed g<sup>-1</sup> fresh weight of seedlings h<sup>-1</sup>.

The assay of protease activity was carried out according the method of Kunitz (1947) modified by Kataria et al. (2023). The 1.0 g seedlings were crushed in 0.2 M phosphate buffer (pH 7.4) and centrifuged at 13800 rpm at 4 °C for 30 min. To 0.5 mL of supernatant, 0.5 mL of casein (1%) prepared in 0.2 M carbonate buffer (pH 9.2) was added and the mixture was incubated for 10 min at 37 °C. The reaction was terminated by the addition of 1 mL of 10% trichloroacetic acid (TCA) and centrifugation at 13800 rpm for 10 min at 4 °C. After centrifugation, 2.5 mL of carbonate buffer (0.44 M, pH 9.5) was added and the development of a blue colour was observed upon addition of 0.5 mL Folin's reagent followed by incubation for 30 min at room temperature. The protein content was measured at 660 nm against carbonate buffer (0.44 M, pH 9.5) and Folin's reagent taken as a blank and the protease activity was represented as mg protein hydrolysed g<sup>-1</sup> fresh weight of seedlings.

### Determination of biochemical parameters

Thiobarbituric acid reactive substances (TBARS) were used to measure the level of malondialdehyde (MDA), according to the method described by Heath and Packer

(1968). Soybean seedlings (100 mg) were homogenized in 1 mL of 0.1% (w/v) TCA, and the resulting homogenate was centrifuged at 12000 rpm for 15 min at 4 °C. The supernatant was used for the TBARS assay. Specifically, 0.5 mL of the supernatant was mixed with 1 mL of 0.5% (w/v) 2-thiobarbituric acid (TBA) prepared in 20% TCA and incubated in a boiling water bath for 30 min. The reaction was stopped by placing the tube in an ice bath, and the mixture was then centrifuged at 12000 rpm for 5 min. The absorbance of the supernatant was measured at 532 nm, and the value for the non-specific absorbance read at 600 nm was subtracted. TBA (0.5%) in TCA (20%) was taken as blank. The amount of MDA was calculated using an extinction coefficient ( $\epsilon = 155 \text{ mM}^{-1} \text{ cm}^{-1}$ ) and expressed as  $\mu\text{mol MDA mg}^{-1}$  fresh weight of seedlings.

The proline content was calculated using the method of Bates et al. (1973). Using a mortar and pestle, 500 mg of soybean seedlings were homogenised in 10 mL of 3% (w/v) aqueous sulphosalicylic acid; the homogenate was then centrifuged for 5 min at 4 °C at 10000 rpm. The proline was estimated using the supernatant. Two mL each of glacial acetic acid, acid ninhydrin reagent, and supernatant were combined. The mixture was boiled at 100 °C for 1 h. The reaction was terminated by cooling in an ice bath and 4 mL of toluene was added. Following thorough mixing, the toluene-containing chromophore was isolated, and the red colour generated absorbance was measured at 520 nm using toluene as the blank. The proline content was calculated through the standard curve prepared in the range of 5–25  $\mu\text{g}$  proline and expressed as  $\mu\text{g proline g}^{-1}$  fresh weight of the seedlings.

The estimation of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) was carried out by the method described by Mukherjee and Choudhuri (1983), which involves the formation of a titanium-hydroperoxide complex. First, 0.5 g of soybean seedlings were crushed in chilled acetone (5 mL) and filtered using Whatman No.1 filter paper. The filtrate (5 mL) was mixed with titanium reagent (2 mL; containing 5% (w/v) titanium oxide and potassium sulphate digested in concentrated sulphuric acid) and ammonium hydroxide solution (2.5 mL) to precipitate the titanium-hydroperoxide complex. The resulting precipitate was centrifuged at 13800 rpm for 15 min at 4 °C, and the pellet was resuspended in concentrated sulphuric acid (2 M) and centrifuged again. The absorbance of the supernatant was then measured at 415 nm against sulphuric acid (2 M) using a Shimadzu Spectrophotometer (UV-1800) and the amount of H<sub>2</sub>O<sub>2</sub> was calculated and expressed as  $\mu\text{mol g}^{-1}$  fresh weight.

### Tolerance index (TI)

Tolerance index (in terms of root length) of seedlings from unprimed and magnetoprimed seeds grown under different concentrations of CdCl<sub>2</sub> was determined with the following formula given by Iqbal and Rahmati (1992):

$$\text{Tolerance index (T.I.)} = \frac{\text{mean root length in metal solution}}{\text{mean root length in control}} \times 100$$

**Statistical analysis**

Three replications and fully randomised designs were used to arrange the samples. The mean ± standard error (S.E.) of the three biological replicates (N = 3) was used to show the data. Ten seedlings were taken for each replication in order to measure the many aspects of seedling growth, including length of seedlings, root and shoot, fresh and dry weight of seedling, and vigour indices. The mean of three seedlings in each replica serves as the data for the biochemical analysis. The data were analysed by Student's t-test and significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as following \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. Statistically significant differences between magnetoprimed and unprimed seedlings grown in non-stress as well as Cd-stress conditions are marked as #P < 0.05, ##P < 0.01, ###P < 0.001.

**Results**

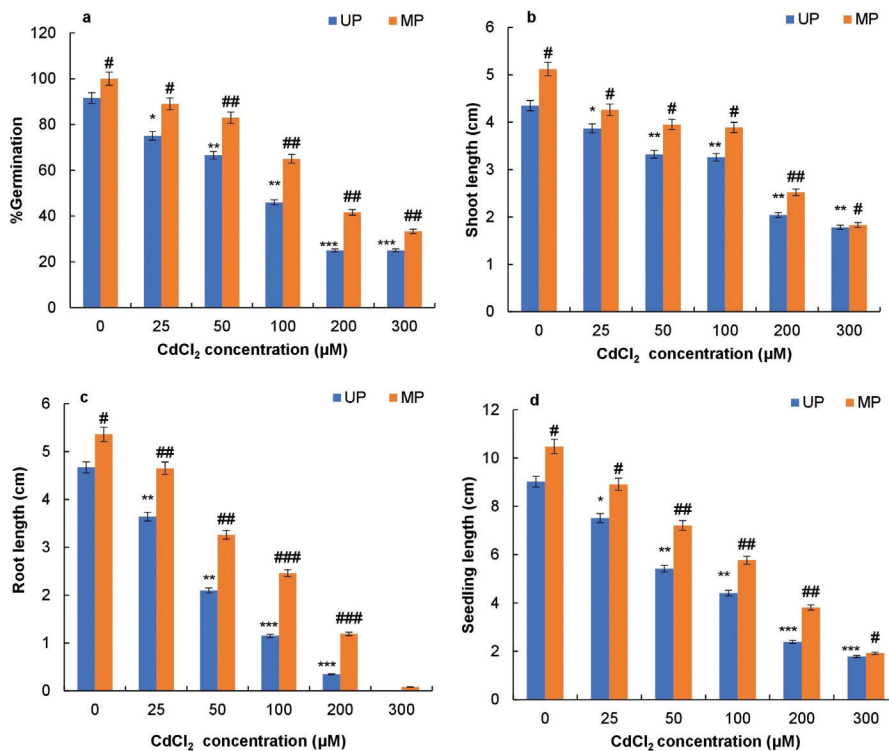
**Effect of MP on seed germination and seedling development under cadmium treatments**

Germination percentage decreased with increasing CdCl<sub>2</sub> concentration in magnetoprimed and unprimed seeds of soybean. A remarkable decrease of 50% was observed at 100 µM CdCl<sub>2</sub> and 72% was observed at 200 and 300 µM CdCl<sub>2</sub> in unprimed seeds (Fig. 1a). Magnetopriming of seeds enhanced percentage germination at all the

concentrations of CdCl<sub>2</sub> used as compared to respective unprimed seeds. Maximum enhancement in percentage germination by MP (41%, 66% and 33%) was obtained at 100, 200 and 300 µM CdCl<sub>2</sub>, respectively (Fig. 1a).

Root, shoot and seedling lengths were reduced with increasing CdCl<sub>2</sub> concentration in magnetoprimed and unprimed seeds of soybean (Fig. 1b-1d). The decrease in root length was more prominent than the decrease in shoot length. Maximum decreases of 75% and 93% were observed in root length of seedlings from unprimed seeds at 100 and 200 µM CdCl<sub>2</sub> and root growth was completely inhibited at 300 µM CdCl<sub>2</sub> (Fig. 1b). Magnetoprimed soybean seeds demonstrated higher shoot and root length at all the concentrations of CdCl<sub>2</sub> used as compared to corresponding controls (Fig. 1b, 1c). Magnetopriming caused increases of 18, 19 and 24% in shoot length of seedlings at 50, 100 and 200 µM CdCl<sub>2</sub>, respectively (Fig. 1b). Similarly, significant promotion in root length by 55, 113 and 240% at 50, 100 and 200 µM CdCl<sub>2</sub> respectively was obtained in seedlings from magnetoprimed seeds as compared to the seedlings of unprimed seeds (Fig. 1c).

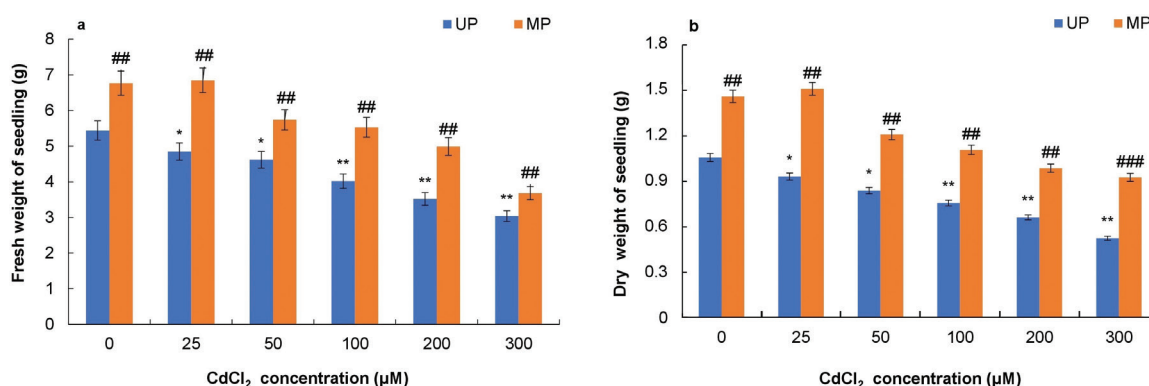
There was substantial decrease in length of seedlings observed at a higher concentration (300 µM) of CdCl<sub>2</sub> (Fig. 1d). The CdCl<sub>2</sub> at 100, 200 and 300 µM concentrations caused significant inhibition of seedling length by 51%, 73% and 80% respectively. On the other hand, a maximum increase of 31% was observed at 100 µM CdCl<sub>2</sub> followed by a 59% increase at 200 µM CdCl<sub>2</sub> in magnetoprimed seeds (Fig. 1d).



**Fig. 1.** Effect of magnetopriming on percentage germination (a), shoot length (b), root length (c) and seedling length (d) of soybean seedlings under Cd toxicity. The vertical lines on the bar indicate ± S.E. for mean of triplicates (N = 3). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as #P < 0.05, ##P < 0.01; ###P < 0.001.

### Effect of MP on fresh weight and dry weight of soybean seedling under cadmium treatments

Fresh and dry weight of unprimed and primed seeds gradually decreased with increasing CdCl<sub>2</sub> concentration (Fig. 2a, 2b). CdCl<sub>2</sub> treatments caused reduction in fresh weight by 26%, 35% and 44% respectively at 100, 200 and 300 μM CdCl<sub>2</sub> concentration as compared to the controls (Fig. 2a). Further, there was significant increase in fresh and dry weight of seedling observed in primed seeds as compared to unprimed ones at all the concentrations of Cd used. Maximum of 24, 37, 42 and 21% promotion in fresh weight of seedlings was found in magnetoprimed as compared to unprimed seed respectively at 50, 100, 200 and 300 μM CdCl<sub>2</sub> (Fig. 2a). Similar results were observed for dry weight of seedlings (Fig. 2b).



**Fig. 2.** Effect of magnetopriming on fresh weight (a) and dry weight (b) of soybean seedlings under Cd toxicity. The vertical lines on the bar indicate ± S.E. for mean of triplicates (N = 3). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \*P < 0.05, \*\*P < 0.01. Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as ##P < 0.01.

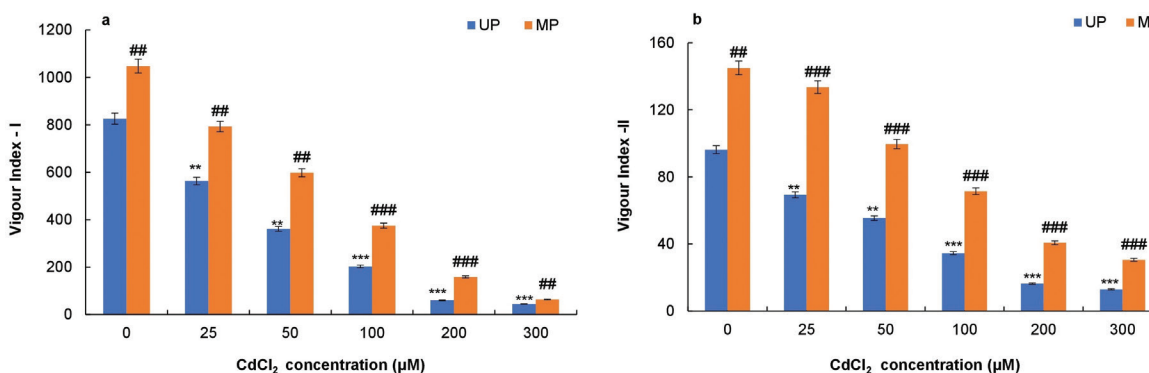
### Effect of MP on vigour indices of soybean seedling under cadmium treatments

Treatment of soybean seeds with different concentrations of CdCl<sub>2</sub> reduced the seed vigour indices gradually from 0 to 100 μM CdCl<sub>2</sub>, thereafter significant decrease was observed at 200 and 300 μM in both primed and unprimed seeds (Fig. 3a, 3b). Vigour index-I was found decreased

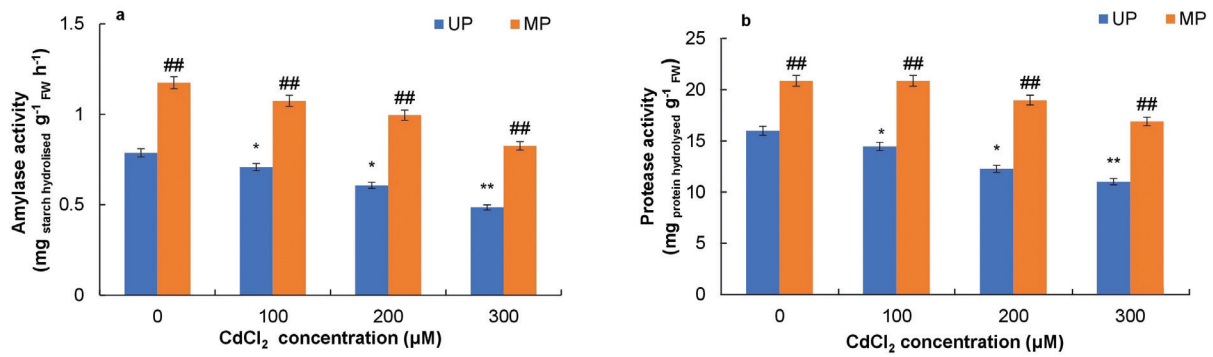
prominently by 75%, 92% and 94% respectively at 100, 200 and 300 μM CdCl<sub>2</sub> concentration in unprimed seeds compared to their controls (Fig. 3a). Similarly, vigour index-II decreased severely at 200 and 300 μM CdCl<sub>2</sub> concentrations by 83% and 87% respectively in seeds compared to the control (Fig. 3b). However, magnetoprimed seeds showed higher vigour indices at all CdCl<sub>2</sub> levels as compared to unprimed seeds. There was prominent increase in vigour index-I at 50, 100 and 200 μM CdCl<sub>2</sub> concentration (84%, 165% and 43% respectively) in magnetoprimed seeds as compared to unprimed seeds (Fig. 3a). Similarly, vigour index-II increased significantly at 100, 200 and 300 μM CdCl<sub>2</sub> concentration by 106, 147 and 135% respectively in magnetoprimed seeds as compared to unprimed seeds (Fig. 3b).

### Effect of MP on total amylase and protease activities in soybean seedlings under cadmium treatments

When soybean seeds were stressed with CdCl<sub>2</sub>, total amylase and protease activities decreased slightly in the seedlings from unprimed seeds as compared to control seedlings. However, primed seeds showed increased amylase activity by 49%, 51%, 64% and 70% at 0, 100, 200 and 300 μM



**Fig. 3.** Effect of magnetopriming on vigour index-I (a) and vigour index-II (b) of soybean seedlings under Cd toxicity. The vertical lines on the bar indicate ± S.E. for mean of triplicates (N = 3). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \*\*P < 0.01, \*\*\*P < 0.001. Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as ##P < 0.01, ####P < 0.001.



**Fig. 4.** Effect of magnetopriming on amount of starch hydrolysed (a) and protein hydrolysed (b) in soybean seedlings under Cd toxicity. The vertical lines on the bar indicate  $\pm$  S.E. for mean of triplicates (N = 3). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \*P < 0.05, \*\*P < 0.01. Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as ##P < 0.001. FW – fresh weigh.

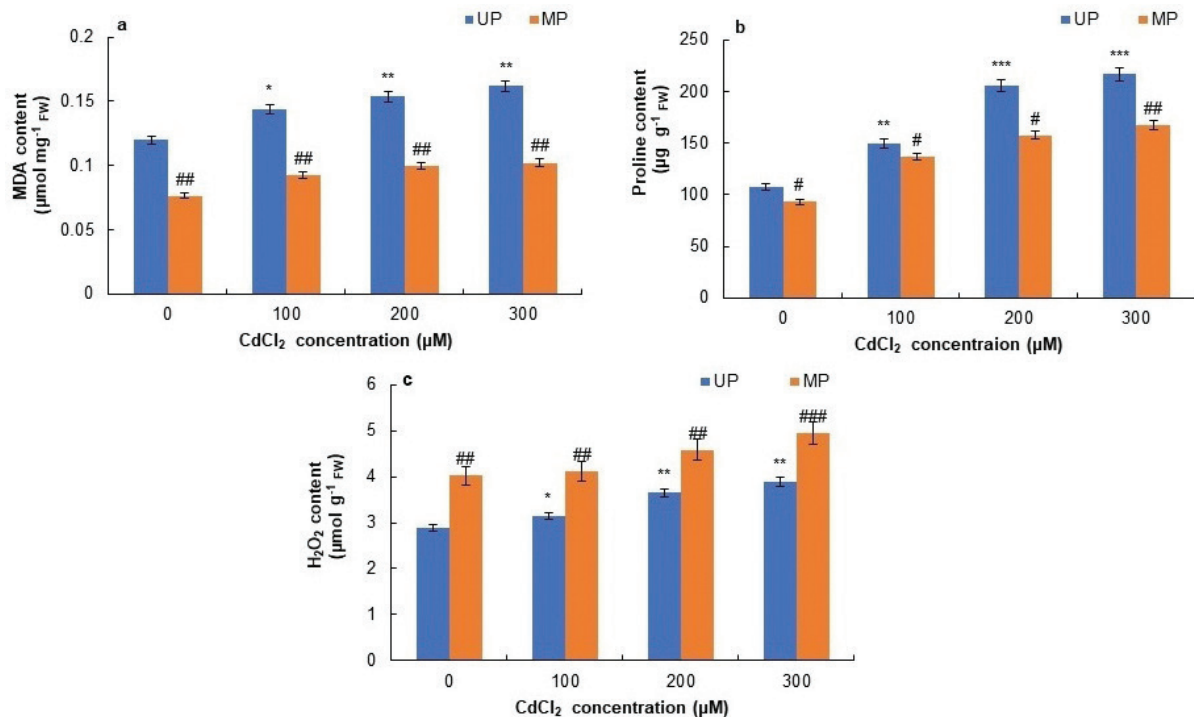
CdCl<sub>2</sub> concentrations, respectively (Fig. 4a). Further, magnetoprimed seeds showed increased protease activity by 30, 44, 54 and 53% at 0, 100, 200 and 300 µM CdCl<sub>2</sub> concentrations, respectively (Fig. 4b).

**Effect of MP on proline, MDA and H<sub>2</sub>O<sub>2</sub> content in soybean seedlings under cadmium treatments**

MDA content was found to rise gradually with increasing concentration of CdCl<sub>2</sub> in unprimed as well as primed seedlings. However, the level of MDA was higher in seedlings emerged from unprimed seeds at all the concentrations of CdCl<sub>2</sub> used (Fig. 5a). MDA content was decreased

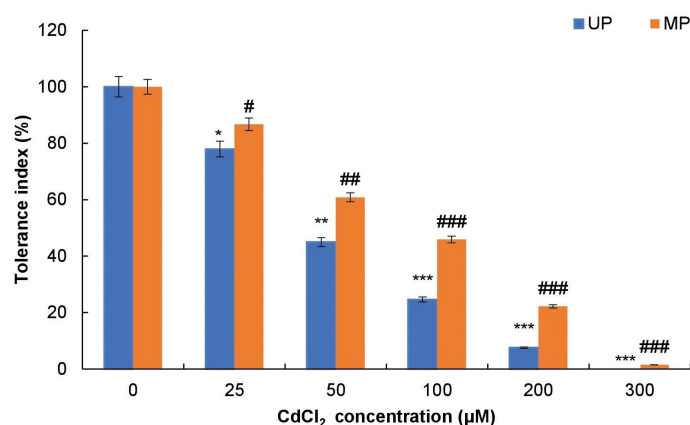
in the seedlings from SMF-primed seeds as compared to the seedlings from unprimed seeds at all the concentrations of CdCl<sub>2</sub> (0 to 300 µM) (Fig. 5a). A remarkable decrease in MDA content (by 37%) was observed at 300 µM CdCl<sub>2</sub> concentration in the seedlings from primed seeds as compared to the seedlings of unprimed seeds.

Proline content was considerably increased in seedlings of unprimed seeds at all the concentrations of CdCl<sub>2</sub> (Fig. 5b). The enhancement in proline content was 39, 92 and 100% in seedlings of unprimed seeds at 100, 200 and 300 µM CdCl<sub>2</sub> concentrations as compared to the control seedlings. Magnetopriming of seeds lowered the proline quan-



**Fig. 5.** Effect of magnetopriming on malondialdehyde – MDA (a), proline (b) and H<sub>2</sub>O<sub>2</sub> (c) content of soybean seedlings under Cd toxicity. The vertical lines on the bar indicate  $\pm$  S.E. for mean of triplicates (N = 3). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001. Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as #P < 0.05, ##P < 0.01. FW – fresh weight.





**Fig. 6.** Effect of magnetopriming on tolerance index of soybean seedlings in terms of root length under Cd toxicity. The vertical lines on the bar indicate  $\pm$  S.E. for mean of triplicates ( $N = 3$ ). Significant differences between non-stressed and Cd-stressed soybean seedlings that emerged from unprimed seeds are marked as follows: \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ . Statistically significant differences between magnetoprimed (MP) and unprimed (UP) seedlings grown in non-stress as well as Cd-stress conditions are marked as \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

tivity at all concentrations of CdCl<sub>2</sub>. There was 23% decrease in proline content at 200 µM and 300 µM CdCl<sub>2</sub> concentration in primed seeds (Fig. 5b).

The cadmium treatments increased the H<sub>2</sub>O<sub>2</sub> content by 26% (200 µM CdCl<sub>2</sub>) and 35% (300 µM CdCl<sub>2</sub>) in the seedlings of unprimed seeds as compared to their controls (Fig. 5c). Magnetopriming also caused a further increase in H<sub>2</sub>O<sub>2</sub> content in soybean seedlings in comparison to unprimed seeds at all the cadmium chloride concentrations.

#### Effect of MP on tolerance index (TI) of soybean seedlings under cadmium treatments

TI was 100% for control seedlings and it decreased with an increase in CdCl<sub>2</sub> toxicity (Fig. 6). The TI after treatment with 25, 50, 100, 200 and 300 µM CdCl<sub>2</sub> was 78, 45, 25, 8 and 0%, respectively, for soybean seedlings obtained from unprimed seeds (Fig. 6). Whereas seeds pre-treated with MP showed higher values of TI at all the levels of CdCl<sub>2</sub> i.e. 87, 61, 46, 22 and 2% TI compared to values from unprimed seeds at 25, 50, 100, 200 and 300 µM CdCl<sub>2</sub>, respectively.

## Discussion

Amongst heavy metals, Cd is a toxic environmental contaminant, which has detrimental effect on plants, animals and humans. It is often found in industrial and agricultural waste and can contaminate soil, water, and food (Yin et al. 2021). The germinating seed serves as the first point of interaction between the developing plant and its surrounding environment, making it particularly susceptible to the toxic effects of Cd (Stefanello 2019, Mahawar et al. 2021). Cd significantly inhibited the seed germination of various plant species such as swiss chard, lettuce, wheat, basil, chia, beans and spinach (Ahmad et al. 2012, Bautista et al. 2013, El Rasafi et al. 2016, Gharebaghi et al. 2017, Stefanello 2019).

The results of the present investigation also showed that, in seedlings derived from unprimed seeds, Cd treatments significantly inhibit seed germination and early seedling

growth parameters (root, shoot, and seedling length, fresh and dry weight of seedlings, and vigour indices); a dose-response inhibition was noted for every parameter examined. Even in the presence of cadmium, however, MP of soybean seeds with SMF at 200 mT for an hour before germination improved the seeds' early development characteristics. The length of both shoot and root is critical for efficient material exchange and nutrient acquisition (Bewley and Black 1994, Stefanello 2019). The inhibition was more pronounced for root length in the seedlings from unprimed seeds grown under Cd treatments. At a higher concentration of Cd (300 mM CdCl<sub>2</sub>), the root growth was completely inhibited. Interestingly, MP with SMF enhanced root length to a greater extent than the shoot length. In the present study, positive effects of MP were observed in soybean seedlings under Cd toxicity, such as increased seed germination percentage, root, shoot, and seedling length, as well as enhanced fresh/dry weight and vigour indices of seedlings. The inhibitory effects of Cd on shoot and root growth lead to stunted seedlings as Cd toxicity disrupts the water and nutrient uptake and transport (Liu et al. 2012, Sheirdil et al. 2012, Stefanello 2019). The increase in root length induced by MP can contribute to enhanced water and nutrient uptake and provide necessary resources for seedlings to cope with the toxic effects of Cd. However, the exposure of seeds to MF has earlier been shown to increase their capacity to absorb moisture and their nutrient content in soybean (Kavi 1977, Radhakrishnan and Kumari 2012). These effects on moisture absorption and physiological activity can contribute to improved germination and seedling vigour. There have also been reports on the effects of very low-frequency alternating magnetic fields on cell membrane ionic permeability (Khizhenkov et al. 2001, Thomas et al. 2013). Studies by Kataria et al. (2019, 2020, 2021, 2023) have confirmed that applying MP with SMF-pretreatment to the seeds can have a positive effect on seed germination and seedling vigour under salt stress. Our results are consistent with the findings of several earlier studies, including those on rice, wheat, maize, soybean and

barley seeds, which have also revealed that MP can improve seed germination percentage, seedling growth and vigour index (Carbonell et al. 2000, Florez et al. 2007, Martinez et al. 2009, Shine et al. 2012, Kataria et al. 2015, Ercan et al. 2022). The tolerance index represents the ability of seedlings to withstand and survive under CdCl<sub>2</sub> toxicity, decreased with increasing concentrations of CdCl<sub>2</sub> in both unprimed and magnetoprimed seedlings. However, the seedlings from magnetoprimed seeds exhibited a higher tolerance index than unprimed seed at all the concentrations of Cd. This suggests that MP helped to mitigate the inhibitory effects of cadmium on seedling growth and increased their tolerance to Cd toxicity. Similarly, Kataria et al. (2020, 2022) have reported that the salt tolerance index of soybean seedlings was increased by MP due to the higher nitric oxide production by SMF pre-treatment, which helps to maintain the balance of abscisic acid (ABA) and gibberellic acid (GA) for higher seed germination and seedling growth under salt stress conditions.

Furthermore, high concentrations of Cd have negative effects on hydrolyzing enzymes including acid phosphatases (ACPs) and  $\alpha$ -amylases, they prevent reserved carbohydrates from being hydrolyzed and from moving from the endosperm to the growing embryonic axis. As a result, the embryo that is germinating becomes starved (Kuriakose and Prasad, 2008). Amylase is an enzyme that breaks down the stored carbohydrate reserves of a seed during germination. Also, it has been shown that Cd's interactions with proteolytic enzymes limit store protein catabolism, which results in the slowing of seedling growth (Gianazza et al., 2007). The magnetoprimed soybean seeds maintained a higher total amylase and protease activity than unprimed seeds under both non-stress and in cadmium stressed conditions. This suggests that the MP may have influenced the biochemical mechanisms involved in seed germination and growth, including amylase and protease activity. Our finding is consistent with the results of previous studies. For instance, Vashisth and Nagarajan (2010) reported that magnetoprimed wheat and sunflower seeds had significantly higher amylase activity than their controls. Previous studies have reported higher  $\alpha$ -amylase and protease activity in magnetoprimed soybean and maize seedlings under various stress conditions such as salt, mercury toxicity and UV-B (Kataria et al. 2017, 2019, 2020, 2023, Raipuria et al. 2021, Prajapati et al. 2023). The higher protease activity in magnetoprimed seeds could contribute to the breakdown of stored proteins into amino acids (Rajendra et al. 2005) that can be utilized for energy and growth during seedling development. The results suggest that MP can enhance the seedling's ability to cope with stress conditions by increasing total amylase and protease activities, which might improve the crop tolerance under Cd toxicity. Theoretical studies have shown that a magnetic field may enhance the density of ions passing through the cellular membrane of a seed, so affecting the osmotic pressure in a way that favours water entering into the cell (Reina et al. 2001). As previously indicated, seeds exposed to either a static or an oscillating magnetic field had higher levels of enzyme activity, particu-

larly  $\alpha$ -amylase (Kataria et al. 2017, 2020). The enzymes  $\alpha$ -amylase which help the seed use its stored starch energy, are essential for seed germination. Therefore, a larger amount of solvent would be supplied to the seed's germination enzymes more quickly by the magnetic field's osmotic pressure action (Diehl 2022).

Interestingly, MP was found to lower the levels of MDA and oxidative stress marker (proline) and enhanced the H<sub>2</sub>O<sub>2</sub> content in soybean seedlings in the presence of Cd toxicity. This suggests that MP may play a protective role against oxidative damage caused by CdCl<sub>2</sub> stress in plants. Similar findings were reported by Chen et al. (2011), where MF treatment was found to reduce the toxic effects of Cd on mung bean by increasing the rate of photosynthesis, nitric oxide concentration, and nitric oxide synthase activity, and reducing the lipid peroxidation. Chen et al. (2017) found the alleviation of the adverse effect of Cd and lead (Pb) through MP due to enhanced lipid peroxidation and antioxidant defence system in wheat seedlings. According to Prajapati et al. (2023), MP reduced lipid peroxidation at 200 mT (1 h) while increased the H<sub>2</sub>O<sub>2</sub> content in soybean seedlings grown under mercury toxicity. This implies that MP alters membrane stability and H<sub>2</sub>O<sub>2</sub> content to regulate antioxidant activity. Fatima et al. (2020) also observed that MP of soybean seeds mitigated the arsenic toxicity in terms of growth, photosynthesis, and water uptake through increase in the thickness of the midrib of leaves.

Several studies have reported changes in proline content in plants under different stress conditions and magnetic treatments. Banerjee and Roychoudhury (2015) stated that proline serves as an osmoprotectant in plants cultivated in a range of stressful conditions. For instance, cadmium toxicity caused a significant increase in proline accumulation in the shoots of *Brassica juncea*, *Triticum aestivum*, and *Vigna radiata*, indicating that proline plays an important role in the response to cadmium stress (Alia and Saradhi 1991, Sharmila et al. 2017). Mercury treatments raised the proline content in *Melissa officinalis* (Safari et al. 2019) and *Eichhornia crassipes* (Malar et al. 2015), which is consistent with our findings. However, magnetic treatment with SMF has been found to decrease the proline content in soybean seedlings under mercury toxicity (Prajapati et al. 2023). Chickpea plants (Hozayn et al. 2022), *Salvia* (Khosrojerdi et al. 2023) and orange seedlings (Mahmoud et al. 2019) treated with magnetic water also showed decreased amount of proline under saline soils. Okba et al. (2022) reported a decrease in proline content with magnetic water treatment, suggesting a positive role of magnetic water in reducing the detrimental effects of water deficit stress. These authors suggested that proline is especially important during stressful situations because it helps in regulation of the osmotic pressure of the cytosol and the vacuole with that of the external environment, which improves the uptake of water and nutrients. Plants have the capability to produce ROS in response to stress, which can have both positive and negative effects on their growth and survival (Hasanuzzaman et al. 2020). H<sub>2</sub>O<sub>2</sub> production was enhanced markedly by Cd toxicity in a num-

ber of plant species (Ahmad et al. 2011, Mahawar et al. 2021). However, Shine et al. (2012) have shown that priming with SMF can increase the content of ROS which can serve as key signalling agents, influencing critical events in seed life, including seed germination by inducing ABA catabolism and gibberellic acid (GA) biosynthesis. Magnetoprimed seeds showed a decrease in ABA concentration and a rise in GA content in soybean under salinity suggesting that MP actively contributes to seed germination and the release of dormancy (Kataria et al. 2022). Thus the production of ROS such as H<sub>2</sub>O<sub>2</sub> induced by MP of seeds represents one of the mechanisms through which seed invigoration and enhanced seedling growth can be achieved under nonstress as well as under abiotic stress conditions (Kataria et al. 2017, 2021, 2023, Raipuria et al. 2021). According to Barba-Espin et al. (2010), during the germination of pea seeds, H<sub>2</sub>O<sub>2</sub> plays a direct role as a signalling molecule within the phytohormone network, causing the induction of proteins linked to plant signalling and development processes. Therefore, H<sub>2</sub>O<sub>2</sub> functions as a central node that harmonizes and integrates phytohormone interactions to improve magnetoprimed soybean seed germination and vigour under Cd toxicity. Therefore, MP with SMF could be a promising technique for improving germination of seeds and seedling growth by alleviating cadmium stress through improving the tolerance of soybean seedlings. But further investigation is required to completely comprehend the processes behind the advantageous effects of-MP in the presence of heavy metal toxicity.

## Conclusion

In conclusion, the present study revealed that the presence of Cd had a detrimental effect on seed germination and seedling growth, and this effect was found to be concentration dependent. Total amylase and protease activity declined under Cd toxicity, suggesting that it might be the result of disruptions in the reserve mobilisation process from cotyledons to the developing embryonic axis. The tolerance index of seedlings emerged from magnetoprimed seeds indicated that MP mitigates the toxic effects induced by Cd and enhances the tolerance in soybean seedlings by promoting root length, seedling vigour, total amylase and protease activity, H<sub>2</sub>O<sub>2</sub> and reducing MDA and proline content as compared to the seedlings from unprimed seeds under Cd toxicity. Overall, the study suggested the possibility that use of a magnetic field might mitigate the harmful effects of Cd toxicity in soybean. Further research in this area could explore the underlying mechanisms and optimize the magnetic field parameters for improved crop productivity and environmental sustainability.

## References

- Abdul-Baki, A. A., Anderson, J. D., 1973: Vigor determination in soybean seed by multiple criteria I. *Crop Science* 13(6), 630-633.
- Abhary, M. K., Akhka, A., 2023: Effects of neodymium magnetopriming on seed germination and salinity tolerance in tomato. *Bioelectromagnetics* 44(1-2), 47-56. <https://doi.org/10.1002/bem.22438>
- Ahmad, K., Bhatti, I. A., Muneer, M., Iqbal, M., Iqbal, Z., 2012: Removal of heavy metals (Zn, Cr, Pb, Cd, Cu and Fe) in aqueous media by calcium carbonate as an adsorbent. *International Journal of Chemical and Biochemical Sciences* 2, 48-53.
- Ahmad, P., Nabi, G., Ashraf, M., 2011: Cadmium-induced oxidative damage in mustard [*Brassica juncea* (L.) Czern. & Coss.] plants can be alleviated by salicylic acid. *South African Journal of Botany* 77(1), 36-44. <https://doi.org/10.1016/j.sajb.2010.05.003>
- Alengebawy, A., Abdelkhalek, S. T., Qureshi, S. R., Wang, M.-Q., 2021: Heavy metals and pesticides toxicity in agricultural soil and plants: Ecological Risks and Human Health Implications. *Toxics* 9(3), 42. <https://doi.org/10.3390/toxics9030042>
- Alia, P. P., Saradhi, N. D., P. Mohanty., 1991: Proline enhances primary photochemical activities in isolated thylakoid membranes of *Brassica juncea* by arresting photoinhibitory damage. *Biochemistry and Biophysics Research Communications* 181(3), 1238-1244. [https://doi.org/10.1016/0006-291X\(91\)92071-Q](https://doi.org/10.1016/0006-291X(91)92071-Q)
- Banerjee, A., Roychoudhury, A., 2015: WRKY proteins: signalling and regulation of expression during abiotic stress responses. *The Scientific World Journal*, 807560. <https://doi.org/10.1155/2015/807560>
- Barba-Espin, G., Diaz-Vivancos, P., Clemente-Moreno, M. J., Albacete, A., Faize, L., Faize, M., Pérez-Alfocea, F., Hernández, J. A., 2010: Interaction between hydrogen peroxide and plant hormones during germination and the early growth of pea seedlings. *Plant Cell and Environment* 33(6), 981-994. <https://doi.org/10.1111/j.1365-3040.2010.02120.x>
- Bates, L. S., Waldren, R. A., Teare, I. D., 1973: Rapid determination of free proline for water-stress studies. *Plant and Soil* 39, 205-207.
- Bautista, O. V., Fischer, G., Cárdenas, J. F., 2013: Cadmium and chromium effects on seed germination and root elongation in lettuce, spinach and Swiss chard. *Agronomía Colombiana* 31(1), 48-57.
- Bewley, J. D., Black, M., 1994: Seeds: physiology of development and germination. Springer Science and Business Media.
- Bilalis, D. J., Katsenios, N., Efthimiadou, A., Karkanis, A., Efthimiadis, P., 2012: Investigation of pulsed electromagnetic field as a novel organic pre-sowing method on germination and initial growth stages of cotton. *Electromagnetic Biology and Medicine* 31(2), 143-150. <https://doi.org/10.3109/15368378.2011.624660>
- Carbonell, M. V., Martinez, E., Amaya, J. M., 2000: Stimulation of germination in rice (*Oryza sativa* L.) by a static magnetic field. *Electro- and Magnetobiology* 19(1), 121-128. <https://doi.org/10.1081/JBC-100100303>
- Chen, Y., Li, R., He, J.-M., 2011: Magnetic field can alleviate toxicological effect induced by cadmium in mungbean seedlings. *Ecotoxicology* 20, 760-769. <https://doi.org/10.1007/s10646-011-0620-6>
- Chen, Y. P., Chen, D., Liu, Q., 2017: Exposure to a magnetic field or laser radiation ameliorates effects of Pb and Cd on physiology and growth of young wheat seedlings. *Journal of Photochemistry and Photobiology B: Biology* 169, 171-177. <https://doi.org/10.1016/j.jphotobiol.2017.03.012>
- Ci, D., Jiang, D., Wollenweber, B., Dai, T., Jing, Q., Cao, W., 2010: Cadmium stress in wheat seedlings: growth, cadmium accumulation and photosynthesis. *Acta Physiologia Plantarum* 32, 365-373. <https://doi.org/10.1007/s11738-009-0414-0>

- Diehl, K. A., 2022: The effects of weak static magnetopriming on seed germination and plant growth. *The Expedition* 13.
- El Rasafi, T., Nouri, M., Bouda, S., Haddioui, A., 2016: The effect of Cd, Zn and Fe on seed germination and early seedling growth of wheat and bean. *Ekológia (Bratislava)* 35(3), 213–223. <https://doi.org/10.1515/eko-2016-0017>
- Ercan, I., Tombuloglu, H., Alqahtani, N., Alotaibi, B., Bamhrez, M., Alshumrani, R., Ozcelik, S., Kayed, T.S., 2022: Magnetic field effects on the magnetic properties, germination, chlorophyll fluorescence, and nutrient content of barley (*Hordeum vulgare* L.). *Plant Physiology and Biochemistry* 170, 36–48. <https://doi.org/10.1016/j.plaphy.2021.11.033>
- Fatima, A., Kataria, S., Prajapati, R., Jain, M., Agrawal, A. K., Singh, B., Kashyap, Y., Tripathi, D. K., Singh, V. P., Gadre, R., 2020: Magnetopriming effects on arsenic stress-induced morphological and physiological variations in soybean involving synchrotron imaging. *Physiol Plantarum* ppl.13211. <https://doi.org/10.1111/ppl.13211>
- Ferguson, B. J., Gresshoff, P. M., 2009: Soybean as a model legume. *Grain Legumes* 53(7).
- Florez, M., Carbonell, M. V., Martínez, E., 2007: Exposure of maize seeds to stationary magnetic fields: Effects on germination and early growth. *Environmental and Experimental Botany* 59(1), 68–75. <https://doi.org/10.1016/j.envexpbot.2005.10.006>
- Gharebaghi, A., Haghighi, M. A., Arouiee, H., 2017: Effect of cadmium on seed germination and earlier basil (*Ocimum basilicum* L. and *Ocimum basilicum* var. *purpurescens*) seedling growth. *Trakia Journal of Sciences* 15(1), 1–4. <https://doi.org/10.15547/tjs.2017.01.001>
- Gianazza, E., Wait, R., Sozzi, A., Regondi, S., Saco, D., Labra, M., Agradi, E., 2007: Growth and protein profile changes in *Lepidium sativum* L. plantlets exposed to cadmium. *Environmental and Experimental Botany* 59(2), 179–187. <https://doi.org/10.1016/j.envexpbot.2005.12.005>
- Hasanuzzaman, M., Bhuyan, M. B., Zulfiqar, F., Raza, A., Mohsin, S. M., Mahmud, J. A., Fujita, M., Fotopoulos, V., 2020: Reactive oxygen species and antioxidant defense in plants under abiotic stress: Revisiting the crucial role of a universal defense regulator. *Antioxidants* 9(8), 681. <https://doi.org/10.3390/antiox9080681>
- He, J., Ren, Y., Chen, X., Chen, H., 2014: Protective roles of nitric oxide on seed germination and seedling growth of rice (*Oryza sativa* L.) under cadmium stress. *Ecotoxicology and Environmental Safety* 108, 114–119. <https://doi.org/10.1016/j.ecoenv.2014.05.021>
- Heath, R. L., Packer, L., 1968: Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. *Archives of Biochemistry and Biophysics* 125, 189–198.
- Hozayn, M., AbdEl-Monem, A. A., El-Mahdy, A. A., 2022: Alleviation salinity stress in germination, seedling vigor, growth, physiochemical, yield and nutritional value of Chickpea (*Cicer arietinum* L.) using magnetic technology in sandy soil. *Egyptian Journal of Chemistry* 65(132), 1317–1331. <https://doi.org/10.21608/ejchem.2022.160474.6910>
- Hussain, A., Rizwan, M., Ali, Q., Ali, S., 2019: Seed priming with silicon nanoparticles improved the biomass and yield while reduced the oxidative stress and cadmium concentration in wheat grains. *Environmental Science and Pollution Research* 26, 7579–7588. <https://doi.org/10.1007/s11356-019-04210-5>
- Iqbal, M. Z., Rahmati, K., 1992: Tolerance of *Albizia lebbek* to Cu and Fe application. *Ekológia, ČSFR* 11, 427–430.
- Kataria, S., Anand, A., Raipuria, R. K., Kumar, S., Jain, M., Watts, A., Brestic, M., 2022: Magnetopriming actuates nitric oxide synthesis to regulate phytohormones for improving germination of soybean seeds under salt stress. *Cells* 11(14), 2174. <https://doi.org/10.3390/cells11142174>
- Kataria, S., Baghel, L., Guruprasad, K. N., 2017: Pre-treatment of seeds with static magnetic field improves germination and early growth characteristics under salt stress in maize and soybean. *Biocatalysis and Agricultural Biotechnology* 10, 83–90. <https://doi.org/10.1016/j.bcab.2017.02.010>
- Kataria, S., Baghel, L., Jain, M., Guruprasad, K. N., 2019: Magnetopriming regulates antioxidant defense system in soybean against salt stress. *Biocatalysis and Agricultural Biotechnology* 18, 101090. <https://doi.org/10.1016/j.bcab.2019.101090>
- Kataria, S., Jain, M., Rastogi, A., Brestic, M., 2021: Static magnetic field treatment enhanced photosynthetic performance in soybean under supplemental ultraviolet-B radiation. *Photosynthesis Research* 150, 263–278. <https://doi.org/10.1007/s11120-021-00850-2>
- Kataria, S., Jain, M., Tripathi, D. K., Singh, V.P., 2020: Involvement of nitrate reductase-dependent nitric oxide production in magnetopriming-induced salt tolerance in soybean. *Physiologia Plantarum* 168(2), 422–436. <https://doi.org/10.1111/ppl.13031>
- Kataria, S., Lokesh, B., Guruprasad, K. N., 2015: Acceleration of germination and early growth characteristics of soybean and maize after pre-treatment of seeds with static magnetic field. *International Journal of Tropical Agriculture* 33(2), 985–992.
- Kataria, S., Shukla, S., Jumrani, K., Jain, M., Gadre, R., 2023: Stimulating role of calcium and cyclic GMP in mediating the effect of magnetopriming for alleviation of salt stress in soybean seedlings. *Seeds* 2(2), 232–245. <https://doi.org/10.3390/seeds2020018>
- Kavi, P. S., 1977: The effect of magnetic treatment of soybean seeds and its moisture absorbing capacity. *Science and Culture* 9, 405–406.
- Khizhenkov, P. K., Dobritsa, N. V., Netsvetov, M. V., Driban, V. M., 2001: Influence of low-and super low-frequency alternating magnetic fields on ionic permeability of cell membranes; Vliyanie nizko-i sverkh nizkochastotnykh peremennykh magnitnykh polej na ionnyu pronitsaemost' kletochnykh membran. *Dopovydyi Natsyional'noyi Akademiyi Nauk Ukrainy* (4), 161-164.
- Khosrojerdi, M., Moghaddam, M., Farhadi, N., 2023: Magnetic water irrigation changes physiological traits and stress tolerance of *Salvia virgata* under saline conditions. *Scientia Horticulturae* 314, 111935. <https://doi.org/10.1016/j.scienta.2023.111935>
- Kunitz, M., 1947: A modified method for the determination of aminoacids. *Journal of General Physiology* 30, 291–298.
- Kuriakose, S. V., Prasad, M. N. V., 2008: Cadmium stress affects seed germination and seedling growth in *Sorghum bicolor* (L.) Moench by changing the activities of hydrolyzing enzymes. *Plant Growth Regulation* 54, 143–156. <https://doi.org/10.1007/s10725-007-9237-4>
- Liu, S., Yang, C., Xie, W., Xia, C., Fan, P., 2012: The effects of cadmium on germination and seedling growth of *Suaeda salsa*. *Procedia Environmental Sciences* 16, 293–298. <https://doi.org/10.1016/j.proenv.2012.10.041>
- Mahawar, L., Popek, R., Shekhawat, G. S., Alyemini, M. N., Ahmad, P., 2021: Exogenous hemin improves Cd<sup>2+</sup> tolerance and remediation potential in *Vigna radiata* by intensifying the HO-1 mediated antioxidant defence system. *Scientific Reports* 11, 2811. <https://doi.org/10.1038/s41598-021-82391-1>
- Mahawar, L., Shekhawat, G. S., 2023: Understanding the physiological mechanism of heme oxygenase for enhanced tolerance and phytoremediation of Cd<sup>2+</sup> in *Eruca sativa*: Co-ordinated function of antioxidant defense system. *Journal of Plant Growth Regulation* 42, 6307–6318. <https://doi.org/10.1007/s00344-022-10825-9>
- Mahmoud, T. A., Youssef, E. A., El-Harouny, S. B., Abo Eid, M. A. M., 2019: Effect of irrigation with magnetic water on ni-

- trogen fertilization efficiency of navel orange trees. *Plant Archives* 19(1), 966–975.
- Malar, S., Sahi, S. V., Favas, P. J. C., Venkatachalam, P., 2015: Mercury heavy-metal-induced physiochemical changes and genotoxic alterations in water hyacinths [*Eichhornia crassipes* (Mart.)]. *Environmental Science and Pollution Research* 22, 4597–4608. <https://doi.org/10.1007/s11356-014-3576-2>
- Martinez, E., Carbonell, M. V., Flórez, M., Amaya, J. M., Maqueda, R., 2009: Germination of tomato seeds [*Lycopersicon esculentum* L.] under magnetic field. *International Agrophysics* 23, 45–49.
- Mukherjee, S. P., Choudhuri, M. A., 1983: Implications of water stress-induced changes in the levels of endogenous ascorbic acid and hydrogen peroxide in *Vigna* seedlings. *Physiologia Plantarum* 58(2), 166–170. <https://doi.org/10.1111/j.1399-3054.1983.tb04162.x>
- Okba, S. K., Mazrou, Y., Mikhael, G. B., Farag, M. E., Alam-Eldein, S. M., 2022: Magnetized water and proline to boost the growth, productivity and fruit quality of ‘Taifi’ pomegranate subjected to deficit irrigation in saline clay soils of semi-arid Egypt. *Horticulturae* 8(7), 564. <https://doi.org/10.3390/horticulturae8070564>
- Prajapati, R., Kataria, S., Gadre, R., Landi, M., Jain, M., 2023: Unveiling the mechanisms underpinning alleviation of mercury toxicity by static magnetic field treatment in soybean. *Journal of Plant Growth Regulation*. <https://doi.org/10.1007/s00344-023-11063-3>
- Prajapati, R., Kataria, S., Jain, M., 2020: Seed priming for alleviation of heavy metal toxicity in plants: An overview. *Plant Science Today* 7(3), 308–313. <https://doi.org/10.14719/pst.2020.7.3.751>
- Radhakrishnan, R., Kumari, B. D. R., 2012: Pulsed magnetic field: A contemporary approach offers to enhance plant growth and yield of soybean. *Plant Physiology and Biochemistry* 51, 139–144. <https://doi.org/10.1016/j.plaphy.2011.10.017>
- Raipuria, R. K., Kataria, S., Watts, A., Jain, M., 2021: Magnetopriming promotes nitric oxide via nitric oxide synthase to ameliorate the UV-B stress during germination of soybean seedlings. *Journal of Photochemistry and Photobiology B: Biology* 220, 112211. <https://doi.org/10.1016/j.jphotobiol.2021.112211>
- Rajendra, P., Sujatha Nayak, H., Sashidhar, R. B., Subramanyam, C., Devendranath, D., Gunasekaran, B., Aradhya, R. S., Bhasakaran, A., 2005: Effects of power frequency electromagnetic fields on growth of Germinating *Vicia faba* L., the broad bean. *Electromagnetic Biology and Medicine* 24(1), 39–54. <https://doi.org/10.1081/JBC-200055058>
- Reina F. G., Pascual L. A., Fundora I. A., 2001: Influence of a stationary magnetic field on water relations in lettuce seeds. Part II: experimental results. *Bioelectromagnetics* 22(8), 596–602. [10.1002/bem.89](https://doi.org/10.1002/bem.89). PMID: 11748678.
- Safari, F., Akramian, M., Salehi-Arjmand, H., Khadivi, A., 2019: Physiological and molecular mechanisms underlying salicylic acid-mitigated mercury toxicity in lemon balm (*Melissa officinalis* L.). *Ecotoxicology and Environmental Safety* 183, 109542. <https://doi.org/10.1016/j.ecoenv.2019.109542>.
- Saharan, B. S., Brar, B., Duhan, J. S., Kumar, R., Marwaha, S., Rajput, V. D., Minkina, T., 2022: Molecular and physiological mechanisms to mitigate abiotic stress conditions in plants. *Life* 12(19), 1634. <https://doi.org/10.3390/life12101634>
- Sarraf, M., Kataria, S., Taimourya, H., Santos, L. O., Menegatti, R. D., Jain, M., Ihtisham, M., Liu, S., 2020: Magnetic field (MF) applications in plants: An overview. *Plants* 9, 1139.
- Sawhney, S., Toky, K. L., Nanda, K. K., 1970: Changes in amylase activity during extension growth and floral induction in *Impatiens balsamina*, a qualitative short day plant. *Indian Journal of Plant Physiology* 13, 198–205.
- Sharmila, P., Kumari, P. K., Singh, K., Prasad, N. V. S. R. K., Pardha-Saradhi, P., 2017: Cadmium toxicity-induced proline accumulation is coupled to iron depletion. *Protoplasma* 254, 763–770. <https://doi.org/10.1007/s00709-016-0988-5>
- Sheirdil, R. A., Bashir, K., Hayat, R., Akhtar, M. S., 2012: Effect of cadmium on soybean (*Glycine max* L) growth and nitrogen fixation. *African Journal of Biotechnology* 11(8), 1886–1891. <https://doi.org/10.5897/AJB11.2849>
- Shine, M. B., Guruprasad, K. N., Anand, A., 2012: Effect of stationary magnetic field strengths of 150 and 200 mT on reactive oxygen species production in soybean. *Bioelectromagnetics* 33(5), 428–437. <https://doi.org/10.1002/bem.21702>
- Singhal, R. K., Kumar, M., Bose, B., Mondal, S., Srivastava, S., Dhankher, O. P., Tripathi, R. D., 2023: Heavy metal (loid)s phytotoxicity in crops and its mitigation through seed priming technology. *International Journal of Phytoremediation* 25(2), 187–206. <https://doi.org/10.1080/15226514.2022.2068502>
- Stefanello, R., 2019: Toxicity of cadmium on the germination of thyme seeds. *Caderno de Pesquisa, Série Biologia* 31(2), 9–21. <https://doi.org/10.17058/cp.v31i2.13091>
- Stutte, G. W., Monje, O., Hatfield, R. D., Paul, A.-L., Ferl, R. J., Simone, C. G., 2006: Microgravity effects on leaf morphology, cell structure, carbon metabolism and mRNA expression of dwarf wheat. *Planta* 224, 1038–1049. <https://doi.org/10.1007/s00425-006-0290-4>
- Thomas, S., Anand, A., Chinnusamy, V., Dahuja, A., Basu, S., 2013: Magnetopriming circumvents the effect of salinity stress on germination in chickpea seeds. *Acta Physiologia Plantarum* 35, 3401–3411. <https://doi.org/10.1007/s11738-013-1375-x>
- Vashisth, A., Nagarajan, S., 2010: Effect on germination and early growth characteristics in sunflower (*Helianthus annuus*) seeds exposed to static magnetic field. *Journal of Plant Physiology* 167(2), 149–156. <https://doi.org/10.1016/j.jplph.2009.08.011>
- Xue, Z., Gao, H., Zhao, S., 2014: Effects of cadmium on the photosynthetic activity in mature and young leaves of soybean plants. *Environmental Science and Pollution Research* 21, 4656–4664. <https://doi.org/10.1007/s11356-013-2433-z>
- Yang, Y.-H., Gu, C.-S., Chen, F.-D., Shao, Y.-F., Wu, K.-W., Fang, W.-M., Liu, Z.-L., Ding, Y.-S., 2015: Physiological responses and bioaccumulation in *Nymphaea tetragona* Georgi under cadmium exposure. *Fresenius Environmental Bulletin* 24, 1716–1724.
- Yin, X., Wei, R., Chen, H., Zhu, C., Liu, Y., Wen, H., Guo, Q. and Ma, J., 2021: Cadmium isotope constraints on heavy metal sources in a riverine system impacted by multiple anthropogenic activities. *Science of the Total Environment* 750, 141233. <https://doi.org/10.1016/j.scitotenv.2020.141233>.

Short communication

## *Isoëtes gymnocarpa* and *Utricularia* × *neglecta* – new taxa for Montenegro

Roman E. Romanov<sup>1\*</sup>, Snežana Dragičević<sup>2</sup>, Angelo Troia<sup>3</sup>

<sup>1</sup> Komarov Botanical Institute of the Russian Academy of Sciences, Professora Popova 2, 197022 St. Petersburg, Russia

<sup>2</sup> Montenegrin Academy of Sciences and Arts, Rista Stijovića 5, 81000 Podgorica, Montenegro

<sup>3</sup> Dipartimento STEBICEF, Università degli Studi di Palermo, via Archirafi 38, 90123 Palermo, Italy

**Abstract** – One lycophyte genus and species (*Isoëtes gymnocarpa*), and one aquatic magnoliophyte species (*Utricularia* × *neglecta*) new for the flora of Montenegro are reported. It is suggested that the new population of *Isoëtes gymnocarpa* and its habitat should be protected.

**Keywords:** aquatic magnoliophytes, the Balkans, flora, lycophytes, Montenegro, new records

### Introduction

The flora of Montenegro (SE Europe) is easily recognized as one of the richest in Europe by the ratio of the number of species per square km (Pulević 2022). This is a reflection of evident habitat heterogeneity and richness resulting in large amounts of different ecological niches available for plants. A long tradition of botanical research has resulted in the more than 3,600 species and subspecies of vascular plants known from the Montenegrin flora (Rohlena 1942, Pulević 2005, 2022, Stešević et al. 2008, Stešević & Caković 2013, 2021). However, some habitat types, e.g., small temporary and permanent water bodies and wetlands in coastal region, have received less attention (cf. Bubanja 2016, Bubanja et al. 2016). A lot of them can be referred to “Mediterranean temporary ponds”, a priority habitat according to the European Union 92/43 Habitats Directive, showing a clearly regressive trend. Distributional patterns of plants associated with these habitats in the Balkan Peninsula are still poorly known.

### Materials and methods

Some localities in the coastal region and Zeta plain of Montenegro were surveyed during 2022 and 2023 as potential habitats for charophytes (Characeae), focussing on species of plants associated with them. All types of water bodies and wetlands encountered were checked, including Mediterranean temporary ponds, inundated sand pits,

quarries, and artificial depressions, excavated ponds, drainage channels, puddles, concrete ponds, temporary and permanent streams and springs and the water bodies associated with them. The strong seasonality in the traits of water bodies and the appearance of aquatic plants as well as habitat heterogeneity are remarkable for the area surveyed. The specimens were collected by hand and studied in a living state. Voucher specimens were deposited in the Herbarium of the Natural History Museum of Montenegro (NHMM) and the Herbarium Mediterraneum Panormitanum (PAL). The nomenclature for lycophytes follows Troia & Greuter (2014), for *Utricularia*, Bobrov et al. (2022).

### Results and discussion

Fifteen species of charophytes, belonging to the genera *Chara* L., *Lamprothamnium* J.Groves, *Nitella* C.Agardh, *Tolypella* (A.Braun) A.Braun, and *Sphaerochara* Mädler, were found in the research area. Our efforts aimed at the listing of all associated vascular plant species resulted in the finding of one genus and two species, new for the flora of Montenegro.

*Isoëtes gymnocarpa* (Gennari) A.Braun (Fig. 1A): North of the Nature Park Ulcinjska Solana (US), unshaded drainage channel within farmland moderately damaged by livestock and probably briefly inundated during winter and early spring, 41.92966 N, 19.27361 E, 3 m a.s.l., 17 IV 2023,

\* Corresponding author e-mail: streptophytes@gmail.com

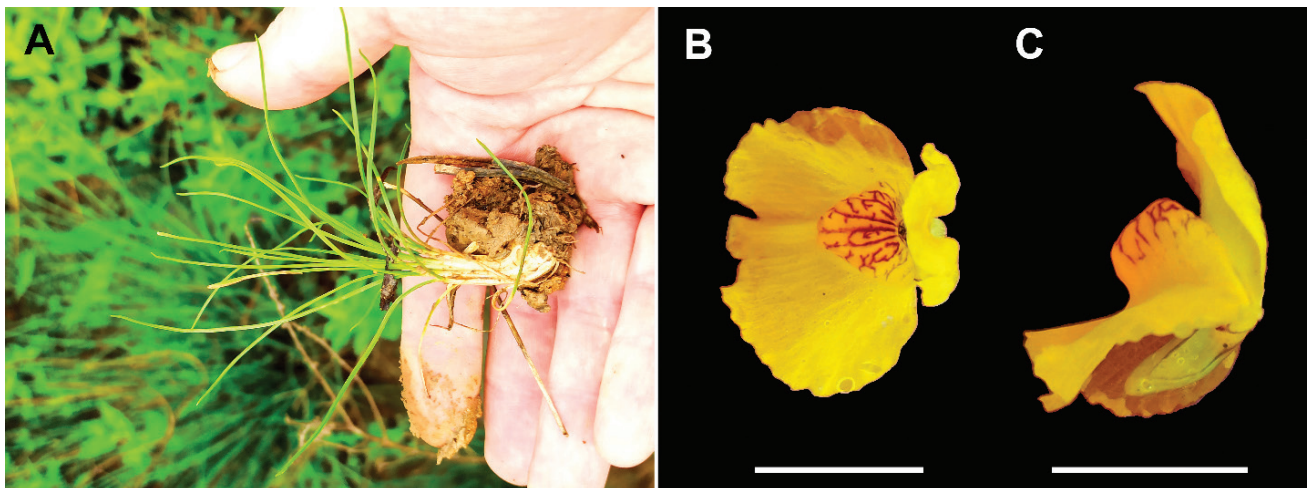


Fig. 1. New genus and species records for Montenegro: A – *Isoëtes gymnocarpa*, general appearance of unearthened plant, B, C – *Utricularia* × *neglecta*, B – isolated corolla showing large flat lower lip, view from above, C – isolated corolla showing short straight spur and nearly right angle between upper and lower lips, lateral view. Scale: B, C – 1 cm. All photos were taken from the voucher specimens by R. E. Romanov.

leg: Roman Romanov (RR) and Snežana Dragičević, det: Angelo Troia (NHMM, PAL).

This habitat is characterized by the specific water regime, which implies an alternation of wet (winter, spring) and dry phases (summer and autumn), just like that in the meadows in the immediate vicinity with mosaic habitats and communities of wet grasslands and pastures which host Mediterranean therophytes and geophytes. These agricultural lands are pastures grazed by livestock (sheep, cows) or partly arable land, with other negative impacts in the immediate environment (like cutting of surrounding trees and shrubs and the associated physical disturbance). The cover abundance of *Isoëtes* was low, not exceeding 20%. The plants were spotted within an area of less than 10 square meters.

The locality is currently outside the Nature Park Ulcinjska Salina, but because of the presence of this rare species of *Isoëtes*, it should be protected by the Park area being extended to include it.

*Isoëtes gymnocarpa* is close to *I. histrix* Bory (for its terrestrial habitat, and for the presence of phyllopodia and tuberculate megaspores), the main diagnostic character being the length of phyllopodia lateral teeth (not longer and thinner than the central tooth as in *I. histrix*, but as long and as thick as the central tooth, cf. Troia & Greuter 2014, 2015a, 2015b). The group needs further investigation, but we follow here the taxonomic view of Troia & Greuter (2015a) with *I. gymnocarpa* = *I. sicula* Tod. = *I. subinermis* (Gennari) Cesca & Peruzzi. The taxon so circumscribed is spread around the Mediterranean area, from the Iberian Peninsula to Anatolia: the population here reported is the northernmost along the Adriatic coasts, both in the Balkan Peninsula and in Italy. A previous report of *I. histrix* for southern Albania (Barina et al. 2013) should be checked to assess if it belongs to *I. histrix* or to *I. gymnocarpa*.

*Isoëtes gymnocarpa* should be recognized as indigenous to Montenegro because no cases of non-native species are

known for this genus. The inconspicuous general habit of terrestrial *Isoëtes*, having the appearance of sterile monocots, can be a possible explanation why species of this genus were not spotted in Montenegro before.

*Utricularia* × *neglecta* Lehm. (Fig. 1B, C): Morača River, the lower reaches, the island of Vranjina, the wetland of Bakine Tigle (BT), 19 VI 2023, 42.28217 N, 19.15145 E, 4 m a.s.l., 19 VI 2023, leg: RR, det: RR (NHMM); BT, small floodplain pool, 42.28746 N, 19.15261 E, 5 m a.s.l., 19 VI 2023, leg: RR, det: RR (NHMM). The cover abundance was low, not exceeding 20 % in both cases.

This hybrid is widely distributed in temperate and tropical regions of the Old World (Taylor 1989, Uotila 2013, Bobrov et al. 2022, <http://www.plantsoftheworldonline.org/>). For a long time, it was reported as *U. australis* R.Br. It was mentioned from Montenegro, from Lake Skadar, based on images of sterile plants, not showing the leaf-teeth bristle character (<https://www.inaturalist.org/observations/31338617>), a trait only visible with microscopy. Moreover, the use of solely this character can be unreliable (Taylor 1989). Therefore, our records are either the first verified ones for Montenegro, or else can be recognized as a confirmation of an earlier uncheckable record. The infrequent and short period of flowering, usually associated with habitats difficult of access, hampers reliable recording of *Utricularia* species. This hybrid seems to be indigenous to Montenegro; no cases of introduction are known for it in the Balkan Peninsula.

## Acknowledgments

The anonymous reviewers are gratefully acknowledged for their encouragement and kind suggestions for the improvement of the manuscript.

This research is supported by the project “BioMon – Establishing biodiversity monitoring in Ulcinj Salina”,

supported by the IKI Small Grants programme, which is part of the International Climate Initiative (IKI). IKI Small Grants is funded by the Federal Ministry of Economic Affairs and Climate Action (BMWK) and the Federal Foreign Office (AA), Germany, and carried out by Deutsche Gesellschaft für Internationale Zusammenarbeit (GIZ) GmbH. The BioMon project is implemented by the Center for Protection and Research of Birds (CZIP), a BirdLife partner in Montenegro. This research is partly supported by the project of the Komarov Botanical Institute RAS (No. 121021600184-6).

### Author contribution statement

Roman E. Romanov – field studies, preparation and identification of specimens, manuscript writing; Snežana Dragićević – field studies, preparation of specimens, manuscript writing; Angelo Troia – preparation and identification of specimens, manuscript writing.

### References

- Barina, Z., Rakaj, M., Pifkó, D., 2013: Contributions to the flora of Albania, 4. Willdenowia 43(1), 165–184. <http://dx.doi.org/10.3372/wi.43.43119>
- Bobrov, A. A., Volkova, P. A., Kopylov-Guskov, Y. O., Mochalova, O. A., Kravchuk, A. E., Nekrasova, D. M., 2022: Unknown sides of *Utricularia* (Lentibulariaceae) diversity in East Europe and North Asia or how hybridization explained old taxonomical puzzles. *Perspectives in Plant Ecology, Evolution and Systematics* 54, 125649. <https://doi.org/10.1016/j.ppees.2021.125649>
- Bubanja, N., 2016: Geobotanical characterization of vascular flora of long Ulcinj beach and its hinterland in Montenegro. Doctoral dissertation. University of Belgrade, Faculty of biology, Belgrade.
- Bubanja, N., Šinžar-Sekulić, J., Stevanović, V., 2016: Assessing the influence of environmental parameters on aquatic plants of ponds in the hinterland of Long Beach in Montenegro. *Limnetica* 35(2), 385–396. <https://doi.org/10.23818/limn.35.31>
- Pulević, V., 2005: Material for the vascular flora of Montenegro. Posebno izdanje Republičkog zavoda za zaštitu prirode Crne Gore, Podgorica.
- Pulević, V. (ed.), 2022: Botanical lexicon of Montenegro. Montenegrin Academy of Sciences and Arts, Lexicographic center, Lexicons, Book 3, Podgorica.
- Rohlena, J., 1942: *Conspectus florae Montenegrinae* [An overview of the Montenegrin flora]. *Preslia* 20–21, 1–506.
- Stešević, D., Caković, D., 2013: *Katalog vaskularne flore Crne Gore* [The catalog of the vascular flora of Montenegro]. T. 1. Podgorica.
- Stešević, D., Caković, D., 2021: *Katalog vaskularne flore Crne Gore* [The catalog of the vascular flora of Montenegro]. T. 2. Podgorica.
- Stešević, D., Petrović, D., Bubanja, N., Vuksanović, S., Biberdžić, V., 2008: Contribution to the flora of Montenegro (Supplementum to the material for vascular flora of Montenegro). *Natura Montenegrina*, Podgorica 7(3), 463–480.
- Taylor, P., 1989: *The genus Utricularia: a taxonomic monograph*. London.
- Troia, A., Greuter, W., 2014: A critical conspectus of Italian *Isoetes* (Isoetaceae). *Plant Biosystems* 148(1), 13–20. <https://doi.org/10.1080/11263504.2013.878409>
- Troia, A., Greuter, W., 2015a: *Isoetaceae* (vers. 1.0). In: Peruzzi, L., Cecchi, L., Cristofolini, G., Domina, G., Greuter, W., Nardi, E., Raimondo, F. M., Selvi, F., Troia, A. (eds.), *Flora critica d'Italia*. Fondazione per la Flora Italiana, Firenze. Published online on 25 February 2015. Available at: <https://floraitaliana.it/contenuti/flora-critica/6>
- Troia, A., Greuter W., 2015b: A conspectus of and key to Greek *Isoetes* (Isoetaceae), based on a reassessment of Hausknecht's gatherings of 1885. *Willdenowia* 45(3), 391–403. <https://doi.org/10.3372/wi.45.45303>
- Uotila, P., 2013: *Lentibulariaceae*. Euro+Med Plantbase – the information resource for Euro-Mediterranean plant diversity.



# Chlorophyll *a* fluorescence measurements in Croatia • the first twenty years

Edited by Hrvoje Lepeduš, Marija Viljevac Vuletić, Zvonimir Zdunić. 2023, 189 pp, ISBN 978-953-7843-11-3 (Print), ISBN 978-953-7843-12-0 (eBook)

Publisher: Agricultural Institute Osijek

Chlorophyll fluorescence (ChlF) techniques are sensitive, rapid and non-invasive and provide a wide range of data on plant health. ChlF has been used for more than two decades to study photosynthesis in plants, algae and bacteria. In Croatia, the pioneering work in ChlF research began in the Department of Biology of the Josip Juraj Strossmayer University of Osijek. Since then, many scientific and agronomic laboratories and institutes in Croatia have used this technique in regular research and applied it in practise. This method has become the basis for many collaborative ventures among scientists in Croatia and abroad. Thanks to this cooperation, many young researchers were given the opportunity to learn the ChlF method and apply it in their research. This led to 14 doctoral theses, one master's thesis and numerous bachelor's theses. This monograph, published in English, is thus a tribute to twenty years of the application of ChlF methods in biology and agronomy in the Republic of Croatia.

The monograph begins with a historical overview of the use of ChlF in scientific research and continues with the application of the ChlF method in fundamental research in plant biology and agronomy. In the era of climate change, photosynthesis has proven to be a reliable marker for the degree of adaptation and tolerance of plants. The four chapters present examples of the application of the ChlF method in research and in the understanding of the responses of plants to various abiotic stress factors. The next six chapters present the practical application of the ChlF method in agronomic research, especially in elucidating the stress responses of plants under adverse environmental conditions and in screening for tolerant genotypes, which are of great importance in breeding programs for various crops. The penultimate chapter presents a case study and gives a detailed insight into the scope and effectiveness of the ChlF



method in phenotyping plant material. The last chapter deals with the recent technological advances in the ChlF method and its future perspectives. As Dr. Šimić and Assoc. Prof. Dr. Mlinarić state in the final chapter of this monograph, "...chlorophyll fluorescence remains an important tool for understanding photosynthesis and its response to environmental stress, the health of plants and ecosystems. Continued advancements in technology and analysis will further improve its usefulness. Its future prospects are

bright, with many potential applications in agriculture, urban farming, forestry, aquatic research, climate change, and bioenergy research.”

The monograph was presented by the editors Dr. Marija Viljevac Vuletić, senior research scientist from the Agri-

cultural Institute Osijek, and Prof. Dr. Hrvoje Lepeduš from the Faculty of Humanities and Social Sciences, Josip Juraj Strossmayer University of Osijek, at a meeting of the Croatian Society of Plant Biologists in February 2024 in Zagreb (Fig. 1).



Fig. 1. Presentation of the monograph *CHLOROPHYLL a FLUORESCENCE MEASUREMENTS IN CROATIA • FIRST TWENTY YEARS*, ed. Dr. Marija Viljevac Vuletić and Prof. Dr. Hrvoje Lepeduš (A) at a meeting of the Croatian Society of Plant Biologists (B).

Dr. Branka Salopek Sondi  
Ruđer Bošković Institute, Zagreb



## INSTRUCTIONS FOR AUTHORS

The interest of the journal is field (terrestrial and aquatic) and experimental botany including plant viruses, bacteria, archaea, algae and fungi, from subcellular level to the ecosystem level with a geographic focus on karstic areas of the southern Europe and the Adriatic Sea (Mediterranean).

The journal welcomes manuscripts for publication in the following categories: original research papers, short communications, book reviews, social news and announcements. Review articles are accepted on editor invitation only.

### Article submission and publishing are free of charge.

Manuscripts should be submitted using On-line Manuscript Submission at <http://www.abc.botanic.hr>. Registration and login are required to submit items on-line and to check the status of current submissions. For submission, after LOGIN find USER HOME then AUTHOR and go to NEW SUBMISSION.

Under SUBMISSION METADATA, fill in the names and e-mail addresses of **all authors**. Criteria for authorship are as set out by the ICMJE and as recommended by the Committee on Publication Ethics (COPE).

### Cover letter

In the cover letter addressed to Editor-in-Chief, the authors should explain how the manuscript meets the scope of the journal and indicate why it will be of interest to the general readership of *Acta Botanica Croatica*. Authors should propose the names and e-mail addresses of at least **five potential reviewers who are experts in the topic of manuscript. Please avoid colleagues with joint publications, or from the same institutions. At least three of them have to be international recognized scientists outside of your home country.** Also, in the cover letter, confirm that the manuscript has not been published or submitted for publication elsewhere and that all authors have read the manuscript and approved it for submission. Include also Founding statement in which any sources of financial support should be specified.

### Type of contribution

#### ORIGINAL RESEARCH PAPER

An original research paper is a fully documented report of original research. The manuscript should be divided into Introduction, Material and methods, Results, Discussion, Acknowledgements, References (**maximum 40**), Table and figure captions, Tables, Figures. There may be up to **12 single-spaced typewritten pages**, excluding figures and tables. There may be up to **8 tables and/or figures in total** per manuscript. Additional figures and/or tables can be published online only as supplementary materials. All tables and figures should be cited in the text properly (Fig. 1, 2, ..., On-line Suppl. Tab. or Fig.). Exceptionally, in papers dealing with conceptual and theoretical bases, especially botanical and phytosociological nomenclature, Results and Discussion can be combined into one section.

### SHORT COMMUNICATIONS

Short communications should consist of not more than **3 single-spaced typewritten pages** and a **maximum of two tables and/or figures**. The text should be divided into Abstract (containing no more than 100 words), Keywords (up to five; listed in alphabetical order), Introduction, Material and methods, Results and discussion, Acknowledgements, References (**maximum 10**), Table and figure captions, Tables and/or Figures.

We may consider longer short communication and research articles with a compelling rationale in the cover letter, contingent upon the content's justification for the extended length.

### REVIEW ARTICLE

Review and mini-review articles are usually accepted for the reviewing process if invited by editor. Authors who wish to contribute a manuscript to this category are encouraged to contact the Editor-in-Chief. The manuscript should be organized according to *Acta Botanica Croatica* guidelines and there are no limitations on the number of references.

### SOCIAL NEWS AND BOOK REVIEWS

We also welcome popular news describing interesting events, anniversaries, as well as short and concise reviews of newly published books in the field of plant sciences.

## PREPARATION OF MANUSCRIPT

### General

The manuscript should be submitted as a Word document. The writing needs to be clear, concise and in correct English. Unfortunately, we do not offer a language editing service as part of the submission process, so it is up to authors to ensure the highest quality of writing in their manuscript. If the language is deemed too inadequate for easy understanding, the manuscript will be returned to authors without review.

The text should be single-spaced and left-adjusted, using Times New Roman and 12 point letter size. The layout of the document should be A4 (21 × 29.5 cm). Adjust indentation to 1 cm (i.e., the first line of all paragraphs and hanging paragraphs of References). Input your text continuously, i.e. insert hard returns exclusively at the ends of paragraphs, headings etc. Do not use the space bar to create indents; the indent command should be used for this purpose. Leave a space between mathematical symbols and numbers (e.g. 2 + 3, 3 < 9). Always leave a space between a number and a Celsius degree symbol (e.g. 12 °C). Do not leave a space when using the multiplication and percentage symbols (e.g. 6×12%). Each page should be numbered.

The metric system should be used throughout the manuscript. If required, equivalent values in other systems may be placed in parentheses immediately after the metric value.

Italicize only the names of genes (e.g. *Arpl* gene), genera, species, subspecies and lower taxonomic units. Genetic information, such as DNA, RNA, or protein sequences, should

be submitted to public data bases (GenBank, EMBL, etc.), and accession numbers should be available in Material and methods. Voucher specimens must be made and deposited in a public herbarium. For endemic and protected taxa, permission has to be obtained from the competent authority. The nomenclature of taxa and syntaxa has to be in strict accordance with international rules (codes).

### Title page

TITLE, should not exceed 120 characters (without spaces).

NAMES OF ALL AUTHORS (name and surname in full), their mailing and e-mail addresses, and institutional affiliations should be given. Include the corresponding author's e-mail address and telephone number.

RUNNING TITLE, should not exceed 50 characters (without spaces).

ABSTRACT of up to 250 words that highlights the objective, results, and conclusion of the paper.

KEY WORDS (up to eight, in alphabetical order), to identify the subjects under which the article may be indexed.

### Content of manuscript

Keep the Introduction brief, stating clearly the purpose of the article and its relation to other papers on the same subject. Do not give an extensive review of literature. Provide enough information in the Material and methods section to enable other investigators to repeat the experiments. Report Results clearly. In the Discussion interpret the results, state their meaning and draw conclusions. Do not simply repeat the results. Proceed with Acknowledgments where any sources of financial support as well as any individuals who were of direct help to the authors should be acknowledged. At the end give Author contribution statement in which the contributions of all authors should be described.

### References

Cite references in the text by name and year in parentheses. Some examples: Wrisher (1998), ... Jones and Smith (1987), ... (Jones 1987a, b), ..... Jones et al. (1986), ... (Facca et al. 2002, 2003, Socal et al. 2006).

Arrange names of authors chronologically in text, e.g: (Jones 1986, Allen 1987). The list of references should be typed in alphabetical order. The articles in English, Spanish, French, German and Italian are accepted in the Reference list. For any other language, please provide the English translation in parentheses. Unpublished materials should be cited in the text as personal observations or unpublished data. Identify authors of unpublished work. Check the text citations against the Reference list to make sure there are no gaps or inconsistencies. Names of journals should be given in full, followed by volume number and pages. To mark a span of pages use en dash (–) instead of a hyphen(-). Please provide doi numbers wherever is possible.

Use the following formats for Reference list style:

Journal article:

Colangelo, E. P., Guerinot, M. L., 2006: Put the metal to the petal: metal uptake and transport throughout plants.

Current Opinion in Plant Biology 9, 322–330. <https://doi.org/10.1016/j.pbi.2006.03.015>.

Books:

Horvat, I., Glavaš, V., Ellenberg, H., 1974: Vegetation Sudosteuropas. Geobotanica selecta 4. Gustav Fischer Verlag, Stuttgart.

Chapter in a book:

Broadwater, S. T., Scott, J., 1994: Ultrastructure of unicellular red algae. In: Seckbach, J. (ed.), Evolutionary pathways and enigmatic algae: *Cyanidium caldarium* (Rhodophyta) and related cells, 215–230. Kluwer Academic Publishers, Amsterdam.

On-line sources: Author, year, title, source. Retrieved October 15, 2015 from <http://www...>

### Table and figures

Table and figure legends should be added following references, on the next page.

**Tables** should be on separate pages. Tables should be prepared in Microsoft Office Word or Excel. Vertical lines should not be visible in tables. The maximum width of a printed table should be 150 characters in broadside. The preferred table organization format can be seen in articles published in previous issues of Acta Botanica Croatica, which are freely available on-line. Large tables and primary data can be published as supplementary materials on-line, but not in the printed version. All tables should be numbered consecutively with Arabic numerals. They should be cited in the text properly (Tab. 1, On-line Suppl. Tab. 1, etc.). Table title should be above the table, on the same page as the table to which it corresponds. The tables need to be self-explanatory: the authors should provide enough information in captions (explain all abbreviations, write full Latin names etc.) so that each table is understandable without reference to the text.

**Figures** should be submitted in appropriate electronic formats as Supplementary files as well as embedded within the manuscript after the tables. Every figure must be referenced in the text. Figures should be numbered in Arabic numerals (below the figure). Figures may be arranged in panels, in which individual images should be divided by white lines no more than 2 mm wide. Line art-works and half-tones or photographic images should be saved as Tagged Image Format (tif) with a resolution of at least 600 dpi or in pdf. The size of tif files can be decreased using Lossless Compression (LZW). Vector graphics (xls, cmx, eps, wmf) should be saved in pdf. All lettering on figures should be in Arial and legible after reduction. Y- and X-axis need to be black, tick marks on axes should be oriented inwards. Graph lines should be thicker than axes lines. Each figure and figure caption should contain all the information necessary for it to be self-explanatory (explain all abbreviations, write full Latin names etc.) so that each figure is understandable without reference to the text. Colour figures are acceptable only if necessary (photographs, not plots and curves) and they are free of charge.

### **Additional guidelines**

All on-line supplementary materials have to be uploaded as a separate Word document and supplementary figures should be submitted in appropriate electronic formats (tif or pdf) as separate files.

Appendices (optional): Each appendix must be numbered as Appendices 1., 2. etc and must have a title.

Footnotes should not be used; information should be integrated into the text.

### **REVIEW PROCESS**

Acta Botanica Croatica is committed to peer-review integrity and upholding the highest standards of review. Once your paper has been assessed for suitability by the Editor-in-Chief and Section Editor, it will then be single blind peer reviewed by independent, anonymous expert referees.

Manuscripts that meet the scientific and journal technical criteria will be sent to the review process. Please note that the journal uses software to screen for plagiarism. Acta Botanica Croatica participates in an initiative by CrossRef (<http://www.crossref.org>) to prevent scholarly and professional plagiarism in scientific publications. This initiative is known as Crossref Similarity Check and provides its members a service to screen received content for originality against a vast database of relevant published material.

### **AHEAD OF PRINT**

The accepted article, including supplementary files, that has been language edited and checked for references, will be published on-line as "Ahead of print" at [https://](https://hrcak.srce.hr/acta-botanica-croatica)

[hrcak.srce.hr/acta-botanica-croatica](https://hrcak.srce.hr/acta-botanica-croatica) and citable with the DOI number.

### **PROOFS**

The proof is sent to the corresponding author for a final check and approval. Corrected proofs must be returned within 72 hours to the Technical Editor-in-Chief. PDF of corrected proofs will be posted on-line and after release of the printed version (1<sup>st</sup> April, 1<sup>st</sup> October), the paper can also be cited by issue and page numbers.

Free unlimited electronic reprints (in pdf.) are available from <http://hrcak.srce.hr/acta-botanica-croatica>.

A printed copy of the journal volume is available for subscribed readers who have paid the annual fee.

### **COPYRIGHT**

Acta Botanica Croatica is an Open Access journal with minimal restrictions regarding content reuse. Immediately after publishing, all content becomes freely available to anyone for unlimited use and distribution, under the sole condition that the author(s) and the original source are properly attributed according to the Creative Commons Attribution 4.0 International License (CC BY 4.0).

CC BY 4.0 represents the highest level of Open Access, which maximizes dissemination of scholarly work and protects the rights of its authors. In Acta Botanica Croatica, authors hold the copyright of their work and retain unrestricted publishing rights.

By approving final Proof the authors grant to the publisher exclusive license to publish their article in print and on-line, in accordance with the Creative Commons Attribution (CC BY 4.0) license.

Indexed in AGRICOLA (National Agricultural Library), CABI – CAB Abstracts, CNKI Scholar (China National Knowledge Infrastructure), DOAJ, EBSCO Discovery Service, Elsevier (BIOBASE/ CABS, SCOPUS), ProQuest (Aquatic Science & Fisheries Abstracts, Microbiology Abstracts A, B, C, Biotechnology and Bioengineering Abstracts, Environmental Science and Pollution Management, Genetics Abstracts, Virology and AIDS Abstracts), Thomson Reuters (BIOSIS Previews, Science Citation Index Expanded, Zoological Record).

---

**ACTA BOTANICA CROATICA**

E-mail: [acta@biol.pmf.hr](mailto:acta@biol.pmf.hr)

## **ORDER FORM:**

Please enter my annual subscription at 50 EUR.

Name (Institution):

Address:

OIB/VAT identification number:

E-mail:

Payments: This order should be sent to the Editorial Office, e-mail: [acta@biol.pmf.hr](mailto:acta@biol.pmf.hr). Payment should be made after receipt of the invoice.

Date:

Signature:

## CONTENTS

### Full Articles

- Massimo Terzi, Nenad Jasprica, Andraž Čarni, Vlado Matevski, Erwin Bergmeier, Jean-Paul Theurillat:  
Nomenclature of the Balkan alliance *Romuleion graecae* (*Poetea bulbosae*) ..... 87
- Sevilay Öztürk, Oğuz Kurt: The taxonomy and distribution of algae in the thermal springs of Türkiye .... 92
- Burcu Yılmaz Çitak: Does palynotaxonomy contribute to the systematics of the genus? The section  
*Multicaulia* of the genus *Hedysarum* (Fabaceae) example in Türkiye ..... 100
- Wafaa Kamal Taia, Rim Samir Hamdy, Amany Mohamed Abd El-Maged: Morpho-palynological assessment  
of the genus *Terminalia* L. (Combretaceae) in Egypt ..... 107
- Duilio Iamónico, Gianluca Nicoletta: First record of the woody *Melaleuca williamsii* s.l. (Myrtaceae) out of  
its native range ..... 115
- Massimo Terzi, Nenad Jasprica: Changes in grassland vegetation on the island of Plavnik (Croatia) over  
100 years ..... 119
- Sabrina Gueridi, Lilya Boucelha, Ouzna Abrous-Belbachir, Réda Djebbar: Effects of hormoprime and  
pretreatment with gibberellic acid on fenugreek (*Trigonella foenum-graecum* L.) seed germination .... 135
- Aslihan Temel, Taylan Kosesakal: Effects of acetic acid treatment on growth and pigment contents in  
barley ..... 145
- Ramazan Karamzahi, Alireza Einali: Trehalose-induced metabolic responses in basil (*Ocimum basilicum*)  
seedlings under salt treatment ..... 153
- Anjali Vyas, Sunita Kataria, Rajkumar Prajapati, Meeta Jain: Mitigation of cadmium toxicity stress  
by magnetoprime during germination of soybean ..... 165

### Short Communication

- Roman E. Romanov, Snežana Dragičević, Angelo Troia: *Isoëtes gymnocarpa* and *Utricularia* × *neglecta* – new  
taxa for Montenegro ..... 176

### Book Review

- Branka Salopek Sondi: Hrvoje Lepeduš, Marija Viljevac Vuletić, Zvonimir Zdunić (eds.): Chlorophyll *a*  
fluorescence measurements in Croatia – the first twenty years (2023) ..... 179